



# Multipathogen Detection in Patients with Respiratory Tract Infection: Identification of Non-respiratory Viruses Using Multiplex Real-time Polymerase Reaction

Khosrow Agin<sup>1,2</sup>, Zahra Heydarifard<sup>3,4</sup>, Leila Ghalichi<sup>5</sup>, Mahmood Yaghoobi<sup>6</sup>, Hamidreza Hagh Ranjbar<sup>2,7,4</sup>, Seyed Mohammad Jazayeri<sup>2,4,6</sup> and Iman Rezaee Azhar<sup>2,4,6,\*</sup>

<sup>1</sup>Loqman Hospital, Shahid Beheshti University of Medical Sciences, Tehran, Iran

<sup>2</sup>Medical Genetics and Molecular Diagnosis Laboratory, Laleh General Hospital, Tehran, Iran

<sup>3</sup>Department of Virology, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran

<sup>4</sup>Research Center for Clinical Virology, Tehran University of Medical Sciences, Tehran, Iran

<sup>5</sup>Mental Health Research Center, Psychosocial Health Research Institute, Iran University of Medical Sciences, Tehran, Iran

<sup>6</sup>Aramesh Pathobiology and Genetic Laboratory, Tehran, Iran

<sup>7</sup>Department of Cell and Molecular biology & Microbiology, Faculty of Biological Science and Technology, University of Isfahan, Isfahan, Iran

\*Corresponding author: Research Center for Clinical Virology, Tehran University of Medical Sciences, Tehran, Iran. Email: stemcellbiologist@gmail.com

Received 2021 October 26; Revised 2021 December 13; Accepted 2021 December 14.

## Abstract

**Background:** Due to the overlapping clinical characteristics of respiratory tract infections (RTIs) and the unavailability of appropriate diagnostic techniques, the diagnosis of RTIs is controversial.

**Objectives:** The study aimed to prompt the diagnosis of RTIs using commercial multiplex real-time PCR.

**Methods:** The survey undertook for two years (2019-2020) on 144 flu-negative immunocompetent outpatients. Respiratory samples were examined by multiplex PCR assays.

**Results:** Study population consisted of females (n=77, 53.5%) and males (n=67, 46.5%). The mean age was  $42.8 \pm 23.7$  years. Thirty-one (21.5%) patients were infected with only one viral or bacterial infection. Eighty-two (57%) were infected with more than one pathogen. Ninety-five (37%) and 161 (62%) tests were positive for bacterial and viral pathogens, respectively. Community-acquired Pneumonia (CAP) and atypical CAP pathogens included 17% and 10% of respiratory specimens, respectively. The predominant pathogens consisted of Human Herpes Virus 7 (HHV-7) (n=38, 15.5%), Epstein-Barr Virus (EBV) (n=34, 13.8%), *Mycoplasma pneumoniae* (n=24, 9.8%), and Human Herpes Virus 6 (HHV-6) (n=21, 8.5%). There were associations between pathogen findings and special age categories. Fever, cough, dyspnea, and hemoptysis were associated with certain pathogens. There was no substantial difference between viral and bacterial Ct concerning gender, age group, and comorbidities.

**Conclusions:** Multiplex diagnostic assays significantly increased the rate of appropriate diagnosis of respiratory pathogens. However, further investigation is needed to find non-respiratory viruses' significance in respiratory specimens of immunocompetent symptomatic patients.

**Keywords:** Molecular Diagnostics, Multiplex Real-time PCR, Respiratory Tract Infections

## 1. Background

Respiratory tract infections (RTIs) impose a significant burden on health systems worldwide. However, current diagnostic tools may detect causative pathogens in only 30-40% of RTI patients. Therefore, the causative agents remain unrecognized in most clinical cases, and nonspecific respiratory symptoms are usually treated empirically. By application of current molecular techniques, it is now established that about 25% of community-acquired Pneumonia (CAP) has viral etiology (1,2). Moreover, disease severity and hospital stay length may increase by viral or viral-bacterial

coinfections in CAP (3), escalating the need for intensive care and increased mortality rate in pneumonia patients (2).

Application of sensitive diagnostic assays could identify more than one pathogen in 4-30% of adults and 23-33% of children in prospective studies of CAP (4-8). The detection of respiratory and non-respiratory viruses in RTI patients is challenging. They could be a sole cause may be of viral origin or a coinfection; besides, they can facilitate or worsen the bacterial infection. Differentiating a viral origin from bacterial infection or mixed viral-bacterial infec-

tion could substantially decrease antibiotic use, avoid the associated risk of adverse reactions, reduce antibiotic resistance development, decrease hospital-acquired infections, and improve clinical outcomes (9).

Respiratory multiplex real-time kits are in vitro tests for the quantitative/qualitative detection of pathogens' nucleic acids in throat/nasal swabs, bronchoalveolar lavage, sputum, and culture of human origins for the evaluation of infections with respiratory pathogens detected by commercial kits. These tests contain separate primers and probes, each targeting the compatible sequence on the desired genes (target of interest) with high sensitivity and specificity. These assays have been optimized for the simultaneous detection of pathogens in unknown samples. Currently, the development and improvement of multiplex PCR tests allow accurate characterization of a wide range of viral and bacterial pathogens in acute and non-acute samples (10-12). At present, multiplex PCR assays are the gold standard for diagnosing viral RTI (13), making it possible to diagnose viral and bacterial RTIs rapidly and simultaneously. In addition to the quantitative measurement ability of real-time PCR assays, semi-quantitative microbial load data could be identified to differentiate colonization from true infection by pathogens in symptomatic and asymptomatic patients (14).

## 2. Objectives

The objectives of the present study were (1) to evaluate a multiplex technique as a "syndromic approach" for the detection of a wide range of bacteria and non-respiratory viruses in different clinical settings of RTIs, (2) to identify the frequency of each respiratory pathogen alone or in combination (co-detection) among patients with RTIs, (3) to evaluate the clinical presentation of patients with regards to the pathogens detected, and (4) to assess the age specific to disease distribution in different viral and bacterial pathogens.

