



# Oral Candidiasis in Hematological Malignancy Patients: Identification and Antifungal Susceptibility Patterns of Isolates

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Received 2020 April 04; Revised 2020 September 14; Accepted 2020 September 14.

## Abstract

**Background:** Oropharyngeal candidiasis is a fungal infection in the mouth caused by *Candida* species. Oropharyngeal candidiasis is a major problem among hematologic malignancy patients.

**Objectives:** The present study was designed to identify and evaluate antifungal susceptibility patterns of *Candida* spp. isolated from hematological malignancy patients with oral candidiasis.

**Methods:** Samples were collected from the oral cavity of 138 patients and confirmed for oropharyngeal candidiasis by microscopic examination and fungal culture. Isolated *Candida* strains were identified by ITS-PCR. Hyphal wall protein 1 (*HWP1*) was amplified to differentiate the *Candida albicans* complex. *In vitro* antifungal susceptibility tests against fluconazole, amphotericin B, and caspofungin were performed according to CLSI M27-A3/S4.

**Results:** The study enrolled 120 patients, including 74 (61.66%) females and 46 (38.33%) males. The mean age of affliction by fungal infections was  $42 \pm 8$  years. Patients with acute myeloid leukemia (44%) comprised the majority of cases. The most commonly isolated species among patients were *C. albicans* (n = 110; 91.6%), *C. glabrata* (n = 8; 6.6%), and *C. africana* (n = 2; 2.8%). The overall resistance of *C. albicans* was 2.7% to fluconazole, and 1.8% to amphotericin B. *Candida glabrata* showed 12.5% resistance to amphotericin B. All *Candida* spp. isolates from patients were susceptible to caspofungin.

**Conclusions:** Local information about the increasing resistance to fluconazole in both *C. albicans* and non *albicans Candida* species and their antifungal susceptibility is useful for deciding on antifungal prophylaxis and selecting the empirical therapy of cancer patients.

**Keywords:** Oral Candidiasis, Hematologic Malignancy, *Candida albicans*

## 1. Background

Oropharyngeal candidiasis is a fungal infection in the mouth caused by *Candida* spp. The disease generates irritating infections accompanied by several symptoms like angular cheilitis, thrush (pseudomembranous), and erythematous (1). The development of this infection may lead to esophagitis and life-threatening systemic infection (2). One of the most important predisposing factors to fungal infections, especially in malignancies, is chemotherapy. The incidence of oropharyngeal candidiasis, as reported in several studies, ranges from 6 to 53% among cancer patients on chemotherapy (3). *Candida* spp. are normal commensals in humans and may be associated with certain virulence factors (4) If not treated rapidly, especially in pa-

tients on chemotherapy, the colonization of *Candida* spp. with invasion to the mucosa may spread the infection to the bloodstream, leading to disseminated disease (5). Recent data suggest an increased incidence of non *albicans Candida* (NAC) (6). The antifungal susceptibility patterns of *Candida albicans* and non *albicans Candida* are different. Microbiological and clinical evaluations should be emphasized in cancer patients.

## 2. Objectives

The present study was designed to identify and evaluate the antifungal susceptibility patterns of *Candida* spp. isolated from patients with oral candidiasis in patients with hematological malignancies.

### 3. Methods

#### 3.1. Study Design

In this study, 138 patients with hematological malignancies were admitted from October 2018 to September 2019 in Iran. Oropharyngeal candidiasis occurred in 120 of them. The study was conducted in the bone marrow, hematology, oncology, and transplant wards of Taleghani Hospital in Tehran, Iran. The patients received antifungal prophylaxis before and during chemotherapy.

#### 3.2. Sample Collection and Initial Identification of Isolates

Two sterile swabs were used to collect samples from the oral cavity. One swab was used for microscopic examination (potassium hydroxide (KOH) 10% preparation and methylene blue staining) and the other one for fungal culture on Sabouraud Dextrose Agar (SDA, Merck, Germany) with chloramphenicol (50 µg/mL) (Merck, Germany). Oropharyngeal candidiasis was confirmed by the existence of budding yeast cells and pseudohyphae in the direct microscopic examination and positive culture of oral swabs. For preliminary species identification, white colonies in positive cultures were inoculated on the CHROMagar candida *Candida* medium (CHROMagar, Paris, France).