## 3. Methods

### 3.1. Study Design

This cross-sectional, double-center investigation was conducted at the Research Center for Clinical Virology (Tehran University of Medical Sciences), collaborating with Laleh general hospital and Aramesh Medical Laboratory, Tehran, Iran, during 2019 - 2020. Patients were referred to Loghman and Laleh Hospitals, related to the Research Center for Clinical Virology Department, with a history of respiratory tract infection associated with the exacerbation of pulmonary symptoms. There was no age limit for the

participants. The inclusion criteria included patients with moderate to severe respiratory symptoms such as cough, dyspnea, hemoptysis, and wheezing, already known for respiratory diseases (small airway diseases) superimposed with acute episodes, and patients who were on antibiotic treatment previously for respiratory infections with unsatisfactory clinical outcomes.

All patients had already been tested for the influenza virus using a triplex real-time PCR assay to detect Flu-A, Flu-B, and H1N1 viruses, all with negative results. The primary clinical manifestations were bronchial asthma, bronchiectasis, tracheobronchitis, and bronchiolitis. A standard chest X-ray showed diffuse and focal reticular patterns, consolidation with lower lobe distribution, air trapping, and congested features in all subjects. Acutely ill patients without previous respiratory tract infection history were excluded from the study.

### 3.2. Sampling

Respiratory specimens were collected from patients, including the sputum, anterior nasal swabs, and throat swabs using stiff synthetic swabs, and transported in the viral transport media to maintain viral nucleic acids in good condition until analysis.

### 3.3. DNA Extraction and Polymerase Chain Reaction

Using a viral/bacterial RNA/DNA nucleic acid extraction kit (ROCHE, Mannheim, Germany), the viral and bacterial nucleic acids were extracted from different respiratory samples according to the manufacturer's instructions. For each multiplex panel, internal control was utilized to exclude false-negative results obtained by PCR inhibitors. Multiplex real-time PCR was carried out on 5 µL of eluted DNA using different panels of respiratory bacterial and respiratory and non-respiratory viral pathogen kits, including Flu, Neuro-9, CAP, and HAP kits (Siemens, Luxembourg), according to the manufacturer's instructions. Semi-quantitative estimation was calculated for each specimen.

### 3.4. Statistical Analyses

The data were analyzed by SPSS version 23. The normality of age data was tested with the Kolmogorov-Smirnov test ( $P = 0.002$ ). The frequency of variables was presented in percentage. The figures were generated with an excel program. The comparison of means was carried out with a nonparametric chi-square test. A  $P$  value  $< 0.05$  was considered statistically significant for all comparisons.

## 4. Results

This study enrolled 144 flu-negative outpatients based on the eligibility criteria. The subjects' age ranged from three to 89 years (mean 42.8, SD 23.7), and 32% of the participants were between 41 and 60 years (Table 1). There were 77 (53.5%) females and 67 (46.5%) males. Thirty-two (22%) patients had a history of comorbidities, including asthma, COPD, and diabetes (data not shown). Thirty-one (21.5%) patients tested negative for all pathogens. Thirty-one (21.5%) patients were infected with only one viral or bacterial agent. Eighty-two (57%) patients were infected with more than one pathogen (viral and/or bacterial). Of a total of 256 multiplex tests, 95 (37%) and 161 (62%) tests were positive for bacterial and viral pathogens, respectively (data not shown). Besides, 23 (16%), 48 (33%), and 61 (42.5%) patients had bacterial, viral, and viral/bacterial coinfections, respectively (Table 1). In 50 (34.7%) cases, we simultaneously detected two organisms (data not shown). The maximum number of organisms detected was six, observed in three cases (data not shown). In addition, CAP and atypical CAP pathogens were found in 17% and 10% of respiratory specimens, respectively (Figure 1).

The distribution of pathogens was higher in females than males; however, this difference was not significant (P value: 0.56, data not shown). Although patients with previous underlying diseases showed a higher rate of bacterial, viral, and viral/bacterial coinfections, it did not reach statistical significance (data not shown). Regarding the history of antibiotic consumption before the tests, 23 had a history of antibiotic therapy. Fourteen and seven of these patients were positive for viral and bacterial infections, respectively, but the positivity rate was not different between those with and without a history of antibiotic therapy (P value: 0.09). The minimum detected Ct was significantly higher in those with a history of antibiotic therapy than others (31 vs. 27, P value: 0.005, data not shown). No other findings were correlated with a history of antibiotic therapy (Table 1). Subjects > 41 years old had more coinfections and multi-infections than other age groups. On the other hand, children (< 15 years old) showed lower rates of coinfections and multi-infections, especially those in the 8-15-year-old group (Table 1).

Figure 1 presents the frequency distribution of viral and bacterial infections. The four predominant pathogens were Human Herpes Virus 7 (HHV-7) (n = 38, 15.5%), Epstein-Barr Virus (EBV) (n = 34, 13.8%), *Mycoplasma pneumoniae* (n = 24, 9.8%), and Human Herpes Virus 6 (HHV-6) (n = 21, 8.5%). Bacterial pathogens responsible for CAP were among the commonest pathogens. Accordingly, *Pseudomonas aeruginosa* and *Klebsiella pneumoniae* responsible for atypical CAP pathogens composed 10% of the pathogens. Table 2 shows