#### 3.3. Molecular Identification of Species

The DNA was extracted from 24 h fresh colony cultures of all 120 isolates, using the method previously described (7). The final solution (DNA template) was kept at -20°C until use as a template for PCR.

#### 3.4. PCR Sequencing of ITS Regions for NAC Identification

The ITS regions of the rDNA gene of isolates were amplified by universal fungal primers, ITS1 (5'-TCCGTAGGTGAACCTGCGG-3') and ITS4 (5'-CCTCCGCTATTGATATGC-3') (8). The contents of the PCR mixture were according to a method previously described (7). The PCR products were subjected to the sequencing for the precise identification of isolates. The PCR products were subjected to sequencing using the ITS1 and ITS4 primers by employing the Big Dye Terminator cycle sequencing kit (Bioneer, Korea).

#### 3.5. Amplification of the HWPI Gene for Differentiating *C. albicans* Complex

The *C. albicans* complex consisting of *C. albicans*, *C. dubliniensis*, and *C. africana* was differentiated using the hyphal wall protein 1 (HWPI) gene with primers CR-f 5'-GCTACCACTTCAGAATCATCATC-3' and CR-r 5'-GCACCTTCAGTCGTAGACG-3'. (9). The PCR condition was defined similarly to the ITS region amplification

condition. The PCR products belonging to two *C. africana* isolates were sequenced (Bioneer, Korea) by CR-f/CR-r primers and compared with the GenBank database using the NCBI database (<https://www.ncbi.nlm.nih.gov>).

#### 3.6. Antifungal Susceptibility Testing

Fluconazole, amphotericin B, and caspofungin were purchased from Sigma-Aldrich (USA). The antifungal agents were diluted with RPMI 1640 medium (Invitrogen, Gibco) and buffered to pH = 7.0 with 0.165 M Morpholine Propane Sulfonic Acid (MOPS) buffer (Sigma, USA) (10). Tests were performed in 96-well microtiter plates. Cell suspensions were adjusted to give a final inoculum concentration of about  $0.5 \times 10^3$  to  $2.5 \times 10^3$  CFU/mL. The plates were incubated at 35°C for 48 h. The minimum inhibitory concentration (MIC) was determined for the identified *Candida* spp. including *C. albicans* (n = 110), *C. glabrata* (n = 8) and *C. africana* (n = 2) according to the CLSI guidelines, M27-A3/S4 document (11). *Candida parapsilosis* ATCC 22019 was used as quality control. All the tests were carried out in duplicate. The MIC breakpoints were defined for each antifungal agent as follows: fluconazole (susceptible MIC  $\leq 8$  µg/mL; resistant MIC  $> 8$  µg/mL), amphotericin B (susceptible MIC  $\leq 1$  µg/mL; resistant MIC  $> 1$  µg/mL), and caspofungin (susceptible MIC  $\leq 0.25$  µg/mL; resistant MIC  $\geq 1$  µg/mL) (12).

## 4. Results

#### 4.1. Demographic Data

Out of 138 patients with hematological malignancy, 120 were included, and 18 were excluded because they had negative culture results. There were 74 (61.66%) females and 46 (38.33%) males (with a male: female ratio of 0.62) in the study sample (Table 1). The mean age of affliction by fungal infections was  $42 \pm 8$  years. It seemed that most patients were afflicted by fungal infections due to the induction phase of chemotherapy. Table 2 shows information regarding the type of hematological malignancy in the patients.

Patients with acute myeloid leukemia (44%) comprised the majority of cases, followed by Hodgkin lymphoma malignancy patients (12.5%). Furthermore, the lowest frequency of leukemia belonged to the non-Hodgkin lymphoma with 4.1% prevalence. All the patients underwent chemotherapy. Based on clinical signs and symptoms, white plaques (72%), dry mouth (65%), pain in the oral cavity (52%), redness of the mucosa (32%), dysphagia (25%), and altered taste sensation (22%) were common clinical findings, in sequence.