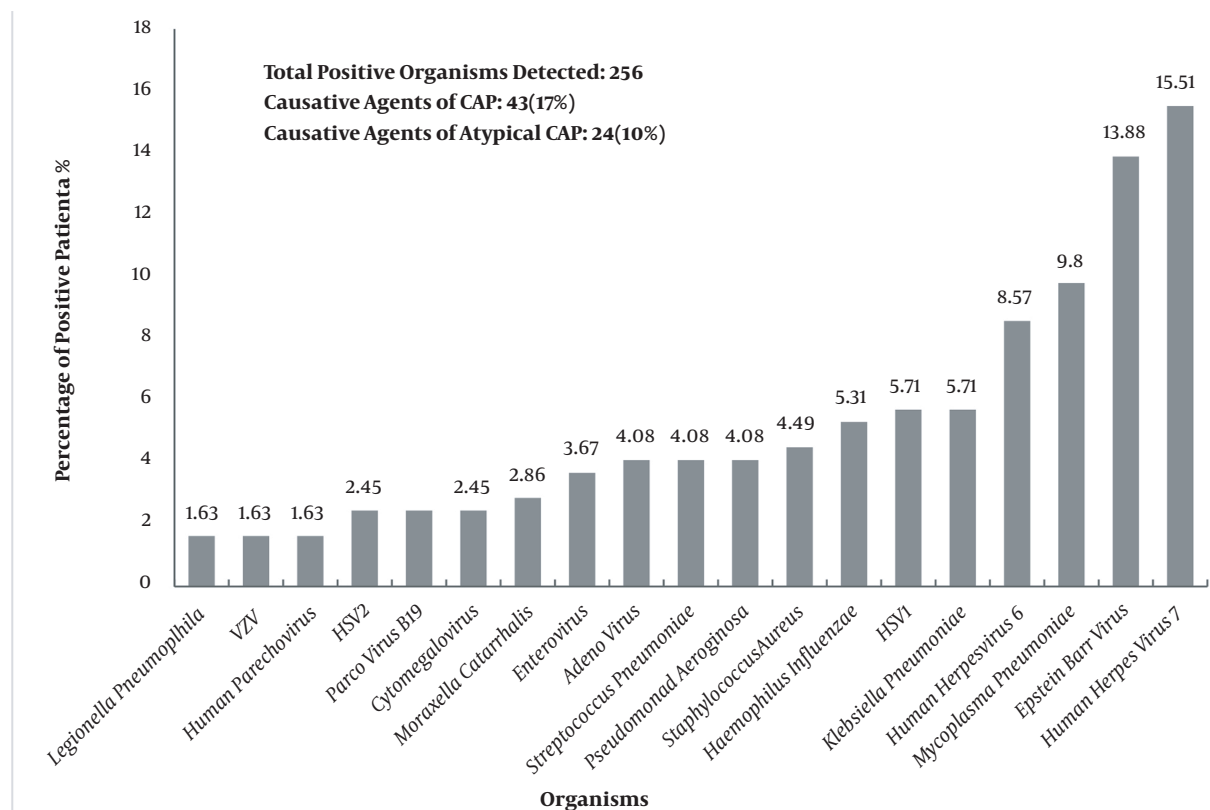
the frequency of viral and bacterial infections against age categories. *Klebsiella pneumoniae* and *P. aeruginosa* (both belonging to atypical CAP) were found mainly in those > 41 years old, especially the elderly (P values: 0.158 and 0.09, respectively). *Mycoplasma pneumoniae*, *Haemophilus influenzae*, HHV-6, and HHV-7 were seen mostly in patients < 7 and 16 - 40 years old (Table 2). *Haemophilus influenzae*, HHV-6, and HHV-7 were mostly observed for those between 8 and 15 years. *Mycoplasma pneumoniae*, HHV-6, HHV-7, EBV, Adenovirus, Parvovirus B19, *K. pneumoniae*, and *Staphylococcus aureus* were dominantly detected in the 41 - 60 age group. Lastly, EBV, *P. aeruginosa*, *K. pneumoniae*, HHV-7, HSV-1, and *S. pneumoniae* were detected in the > 60 age group (Table 2).

Next, we compared the association between clinical symptoms and microbial findings. We found that acute cough, chronic cough (more than 35 days), and dyspnea were the common main symptoms between patients. Except for dyspnea that was more common in adults > 60 years old, the other symptoms were proportionally distributed between different age groups and genders (data not shown). Accordingly, the frequency of wheezing was higher in patients infected with EBV and *M. pneumoniae* (27.78% and 16.67%, respectively), although both were statistically insignificant (Table 3). Hemoptysis was more observed in patients with *M. pneumoniae* (the highest frequency), followed by adenovirus, HSV-1, and *Moraxella catarrhalis* (with equal frequencies) (Table 3). Moreover, fever was predominant in patients infected with EBV, HHV-1, HHV-7, and *M. pneumoniae* (Table 3). Furthermore, dyspnea was more associated with detecting EBV, HHV-7, *S. aureus*, and HHV-6 (listed in the order of significance from the highest to lowest) (Table 3).

We divided patients into acute, chronic, and acute-chronic cough patients regarding the history of cough. Acute cough was accompanied mainly by *M. pneumoniae* and HHV-7 with an equal frequency, followed by EBV, HHV-6, and adenovirus. However, EBV was the dominant pathogen for chronic cough, followed by *M. pneumoniae*, HHV-7, HHV-6, and *K. pneumoniae* (Table 3). Lastly, three pathogens were mainly associated with cough, regardless of being acute or chronic, including *M. pneumoniae*, EBV (18.75%), and HHV-7 (18.75%) (Table 3). Dyspnea and cough were associated with multiple pathogen detection (viral, bacterial, and viral-bacterial); however, these correlations were insignificant compared to other symptoms (data not shown). Regarding the Ct values of the tests, there was no substantial difference between viral and bacterial Ct median values concerning gender, age, and comorbidities (Table 1). However, the minimum detected Ct was significantly higher in those with a history of antibiotic therapy than others (31 vs. 27, P value: 0.005).

**Table 1.** Demographics, Microbiological, and Antibiotic History of Patients <sup>a, b</sup>

Group	All Patients	Any Co-morbidity	Negative for All Microbes	1 Microbe Detected	> 1 Microbe Detected	Bacteria All	Bacteria Median CT	Viruses All	Viruses Median CT	Bacterial Co-detection	Viral Co-detection	Bac/Vir Co-detection
<b>Age (y)</b>												
2 - 7	16 (11.1)	4 (25)	3 (18.7)	2 (12.5)	11 (68.7)	13 (14.2)	31.00	22 (13.6)	32.00	4 (17.3)	7 (14.5)	9 (14.7)
8 - 15	11 (7.6)	4 (36)	2 (18)	1 (9)	8 (72.7)	6 (6.5)	31.50	15 (9.3)	32.00	1 (4.3)	4 (8.3)	6 (9.8)
16 - 40	29 (20.1)	3 (10)	8 (27.5)	5 (17.2)	16 (55)	15 (16.4)	33.00	34 (21.1)	32.00	4 (17.3)	9 (18.7)	11 (18)
41 - 60	46 (31.9)	13 (28)	10 (21.7)	11 (23.9)	25 (54.3)	25 (27.4)	32.00	51 (31.6)	31.50	6 (26)	18 (37.5)	17 (27.8)
> 60	42 (29.1)	8 (19)	8 (19)	12 (28.5)	22 (52.3)	32 (35.1)	32.00	39 (24.2)	32.00	8 (34.7)	10 (20)	18 (29.5)
<b>Gender</b>												
Female	77 (53)	18 (23)	18 (58)	14 (45)	45 (54.8)	38 (54.2)	32.00	53 (53)	32.00	10 (43.4)	25 (52)	34 (55.7)
Male	67 (47)	14 (21)	13 (42)	17 (54.8)	37 (45.1)	32 (45.7)	32.00	47 (47)	32.00	13 (56.5)	23 (48)	27 (44.2)
<b>Antibiotic therapy</b>	23 (15.9)	-	8 (34)	5 (21)	10 (43)	11 (33.3)	34	22 (66.6)	31.5	4 (17)	7 (30)	6 (26)
<b>Total</b>	144	32 (22)	31 (21.5)	31 (21.5)	82 (57)	95 (37)	32.00	161 (62)	32.00	23 (16)	48 (33)	61 (42.5)

<sup>a</sup>Values are expressed as No. (%).<sup>b</sup>Indicates the number of positive tests (not the patients).**Figure 1.** Distribution of pathogen identification

## 5. Discussion

The present study was the first Iranian survey of the frequency of pathogens among RTI outpatients referred to Lohman and Laleh hospitals using multiplex assays during 2019 - 2020. The results clearly showed that us-

ing targeted panels of real-time multiplex PCR for viral and bacterial pathogens increased the chance of a proper diagnosis in patients with respiratory tract infection and provided valuable information about the distribution of pathogens among different age groups. Of all tests, 36% and 64% were positive for bacterial and viral pathogens,

**Table 2.** Prevalence of Pathogens in Different Age Groups <sup>a</sup>

Pathogens	2 - 7 Years	8 - 15 Years	16 - 40 Years	41 - 60 Years	> 60 Years
<i>Klebsiella pneumoniae</i>	1 (3.4)	0 (0.0)	2 (4.1)	6 (7.5)	5 (7.2)
<i>Pseudomonas aeruginosa</i>	0 (0.0)	0 (0.0)	2 (4.1)	8 (9.7)	8 (11.6)
<i>Mycoplasma pneumoniae</i>	6 (20.6)	1 (5.2)	7 (14.2)	8 (9.7)	2 (2.8)
<i>Moraxella catarrhalis</i>	2 (6.8)	1 (5.2)	0 (0.0)	1 (1.2)	3 (4.3)
<i>Haemophilus influenzae</i>	0 (0.0)	3 (16.0)	4 (8.1)	2 (2.4)	4 (5.7)
<i>Staphylococcus aureus</i>	1 (3.4)	0 (0.0)	2 (4.1)	4 (4.7)	4 (5.7)
<i>Chlamydia pneumoniae</i>	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)
<i>Streptococcus pneumoniae</i>	3 (10.6)	1 (5.2)	0 (0.0)	2 (2.4)	4 (5.7)
<i>Legionella pneumophila</i>	0 (0.0)	1 (5.2)	1 (2.0)	1 (1.2)	1 (1.4)
<i>Enterovirus</i>	2 (6.8)	1 (5.2)	2 (4.1)	2 (2.4)	2 (2.8)
Human parechovirus	0 (0.0)	1 (5.2)	1 (2.0)	1 (1.2)	1 (1.4)
Human Herpesvirus 6	4 (14.0)	2 (11.0)	4 (8.1)	7 (8.4)	4 (5.7)
Human herpesvirus 7	4 (14.0)	4 (21.0)	9 (18.3)	15 (18)	6 (9.6)
Parvovirus B19	2 (6.8)	0 (0.0)	0 (0.0)	4 (4.7)	0 (0.0)
Adenovirus	0 (0.0)	1 (5.2)	1 (2.0)	6 (7.5)	2 (2.8)
Cytomegalovirus	0 (0.0)	1 (5.2)	2 (4.1)	2 (2.4)	1 (1.4)
Epstein-Barr virus	2 (6.8)	1 (5.2)	4 (8.1)	9 (10.9)	18 (26.2)
Herpes simplex virus 1	1 (3.4)	1 (5.2)	5 (10.6)	3 (3.3)	4 (5.7)
Herpes simplex virus 2	1 (3.4)	0 (0.0)	3 (6.1)	2 (2.4)	0 (0.0)
<b>Total</b>	<b>29 (100)</b>	<b>19 (100)</b>	<b>49 (100)</b>	<b>83 (100)</b>	<b>69 (100)</b>

<sup>a</sup>Values are expressed as No. (%).

respectively. Although the differences were not statistically significant, we observed higher positivity rates in females, those without a history of antibiotic therapy, and those with underlying disease. Previously, the viral etiology of CAP and the nature of mixed infections with different pathogens had been underestimated due to the limited range of diagnostic methods. However, recent studies on multi-targeted molecular diagnostics have indicated that the viral-bacterial co-detection rate was 59.8% (15). Crotty et al. reported bacterial coinfection and viral coinfection rates of 57.6% and 33%, respectively (16). Another study reported bacterial, viral, and viral-bacterial coinfection rates of 28%, 74%, and 17%, respectively (17).