**Table 1.** Age and Gender Distribution of Patients

Gender	Age Range, y				Total	Total, %
	< 20	21 - 40	41 - 60	> 60		
Male	2	23	10	11	46	38.33
Female	3	46	20	5	74	61.66
Total	5	69	30	16	120	100

**Table 2.** The Distribution of Patients Based on Hematological Malignancy and Sex<sup>a</sup>

Hematological Malignancy	Male	Female	Total
Acute myeloid leukemia	24 (45.3)	29 (54.7)	53 (44)
Hodgkin lymphoma	6 (40)	9 (60)	15 (12.5)
Chronic myelogenous leukemia	2 (16.7)	10 (83.3)	12 (10)
Multiple myeloma	2 (18.1)	9 (81.8)	11 (9.1)
Acute lymphoblastic leukemia	4 (40)	6 (60)	10 (8.3)
Myelodysplastic syndrome	2 (28.5)	5 (71.5)	7 (5.8)
Chronic lymphocytic leukemia	4 (57.1)	3 (42.9)	7 (5.8)
Non-Hodgkin lymphoma	2 (40)	3 (60)	5 (4.1)
Total	46 (38.4)	74 (61.6)	120 (100)

<sup>a</sup>Values are expressed as No. (%).

#### 4.2. Results of Initial and Molecular Identification of Isolates

Overall, 135 (97.8%) and 138 (100%) of these patients had positive results for KOH smear and methylene blue staining (microscopic examination), respectively. Out of 138 patients with positive results in methylene blue staining, 15 (10.8%) showed yeast and budding yeast and 123 (89%) exhibited budding yeast and pseudohyphae. Eighteen (13%) patients had positive results in microscopic examination, which were not detected in culture. All positive cultures were speciated by the CHROMagar *Candida* medium. The specific patterns of non-green colonies of *Candida* spp. using the amplification of the ITS region after sequencing indicated *C. glabrata* (n = 8).

#### 4.3. Results of Amplification of *HWPI* Gene

Based on the amplification of the *HWPI* gene and production of DNA fragment, out of 112 colonies of *C. albicans*, 110 isolates were *C. albicans* (900 bp), and only two isolates were identified as *C. africana* (700 bp) (Figure 1). In this survey, *C. dubliniensis* (569 bp) was not found. The identification of the two isolates of *C. africana* was confirmed by *HWPI* sequencing (12). The alignment of the obtained sequence in BLAST revealed a 98.98% identity with *C. africana*, and the sequences were deposited in the GenBank under accession number 2275155.



**Figure 1.** Discrimination of *Candida africana* and *C. albicans*. Lanes 1, *C. albicans* ATCC 10231 (900 bp); lanes 2 and 4, *C. africana* (700 bp); lane 3 and 5, clinical isolates of *C. albicans* (900 bp); lane M, molecular size marker 100 bp.

#### 4.4. Results of Antifungal Susceptibility Testing

Antifungal susceptibility testing was performed using the broth microdilution method according to CLSI document M27-A3/S4 (Table 3). None of the isolates of *C. albicans* and *C. glabrata* were resistant to caspofungin. In the present study, non *albicans* *Candida* isolates showed a lower percentage of resistance than *C. albicans* did.

#### 4.5. Results of Antifungal Prophylaxis

The resistance rate to fluconazole was 2.7% among *C. albicans* species isolated from patients. There was no relationship between prophylaxis with voriconazole and resistance to amphotericin B (P = 0.309). There was a significant association between prophylaxis with fluconazole and resistance to fluconazole (P < 0.0001) (Table 4).