On the other hand, the above data included the respiratory viral causes of CAP. The present study showed that multiple non-respiratory viruses were detected in at least 62% of moderate cases of CAP. Among non-respiratory viruses, HHV-7, EBV, and HHV-6 were the most frequent ones, comprising 14.9%, 13.3%, and 8.2% of all detected pathogens, respectively. There are several reports on the increased detection of non-respiratory viruses involved in viral CAP in respiratory samples from immunocompetent

patients, although at a lower frequency than in immunocompromised hosts. Besides, EBV has been the most commonly detected one, followed by HHV-7, HHV-6, and CMV at different rates among patients with various respiratory diseases (18-20).

In two different reports on ARDS patients with unknown etiology, Bonizzoli reported a prevalence of 43% for EBV and 28% for HSV-1 and CMV (21), and in the second survey, Tachikawa et al. found CMV (60%), EBV (45%), HSV-1 (31%), HHV-7 (2%), and HHV-6 (5%) (22). Among 108 bronchiectasis patients, 48% were positive for EBV (23). Of 136 patients with chronic obstructive pulmonary disease (COPD), EBV was observed in 65 (48%) severe patients and 31 (46%) moderate patients (24). In patients with interstitial pneumonia, HHV-7 DNA was detected in 79.2%, and HHV-6 DNA was discovered in 12.5% of biopsy tissues (25). On the other hand, EBV and HHV-7 were detected frequently in asymptomatic immunocompetent healthy adults as 9 - 50% (4, 5, 26, 27) and 34% (8, 28), respectively.

A question is raised "What are the roles of viral agents in respiratory secretions either alone or combined with bacterial agents?" Over the past few years, several her-

**Table 3.** Frequency of Clinical Symptoms Concerning Isolated Pathogens <sup>a</sup>

Pathogens	Clinical Symptoms						
	Fever (N = 17)	Wheezing (N = 18)	Hemoptysis (N = 16)	Dyspnea (N = 83)	Chronic Cough (N = 37)	Acute Cough (N = 148)	Chronic and Acute Cough (N = 14)
<i>Legionella pneumophila</i>	0	1 (5.56)	0	1 (1.15)	1 (2.44)	2 (1.25)	0
VZV	1 (5.26)	1 (5.56)	0	0	0	3 (1.88)	0
Human parechovirus	1 (5.26)	0	0	0	0	4 (2.5)	0
Herpes simplex virus 2	0	1 (5.56)	1 (6.25)	1 (1.15)	1 (2.44)	5 (3.12)	1 (6.25)
Parvovirus B19	0	0	1 (6.25)	2 (2.3)	0	5 (3.12)	0
Cytomegalovirus	0	1 (5.56)	0	3 (3.45)	4.88	1 (0.52)	0
<i>Moraxella catarrhalis</i>	0	0	2 (12.5)	3 (3.45)	0	4 (2.5)	0
Enterovirus	0	1 (5.56)	0	0	0	6 (3.75)	0
Adenovirus	0	1 (5.56)	2 (12.5)	4 (4.6)	4 (9.76)	3 (1.88)	0
<i>Streptococcus pneumoniae</i>	2 (10.53)	0	1 (6.25)	4 (4.6)	1 (2.44)	6 (3.75)	0
<i>Pseudomonas aeruginosa</i>	0	1 (5.56)	1 (6.25)	7 (8.05)	0	5 (3.12)	0
<i>Staphylococcus aureus</i>	0	1 (5.56)	0	8 (9.2)	1 (2.44)	7 (4.38)	0
<i>Haemophilus influenzae</i>	0	0	1 (6.25)	4 (4.6)	1 (2.44)	8 (5)	0
Herpes simplex virus 1	3 (15.79)	0	2 (12.5)	5 (5.75)	3 (7.32)	6 (3.75)	0
<i>Klebsiella pneumoniae</i>	0	1 (5.56)	0	4 (4.6)	1 (2.44)	12 (7.5)	1 (6.25)
Human herpesvirus 6	0	0	0	6 (6.9)	4 (9.76)	13 (8.12)	1 (6.25)
<i>Mycoplasma pneumoniae</i>	3 (15.79)	3 (16.67)	3 (18.7)	5 (5.75)	7 (17.07)	19 (11.88)	5 (31.25)
Epstein-Barr virus	4 (21.05)	5 (27.78)	1 (6.25)	13 (14.94)	6 (14.63)	23 (14.38)	3 (18.75)
Human herpes virus 7	3 (15.79)	1 (5.56)	1 (6.25)	13 (14.94)	7 (17.07)	21 (13.12)	3 (18.75)

<sup>a</sup>Values are expressed as No. (%).

pesviruses have been isolated in patients with respiratory tract infections. As known, EBV, HHV-6, and HHV-7 can cause severe pneumonia in immunocompromised hosts and pneumonia in normal individuals without underlying diseases (29, 30). The interaction of these co-pathogens may worsen respiratory disease outcomes. Viral infections may predispose the respiratory tract for secondary bacterial infections by an interaction between bacteria and respiratory viruses (31, 32). However, there is still scarce data on the occurrence of non-respiratory viruses in immunocompetent CAP patients; therefore, further observational data using multiplex assays are needed to fulfill this gap of

knowledge.