### 5. Discussion

Oral candidiasis is a major problem among cancer patients on cytotoxic therapy. Malignancy, chemotherapy, and radiotherapy are the main predisposing factors that compromise the immune system and put the patient at the risk of oropharyngeal candidiasis. In the present study, the most common sign observed was the presence of white

**Table 3.** Antifungal Susceptibility Testing Against Yeast Species Isolated From Patients with Hematological Malignancies

<i>Candida</i> species	Antifungal	Breakpoints, $\mu\text{g/mL}$	Number of Isolates	Resistance, %
<i>C. albicans</i> (n = 110)	Fluconazole	R > 8	3	2.7
		I = 4	28	
		S $\leq$ 2	79	
	Amphotericin B	R > 1	2	1.8
		S $\leq$ 1	108	
	Caspofungin	R $\geq$ 1	0	0
I = 0.5		2		
S $\leq$ 0.25		108		
<i>C. glabrata</i> (n = 8)	Fluconazole	R > 64	0	0
		I $\leq$ 32	4	
		S = ND	4	
	Amphotericin B	R > 1	1	12.5
		S $\leq$ 1	7	
	Caspofungin	R $\geq$ 0.5	0	0
I = 0.25		1		
S $\leq$ 0.12		7		
<i>C. africana</i> (n = 2)	Fluconazole	R > 8	1	50
		I = 4	1	
		S $\leq$ 2	-	
	Amphotericin B	R > 1	1	50
		S $\leq$ 1	1	
	Caspofungin	R $\geq$ 1	-	0
I = 0.5		-		
S $\leq$ 0.25		2		

<sup>z</sup>Abbreviations: S, susceptible; I, intermediate; R, resistant; ND, not determined.

**Table 4.** Antifungal Prophylaxis of Patients with Hematological Malignancies Receiving Chemotherapy<sup>a</sup>

Hematological Malignancies	Prophylaxis		Without Prophylaxis	Cases
	Voriconazole	Fluconazole		
Acute myeloid leukemia	36 (75)	10 (21)	2 (4)	48
Hodgkin lymphoma	3 (30)	7 (70)	0	10
Chronic myelogenous leukemia	2 (25)	6 (75)	0	8
Multiple myeloma	5 (38)	8 (62)	0	13
Acute lymphoblastic leukemia	4 (36)	7 (64)	0	11
Myelodysplastic syndrome	10 (83)	2 (17)	0	12
Chronic lymphocytic leukemia	0	8 (100)	0	8
Non-Hodgkin lymphoma	3 (30)	7 (70)	0	10
<b>Total cases</b>				120

<sup>a</sup>Values are expressed as No. (%).

plaques (72%), followed by dryness of the mouth. In a similar work conducted by Jayachandran et al., the most commonly encountered symptom was dryness of the mouth (65%), and the most common sign among them was the presence of white plaques (3). Also, other previous studies reported other common oral manifestations such as the hairy tongue, oral mucosa atrophy, ulcers, and red and white lesions (13). By evaluating the clinical details, there

was a significant association between the dryness of the mouth and the isolation of *Candida* from the oral cavity ( $P < 0.0001$ ) (3). Malignancies and associated therapeutic interventions like chemotherapy and radiotherapy can weaken the cell-mediated host immunity, which can control invasive fungal infections. These cancer therapies can induce hyposalivation and xerostomia with increased oral yeast proliferation, colonization, and infection (14).

*Candida* spp. can be found as commensal microorganisms in the oral cavity in approximately 10% to 70% of the total population (15). In such conditions, the proliferation rate of *Candida* spp. may increase and make an infection. In this study, the most commonly isolated species among patients were *C. albicans* (91.6%), followed by *C. glabrata* (6.6%) and *C. africana* (2.8%). Previous studies showed that *C. albicans* was the most frequently isolated species, followed by *C. glabrata*, but *C. africana* was not found in these studies (16, 17). Notably, this is the first survey from Iran reporting that *C. africana* is responsible for oropharyngeal candidiasis in hematopoietic malignancy patients. *Candida albicans* is the most commonly encountered species in oropharyngeal candidiasis. The change in the etiology of oral candidiasis from *C. albicans* to non *albicans* *Candida* is particularly challenging for choosing an appropriate antifungal drug. The immunosuppressed condition of cancer patients may lead to the colonization of non *albicans* *Candida*.

One of the most vital implications of these findings was highlighting the importance of non-*albicans* *Candida* species and their significant role in mortality (3, 14). Out of the 120 patients surveyed in the present study, there were 38.4% male patients and 61.6% female patients. Most patients (57.5%) were between 21 and 40-years-old. However, patients younger than 20-years-old comprised a minority of the patients (4%). Based on clinical symptoms, non-Hodgkin lymphoma was the least frequent class of hematologic malignancies among patients (4.1%). The most frequent hematological malignancy among patients was acute myeloid leukemia (44.1%).