Patients > 41 years had more coinfections and multi-infections than other age groups. On the other hand, children < 15 years showed lower rates of coinfection and multi-infection, especially those in the 8 - 15 years old group. Despite a high rate of herpesviruses primary infection in immunocompetent children, detecting these viruses in respiratory secretions might not be a common phenomenon. The other reason could be that children < 15 years old comprised less than 20% of the patient populations in the present study. We found some associations between four significant symptoms and pathogen predom-



inance. Wheezing and cough (both acute and chronic) were primarily observed in the presence of *M. pneumoniae* and EBV. Hemoptysis was mainly associated with *M. pneumoniae*, adenovirus, and HSV-1, indicating the potential of a correlation between major symptoms and specific pathogens. On the other hand, fever, acute/chronic cough, and dyspnea were strongly associated with versatile lists of pathogens. Likewise, in some surveys, wheezing in children and adults had been meaningfully linked with viral pneumonia compared to mixed viral or bacterial infections (33,34). However, we appreciate that clinical manifestations may differ depending on specific coinfection patterns, and significant overlaps in the present study and other reports mitigate the utility of these findings in respiratory-infected patients.

Herpesviruses could persist and establish latency; therefore, differentiation between latent infection and active viral replication by diagnostic assays is paramount. Positive results on qualitative assays may indicate respiratory disease etiology, asymptomatic colonization, microorganism shedding, or upcoming infection (35). Using a semi-quantitative approach, we aimed to use the *Ct* values; however, due to significant overlap in *Ct* value diversities between different detected pathogens, identifying a certain *Ct* value threshold to show the cause and effect relatedness was not possible. The present investigation also has several limitations. The obtained results focused only on pathogens included in the employed respiratory assays; therefore, the presence of other non-detected pathogens could not be excluded. Moreover, the obtained data from a double-center study could not be attributed to the whole country.

### 5.1. Conclusions

In conclusion, given that a substantial proportion of respiratory diseases has been caused by viral pathogens, the unnecessary usage of antibiotics is puzzling due to its unfavorable health consequences and antimicrobial resistance complications. Hence, the multiplex respiratory panels provide the early detection of the pathogen spectrum and benefit patients in the clinical decision-making process. The role of herpesviruses in disease worsening and complications deserved further investigations.

### Acknowledgments

The authors acknowledge the help of the members of Aramesh Genetic Laboratory, especially Mina Hamed-naghsheh and Amir Roshanai.

### Footnotes

**Authors' Contribution:** KHA did conceptualization, investigation, and manuscript preparation; ZH and LGH performed the data analysis and interpretation; SMJ edited the manuscript and arranged work resources; MY, HHR, and IRA performed sampling and experiments; SMJ and IRA supervised the whole project. All authors approved the final version of the manuscript before submission.

**Conflict of Interests:** The authors report no conflict of interest.

**Ethical Approval:** The Ethics Committee of Tehran University of Medical Sciences approved this project (IR.TUMS.SPH.REC.1400.165).

**Funding/Support:** The authors received no financial support for the research, authorship, and/or publication of this article.

**Informed Consent:** The consent documents were signed and dated by the patients.

### References

- Burk M, El-Kersh K, Saad M, Wiemken T, Ramirez J, Cavallazzi R. Viral infection in community-acquired pneumonia: a systematic review and meta-analysis. *Eur Respir Rev.* 2016;**25**(140):178–88. doi: [10.1183/16000617.0076-2015](https://doi.org/10.1183/16000617.0076-2015). [PubMed: [27246595](https://pubmed.ncbi.nlm.nih.gov/27246595/)].
- Voiriot G, Visseaux B, Cohen J, Nguyen LB, Neuville M, Morbieu C, et al. Viral-bacterial coinfection affects the presentation and alters the prognosis of severe community-acquired pneumonia. *Crit Care.* 2016;**20**(1):375. doi: [10.1186/s13054-016-1517-9](https://doi.org/10.1186/s13054-016-1517-9). [PubMed: [27852281](https://pubmed.ncbi.nlm.nih.gov/27852281/)]. [PubMed Central: [PMC5112669](https://pubmed.ncbi.nlm.nih.gov/PMC5112669/)].
- Rappo U, Schuetz AN, Jenkins SG, Calfee DP, Walsh TJ, Wells MT, et al. Impact of early detection of respiratory viruses by multiplex pcr assay on clinical outcomes in adult patients. *J Clin Microbiol.* 2016;**54**(8):2096–103. doi: [10.1128/JCM.00549-16](https://doi.org/10.1128/JCM.00549-16). [PubMed: [27225406](https://pubmed.ncbi.nlm.nih.gov/27225406/)]. [PubMed Central: [PMC4963510](https://pubmed.ncbi.nlm.nih.gov/PMC4963510/)].
- Johansson N, Kalin M, Tiveljung-Lindell A, Giske CG, Hedlund J. Etiology of community-acquired pneumonia: increased microbiological yield with new diagnostic methods. *Clin Infect Dis.* 2010;**50**(2):202–9. doi: [10.1086/648678](https://doi.org/10.1086/648678). [PubMed: [20014950](https://pubmed.ncbi.nlm.nih.gov/20014950/)]. [PubMed Central: [PMC7107844](https://pubmed.ncbi.nlm.nih.gov/PMC7107844/)].
- Johnstone J, Majumdar SR, Fox JD, Marrie TJ. Viral infection in adults hospitalized with community-acquired pneumonia: prevalence, pathogens, and presentation. *Chest.* 2008;**134**(6):1141–8. doi: [10.1378/chest.08-0888](https://doi.org/10.1378/chest.08-0888). [PubMed: [18689592](https://pubmed.ncbi.nlm.nih.gov/18689592/)]. [PubMed Central: [PMC7094572](https://pubmed.ncbi.nlm.nih.gov/PMC7094572/)].
- Lieberman D, Shimoni A, Shemer-Avni Y, Keren-Naos A, Shtainberg R, Lieberman D. Respiratory viruses in adults with community-acquired pneumonia. *Chest.* 2010;**138**(4):811–6. doi: [10.1378/chest.09-2717](https://doi.org/10.1378/chest.09-2717). [PubMed: [20363845](https://pubmed.ncbi.nlm.nih.gov/20363845/)]. [PubMed Central: [PMC7094496](https://pubmed.ncbi.nlm.nih.gov/PMC7094496/)].
- Cevy-Macherel M, Galetto-Lacour A, Gervais A, Siegrist CA, Bille J, Bescher-Ninet B, et al. Etiology of community-acquired pneumonia in hospitalized children based on WHO clinical guidelines. *Eur J Pediatr.* 2009;**168**(12):1429–36. doi: [10.1007/s00431-009-0943-y](https://doi.org/10.1007/s00431-009-0943-y). [PubMed: [19238436](https://pubmed.ncbi.nlm.nih.gov/19238436/)]. [PubMed Central: [PMC7087130](https://pubmed.ncbi.nlm.nih.gov/PMC7087130/)].
- Tsolia MN, Psarras S, Bossios A, Audi H, Paldanius M, Gourgiotis D, et al. Etiology of community-acquired pneumonia in hospitalized school-age children: evidence for high prevalence of viral infections. *Clin*