Patients suffering from acute myeloid leukemia had the lowest rate of absolute neutrophil count among all patients. This finding can justify the higher rate of patients with acute myeloid leukemia suffering from oropharyngeal candidiasis among all patients. A previous epidemiological study reported that the total cancer incidence in Iran was higher in men than in women. However, gender differences were not significant (18). Morbidity and mortality of invasive fungal infections after induction chemotherapy of leukemia patients and after allogeneic stem cell transplantation are not to be overlooked. The risk of invasive fungal infections increases significantly with the use of more intensive chemotherapy (19).

Patients with prolonged neutropenia (< 500 cells/ $\mu$ L for > 7 days) have substantial risks for invasive fungal infection, therefore prophylaxis should be prescribed for them (20). In the present study, fluconazole and voriconazole were used as prophylactic drugs to protect patients against invasive fungal infections. There was a significant association between prophylaxis with fluconazole and resistance to fluconazole ( $P < 0.0001$ ). It seems that exposure to fluconazole increases the colonization of fluconazole-

resistant *Candida* spp. in cancer patients. Fluconazole exposure may be an independent risk factor for resistance (21). Approximately, 27.12% of the isolates of *Candida* spp. were intermediated to fluconazole, revealing a decreased sensitivity to fluconazole in *C. albicans* ( $n = 28$ ) and *C. glabrata* ( $n = 4$ ), (Table 3). Nystatin, fluconazole, and itraconazole are commonly used for the treatment of oral candidiasis. Voriconazole, posaconazole, and sometimes echinocandins are reserved for complicated cases (21).

Amphotericin B is recommended especially for invasive infections caused by fluconazole-resistant strains (8). Finally, the efficacy of caspofungin is similar to that of amphotericin B and is highly effective in achieving favorable responses in patients (22). Many studies have revealed that fluconazole resistance is increasing among *C. albicans* and non *albicans* *Candida* isolates causing oropharyngeal candidiasis infection (23). The molecular mechanisms of *C. albicans* resistance to fluconazole can be summarized as decrease drug concentration, target site alteration, upregulation of the target enzyme, and development of bypass pathways (24).

A previous study confirmed our results, reporting that the resistance rate to fluconazole was 3% among *C. albicans* isolates (25). According to some meta-analyses, the use of fluconazole as a prophylaxis of invasive fungal infections of cancer patients should be limited to populations whose incidence of invasive candidemia is higher than 7% to 10% for decreasing fluconazole exposure and resistance (26). All *C. glabrata* strains were susceptible and intermediate to fluconazole. Furthermore, the resistance rates of *C. albicans*, *C. glabrata*, and *C. africana* isolates to amphotericin B were 1.8%, 12.5%, and 50%, respectively. Also, all *Candida* spp. isolates from patients were susceptible to caspofungin. Badiie et al. showed the resistance rates of *C. albicans* and *C. glabrata* to amphotericin B were 3.3% and 11.1%, respectively (5).

### 5.1. Conclusions

Due to the significance of oral candidiasis in hematological malignancies, the identification and antifungal susceptibility of *Candida* spp. have particular importance. Our local information about the increasing resistance to fluconazole in *Candida* spp. is useful for deciding on the antifungal prophylaxis and selecting the empirical therapy of cancer patients.

### Footnotes

**Authors' Contribution:** Study concept: Masoud Mardani, Sara Abolghasemi and Ensieh Lotfali. Analysis and interpretation of data: David Darvishnia. Drafting of the manuscript: Reza Ghasemi, Mohammad Mahdi Rabiei, and



Ensieh Lotfali. Critical revision: Masoud Mardani, Sara Abolghasemi, and Azam Fattahi.

**Conflict of Interests:** None.

**Ethical Approval:** The project received approval from the Ethics Committee of Shahid Beheshti University of Medical Sciences, Tehran, Iran (code: IR.SBMU.RETECH.REC.1398.208).

**Funding/Support:** This study was financially supported by the Infectious Diseases and Tropical Medicine Research Center, Shahid Beheshti University of Medical Sciences (SBMU), Tehran, Iran.

## References

- Koohi F, Salehiniya H, Shamlou R, Eslami S, Ghojogh ZM, Kor Y, et al. Leukemia in Iran: epidemiology and morphology trends. *Asian Pac J Cancer Prev*. 2015;**16**(17):7759–63. doi: [10.7314/apjcp.2015.16.17.7759](https://doi.org/10.7314/apjcp.2015.16.17.7759). [PubMed: [26625794](https://pubmed.ncbi.nlm.nih.gov/26625794/)].
- Shankland KR, Armitage JO, Hancock BW. Non-Hodgkin lymphoma. *Lancet*. 2012;**380**(9844):848–57. doi: [10.1016/S0140-6736\(12\)60605-9](https://doi.org/10.1016/S0140-6736(12)60605-9). [PubMed: [22835603](https://pubmed.ncbi.nlm.nih.gov/22835603/)].
- Jayachandran AL, Katragadda R, Thyagarajan R, Vajravelu L, Manikesi S, Kaliappan S, et al. candidiasis among cancer patients attending a tertiary care hospital in Chennai, south India: an evaluation of clinico-mycological association and antifungal susceptibility pattern. *Can J Infect Dis Med Microbiol*. 2016;**2016**:8758461. doi: [10.1155/2016/8758461](https://doi.org/10.1155/2016/8758461). [PubMed: [27403171](https://pubmed.ncbi.nlm.nih.gov/27403171/)]. [PubMed Central: [PMC4923570](https://pubmed.ncbi.nlm.nih.gov/PMC4923570/)].
- Ansari S, Shirzadi E, Elahi M. The prevalence of fungal infections in children with hematologic malignancy in Ali-Asghar children hospital between 2005 and 2010. *Iran J Ped Hematol Oncol*. 2015;**5**(1):1–10. [PubMed: [25914797](https://pubmed.ncbi.nlm.nih.gov/25914797/)]. [PubMed Central: [PMC4402151](https://pubmed.ncbi.nlm.nih.gov/PMC4402151/)].
- Badiee P, Badali H, Boekhout T, Diba K, Moghadam AG, Hossaini Nasab A, et al. Antifungal susceptibility testing of Candida species isolated from the immunocompromised patients admitted to ten university hospitals in Iran: comparison of colonizing and infecting isolates. *BMC Infect Dis*. 2017;**17**(1):727. doi: [10.1186/s12879-017-2825-7](https://doi.org/10.1186/s12879-017-2825-7). [PubMed: [29157206](https://pubmed.ncbi.nlm.nih.gov/29157206/)]. [PubMed Central: [PMC5697407](https://pubmed.ncbi.nlm.nih.gov/PMC5697407/)].
- Akpan A, Morgan R. Oral candidiasis. *Postgrad Med J*. 2002;**78**(922):455–9. doi: [10.1136/pmj.78.922.455](https://doi.org/10.1136/pmj.78.922.455). [PubMed: [12185216](https://pubmed.ncbi.nlm.nih.gov/12185216/)]. [PubMed Central: [PMC1742467](https://pubmed.ncbi.nlm.nih.gov/PMC1742467/)].
- Ghajari A, Lotfali E, Ahmadi NA, Nazer Fassihi P, Shahmohammadi N, Ansari S, et al. Isolation of different species of Candida in patients with vulvovaginal candidiasis from Damavand, Iran. *Arch Clin Infect Dis*. 2018;**13**(6). doi: [10.5812/archcid.59291](https://doi.org/10.5812/archcid.59291).
- Malani AN, Kauffman CA. Candida urinary tract infections: treatment options. *Expert Rev Anti Infect Ther*. 2007;**5**(2):277–84. doi: [10.1586/14787210.5.2.277](https://doi.org/10.1586/14787210.5.2.277). [PubMed: [17402842](https://pubmed.ncbi.nlm.nih.gov/17402842/)].