- Infect Dis.* 2004;**39**(5):681–6. doi: [10.1086/422996](#). [PubMed: [15356783](#)]. [PubMed Central: [PMC7107828](#)].
9. Davey P, Brown E, Charani E, Fenelon L, Gould IM, Holmes A, et al. Interventions to improve antibiotic prescribing practices for hospital inpatients. *Cochrane Database Syst Rev.* 2013;**(4)**. CD003543. doi: [10.1002/14651858.CD003543.pub3](#). [PubMed: [23633313](#)].
  10. Templeton KE, Scheltinga SA, Graffelman AW, Van Schie JM, Crielaard JW, Sillekens P, et al. Comparison and evaluation of real-time PCR, real-time nucleic acid sequence-based amplification, conventional PCR, and serology for diagnosis of *Mycoplasma pneumoniae*. *J Clin Microbiol.* 2003;**41**(9):4366–71. doi: [10.1128/JCM.41.9.4366-4371.2003](#). [PubMed: [12958270](#)]. [PubMed Central: [PMC193789](#)].
  11. Templeton KE, Scheltinga SA, Beersma MF, Kroes AC, Claas EC. Rapid and sensitive method using multiplex real-time PCR for diagnosis of infections by influenza A and influenza B viruses, respiratory syncytial virus, and parainfluenza viruses 1, 2, 3, and 4. *J Clin Microbiol.* 2004;**42**(4):1564–9. doi: [10.1128/JCM.42.4.1564-1569.2004](#). [PubMed: [15071005](#)]. [PubMed Central: [PMC387552](#)].
  12. Scheltinga SA, Templeton KE, Beersma MF, Claas EC. Diagnosis of human metapneumovirus and rhinovirus in patients with respiratory tract infections by an internally controlled multiplex real-time RNA PCR. *J Clin Virol.* 2005;**33**(4):306–11. doi: [10.1016/j.jcv.2004.08.021](#). [PubMed: [15994117](#)]. [PubMed Central: [PMC7185544](#)].
  13. Bonnin P, Mischak F, Kin N, Resa C, Dina J, Gouarin S, et al. Study and interest of cellular load in respiratory samples for the optimization of molecular virological diagnosis in clinical practice. *BMC Infect Dis.* 2016;**16**:384. doi: [10.1186/s12879-016-1730-9](#). [PubMed: [27503120](#)]. [PubMed Central: [PMC4977610](#)].
  14. Jansen RR, Wieringa J, Koekkoek SM, Visser CE, Pajkrt D, Molenkamp R, et al. Frequent detection of respiratory viruses without symptoms: toward defining clinically relevant cutoff values. *J Clin Microbiol.* 2011;**49**(7):2631–6. doi: [10.1128/JCM.02094-10](#). [PubMed: [21543571](#)]. [PubMed Central: [PMC3147826](#)].
  15. Jung J, Seo E, Yoo RN, Sung H, Lee J. Clinical significance of viral-bacterial codetection among young children with respiratory tract infections: Findings of RSV, influenza, adenoviral infections. *Medicine (Baltimore).* 2020;**99**(2). e18504. doi: [10.1097/MD.00000000000018504](#). [PubMed: [31914021](#)]. [PubMed Central: [PMC6959858](#)].
  16. Crotty MP, Meyers S, Hampton N, Bledsoe S, Ritchie DJ, Buller RS, et al. Epidemiology, Co-Infections, and Outcomes of Viral Pneumonia in Adults: An Observational Cohort Study. *Medicine (Baltimore).* 2015;**94**(50). e2332. doi: [10.1097/MD.0000000000002332](#). [PubMed: [26683973](#)]. [PubMed Central: [PMC5058945](#)].
  17. Wei L, Liu W, Zhang XA, Liu EM, Wo Y, Cowling BJ, et al. Detection of viral and bacterial pathogens in hospitalized children with acute respiratory illnesses, Chongqing, 2009–2013. *Medicine (Baltimore).* 2015;**94**(16). e742. doi: [10.1097/MD.0000000000000742](#). [PubMed: [25906103](#)]. [PubMed Central: [PMC4602679](#)].
  18. Gambarino S, Mantovani S, Astegiano S, Libertucci D, Solidoro P, Baldi S, et al. Lower respiratory tract viral infections in hospitalized adult patients. *Minerva Med.* 2009;**100**(5):349–55. [PubMed: [19910888](#)].
  19. Astegiano S, Costa C, Terlizzi ME, Sidoti F, Gambarino S, Mantovani S, et al. Detection of human herpesvirus-7 DNA in bronchoalveolar lavage. *Intervirology.* 2010;**53**(2):119–23. doi: [10.1159/000264202](#). [PubMed: [19955817](#)].
  20. Avnon LS, Smolnikov A, Almog Y. Varicella pneumonia in southern Israel: clinical characteristics, diagnosis and therapeutic considerations. *Isr Med Assoc J.* 2009;**11**(5):261–5. [PubMed: [19637501](#)].
  21. Bonizzoli M, Arvia R, di Valvasone S, Liotta F, Zakrzewska K, Azzi A, et al. Human herpesviruses respiratory infections in patients with acute respiratory distress (ARDS). *Med Microbiol Immunol.* 2016;**205**(4):371–9. doi: [10.1007/s00430-016-0456-z](#). [PubMed: [27138606](#)]. [PubMed Central: [PMC7086591](#)].
  22. Tachikawa R, Tomii K, Seo R, Nagata K, Otsuka K, Nakagawa A, et al. Detection of herpes viruses by multiplex and real-time polymerase chain reaction in bronchoalveolar lavage fluid of patients with acute lung injury or acute respiratory distress syndrome. *Respiration.* 2014;**87**(4):279–86. doi: [10.1159/000355200](#). [PubMed: [24334877](#)].
  23. Chen CL, Huang Y, Martinez-Garcia MA, Yuan JJ, Li HM, de la Rosa-Carrillo D, et al. The role of Epstein-Barr virus in adults with bronchiectasis: a prospective cohort study. *Open Forum Infect Dis.* 2020;**7**(8):ofaa235. doi: [10.1093/ofid/ofaa235](#). [PubMed: [32766379](#)]. [PubMed Central: [PMC7397835](#)].
  24. McManus TE, Marley AM, Baxter N, Christie SN, Elborn JS, O'Neill HJ, et al. High levels of Epstein-Barr virus in COPD. *Eur Respir J.* 2008;**31**(6):1221–6. doi: [10.1183/09031936.00107507](#). [PubMed: [18287127](#)].
  25. Yamamoto K, Yoshikawa T, Okamoto S, Yamaki K, Shimokata K, Nishiyama Y. HHV-6 and 7 DNA loads in lung tissues collected from patients with interstitial pneumonia. *J Med Virol.* 2005;**75**(1):70–5. doi: [10.1002/jmv.20239](#). [PubMed: [15543584](#)].
  26. Johnson KH, Webb CH, Schmeling DO, Brundage RC, Balfour HJ. Epstein-Barr virus dynamics in asymptomatic immunocompetent adults: an intensive 6-month study. *Clin Transl Immunol.* 2016;**5**(5). e81. doi: [10.1038/cti.2016.28](#). [PubMed: [27350880](#)]. [PubMed Central: [PMC4910122](#)].
  27. Gleeson M, Pyne DB, Austin JP, Lynn Francis J, Clancy RL, McDonald WA, et al. Epstein-Barr virus reactivation and upper-respiratory illness in elite swimmers. *Med Sci Sports Exerc.* 2002;**34**(3):411–7. doi: [10.1097/00005768-200203000-00005](#). [PubMed: [11880803](#)].
  28. Ihira M, Yoshikawa T, Ohashi M, Enomono Y, Akimoto S, Suga S, et al. Variation of human herpesvirus 7 shedding in saliva. *J Infect Dis.* 2003;**188**(9):1352–4. doi: [10.1086/379040](#). [PubMed: [14593593](#)].
  29. Harvala H, Wolthers KC, Simmonds P. Parechoviruses in children: understanding a new infection. *Curr Opin Infect Dis.* 2010;**23**(3):224–30. doi: [10.1097/qco.0b013e32833890ca](#). [PubMed: [20414971](#)].
  30. Abed Y, Boivin G. Human parechovirus types 1, 2 and 3 infections in Canada. *Emerg Infect Dis.* 2006;**12**(6):969–75. doi: [10.3201/eid1206.051675](#). [PubMed: [16707054](#)]. [PubMed Central: [PMC3373051](#)].
  31. Hakansson A, Kidd A, Wadell G, Sabharwal H, Svanborg C. Adenovirus infection enhances in vitro adherence of *Streptococcus pneumoniae*. *Infect Immun.* 1994;**62**(7):2707–14. doi: [10.1128/iai.62.7.2707-2714.1994](#). [PubMed: [8005661](#)]. [PubMed Central: [PMC302872](#)].
  32. McCullers JA. Insights into the interaction between influenza virus and pneumococcus. *Clin Microbiol Rev.* 2006;**19**(3):571–82. doi: [10.1128/CMR.00058-05](#). [PubMed: [16847087](#)]. [PubMed Central: [PMC1539103](#)].
  33. Garcia-Garcia ML, Calvo C, Pozo F, Villadangos PA, Perez-Brena P, Casas I. Spectrum of respiratory viruses in children with community-acquired pneumonia. *Pediatr Infect Dis J.* 2012;**31**(8):808–13. doi: [10.1097/INF.0b013e3182568c67](#). [PubMed: [22531244](#)].
  34. Dowell SF, Anderson LJ, Gary HJ, Erdman DD, Plouffe JF, File TJ, et al. Respiratory syncytial virus is an important cause of community-acquired lower respiratory infection among hospitalized adults. *J Infect Dis.* 1996;**174**(3):456–62. doi: [10.1093/infdis/174.3.456](#). [PubMed: [8769600](#)].
  35. Brittain-Long R, Westin J, Olofsson S, Lindh M, Andersson LM. Prospective evaluation of a novel multiplex real-time PCR assay for detection of fifteen respiratory pathogens-duration of symptoms significantly affects detection rate. *J Clin Virol.* 2010;**47**(3):263–7. doi: [10.1016/j.jcv.2009.12.010](#). [PubMed: [20080440](#)]. [PubMed Central: [PMC7108433](#)].