- Romeo O, Criseo G. First molecular method for discriminating between Candida africana, Candida albicans, and Candida dubliniensis by using hwp1 gene. *Diagn Microbiol Infect Dis*. 2008;**62**(2):230–3. doi: [10.1016/j.diagmicrobio.2008.05.014](https://doi.org/10.1016/j.diagmicrobio.2008.05.014). [PubMed: [18640803](https://pubmed.ncbi.nlm.nih.gov/18640803/)].
- Lotfali E, Kordbacheh P, Mirhendi H, Zaini F, Ghajari A, Mohammadi R, et al. Antifungal susceptibility analysis of clinical isolates of Candida parapsilosis in Iran. *Iran J Public Health*. 2016;**45**(3):322–8. [PubMed: [27141494](https://pubmed.ncbi.nlm.nih.gov/27141494/)]. [PubMed Central: [PMC4851746](https://pubmed.ncbi.nlm.nih.gov/PMC4851746/)].
- Wayne P. Clinical and Laboratory Standards Institute: Reference method for broth dilution antifungal susceptibility testing of yeasts; approved standard. *CLSI document M27-A3 and Supplement S*. 2008;**3**:6–12.
- Lotfali E, Mardani M, Abolghasemi S, Darvishnia D, Rabiei MM, Ghasemi R, et al. Isolation of Candida africana in oral candidiasis: First report among cancer patients in Iran. *Curr Med Mycol*. 2020. doi: [10.18502/cmm.6.2.2695](https://doi.org/10.18502/cmm.6.2.2695).
- Ghapanchi J, Rezaee M, Kamali F, Lavaee F, Shakib E. Prevalence of oral and craniofacial manifestations of hematological dyscrasias at Shiraz Nemazee Hospital. *Middle East J Cancer*. 2014;**5**(3):145–9.
- Ramla S, Sharma V, Patel M. Influence of cancer treatment on the Candida albicans isolated from the oral cavities of cancer patients. *Support Care Cancer*. 2016;**24**(6):2429–36. doi: [10.1007/s00520-015-3035-8](https://doi.org/10.1007/s00520-015-3035-8). [PubMed: [26638003](https://pubmed.ncbi.nlm.nih.gov/26638003/)].
- Bunetel L, Tamanai-Shacoori Z, Martin B, Autier B, Guiller A, Bonnaure-Mallet M. Interactions between oral commensal Candida and oral bacterial communities in immunocompromised and healthy children. *J Mycol Med*. 2019;**29**(3):223–32. doi: [10.1016/j.mycmed.2019.06.004](https://doi.org/10.1016/j.mycmed.2019.06.004). [PubMed: [31235209](https://pubmed.ncbi.nlm.nih.gov/31235209/)].
- Alborzi A, Davarpanah MA. Distributions and antifungal susceptibility of Candida species from mucosal sites in HIV positive patients. *Arch Iran Med*. 2010;**13**(4):282–7. [PubMed: [20597560](https://pubmed.ncbi.nlm.nih.gov/20597560/)].
- Sobel JD, Ohmit SE, Schuman P, Klein RS, Mayer K, Duerr A, et al. The evolution of Candida species and fluconazole susceptibility among oral and vaginal isolates recovered from human immunodeficiency virus (HIV)-seropositive and at-risk HIV-seronegative women. *J Infect Dis*. 2001;**183**(2):286–93. doi: [10.1086/317936](https://doi.org/10.1086/317936). [PubMed: [11204125](https://pubmed.ncbi.nlm.nih.gov/11204125/)].
- Beiranvand S, Zarea K, Ghanbari S, Tuvesson H, Keikhaei B. Ten years incidence of cancer in Iran; a systematic review and meta-analysis. *Clin Epidemiol Glob Health*. 2018;**6**(2):94–102. doi: [10.1016/j.cegh.2017.10.005](https://doi.org/10.1016/j.cegh.2017.10.005).
- Lee CH, Lin C, Ho CL, Lin JC. Primary fungal prophylaxis in hematological malignancy: a network meta-analysis of randomized controlled trials. *Antimicrob Agents Chemother*. 2018;**62**(8). doi: [10.1128/AAC.00355-18](https://doi.org/10.1128/AAC.00355-18). [PubMed: [29866872](https://pubmed.ncbi.nlm.nih.gov/29866872/)]. [PubMed Central: [PMC6105838](https://pubmed.ncbi.nlm.nih.gov/PMC6105838/)].
- Mellinghoff SC, Panse J, Alakel N, Behre G, Buchheidt D, Christopheit M, et al. Primary prophylaxis of invasive fungal infections in patients with haematological malignancies: 2017 update of the recommendations of the Infectious Diseases Working Party (AGIHO) of the German Society for Haematology and Medical Oncology (DGHO). *Ann Hematol*. 2018;**97**(2):197–207. doi: [10.1007/s00277-017-3196-2](https://doi.org/10.1007/s00277-017-3196-2). [PubMed: [29218389](https://pubmed.ncbi.nlm.nih.gov/29218389/)]. [PubMed Central: [PMC5754425](https://pubmed.ncbi.nlm.nih.gov/PMC5754425/)].
- Garnacho-Montero J, Diaz-Martin A, Garcia-Cabrera E, Ruiz Perez de Pipaon M, Hernandez-Caballero C, Aznar-Martin J, et al. Risk factors for fluconazole-resistant candidemia. *Antimicrob Agents Chemother*. 2010;**54**(8):3149–54. doi: [10.1128/AAC.00479-10](https://doi.org/10.1128/AAC.00479-10). [PubMed: [20498325](https://pubmed.ncbi.nlm.nih.gov/20498325/)]. [PubMed Central: [PMC2916332](https://pubmed.ncbi.nlm.nih.gov/PMC2916332/)].
- Villanueva A, Gotuzzo E, Arathoon EG, Noriega LM, Kartsonis NA, Lupinacci RJ, et al. A randomized double-blind study of caspofungin versus fluconazole for the treatment of esophageal candidiasis. *Am J Med*. 2002;**113**(4):294–9. doi: [10.1016/s0002-9343\(02\)01191-9](https://doi.org/10.1016/s0002-9343(02)01191-9). [PubMed: [12361815](https://pubmed.ncbi.nlm.nih.gov/12361815/)].
- Jahanshiri Z, Manifar S, Moosa H, Asghari-Paskiabi F, Mahmoodzadeh H, Shams-Ghahfarokhi M, et al. Oropharyngeal candidiasis in head and neck cancer patients in Iran: Species identification, antifungal susceptibility and pathogenic characterization. *J Mycol Med*. 2018;**28**(2):361–6. doi: [10.1016/j.mycmed.2018.01.001](https://doi.org/10.1016/j.mycmed.2018.01.001). [PubMed: [29602636](https://pubmed.ncbi.nlm.nih.gov/29602636/)].
- Ngouana TK, Drakulovski P, Krasteva D, Toghueo RK, Kouanfack C, Reynes J, et al. Genetic diversity of the Hwp1 gene and HIS3, EF3, CDC3 microsatellites and antifungal susceptibility profiles of Candida albicans isolates from Yaounde HIV-infected patients. *Med Mycol*. 2017;**55**(5):546–54. doi: [10.1093/mmy/myw108](https://doi.org/10.1093/mmy/myw108). [PubMed: [27744307](https://pubmed.ncbi.nlm.nih.gov/27744307/)].
- Arastehfar A, Daneshnia F, Farahyar S, Fang W, Salimi M, Salehi M, et al. Incidence and spectrum of yeast species isolated from the oral cavity of Iranian patients suffering from hematological malignancies. *J Oral Microbiol*. 2019;**11**(1):1601061. doi: [10.1080/20002297.2019.1601061](https://doi.org/10.1080/20002297.2019.1601061). [PubMed: [31044032](https://pubmed.ncbi.nlm.nih.gov/31044032/)]. [PubMed Central: [PMC6484487](https://pubmed.ncbi.nlm.nih.gov/PMC6484487/)].
- Bennett JE, Dolin R, Blaser MJ. *Mandell, Douglas, and Bennett's principles and practice of infectious diseases*. Philadelphia, PA: Elsevier Saunders; 2020.