Published Online: 2024 September 24



Mutations in the X Gene of the Hepatitis B Virus and Their Influence on Outcome CHB Infection in Three-Generations in the Family

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Received: 8 May, 2024; Revised: 23 July, 2024; Accepted: 3 August, 2024

Abstract

Background: The occurrence of specific mutations within the Hepatitis B virus (HBV) genome is associated with the progression of chronic hepatitis B infection towards more severe outcomes.

Objectives: This study aimed to investigate mutational patterns in the *X*-gene and their influence on the outcome of chronic HBV infection (CHB) across three generations in a family.

Methods: Ninety CHB patients, meeting the inclusion criteria, were recruited from cases referred to the Center of Hepatology at Golestan University of Medical Sciences between September 2020 and January 2021. The *HBx* gene was amplified using seminested PCR from serum samples and then subjected to sequencing.

Results: A comparison of the sequences from CHB patients indicated that children and mothers in the two-generation group exhibited the highest similarity (79.3%) in the *X*-gene, with the lowest mutation rate (20.7%). The N-terminal region of the *X*-gene showed the highest mutation frequency in the three-generation group, including C1491G (25%), G1613T (23.9%), C1500T (43.4%), and G1658T (33.4%). The mutation rate was notably higher in *HBeAg*-negative patients across the three groups compared to *HBe*-Ag-positive CHB patients, with a statistically significant difference (P = 0.03). A1762T/G1764A mutations were observed in 15.6% of patients, and their presence showed a significant difference (P = 0.03). Additionally, in the three-generation group, a silent mutation (A1727G, 10%) and a missense mutation (A1727T, 30%) were detected.

Conclusions: Specific mutational patterns in the *HBx* gene may be valuable in predicting clinical outcomes in CHB patients and could serve as warning indicators for increased susceptibility to hepatocellular carcinoma (HCC).

Keywords: Hepatitis B-virus, X Protein, Mutation, Three Generations Group

1. Background

Hepatitis B virus (HBV) is a major etiological agent of hepatitis globally (1), with chronic cases leading to endstage liver diseases such as hepatocellular carcinoma (HCC) and cirrhosis (2). The prevalence of HBV infection among the Iranian population is approximately 3%, and the progression of related liver disease is influenced by host, viral, and environmental factors (3). Mutations in the viral genome play a critical role in exacerbating the infection (4). Some genomic regions, such as the precore/basal core promoter (BCP), direct repeat sequences, and enhancer II (EnH II), overlap with the HBV X-gene (5). The *HBX* protein, an important non-structural protein with transcriptional transactivator capabilities, affects both cellular and viral promoters (6). It functions as a multifaceted oncoprotein related to HCC in chronic HBV patients (CHB) through a multistep process, which includes the induction of reactive oxygen species,

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alteration of mitochondrial function and physiology, and DNA damage (7, 8).

Different genetic variations in the X-gene, such as mutations during chronic infection (9), can impact not only the amino acid sequence but also HBV expression and the expression of other genes (10). These mutations can influence the physiological functions of the *HBX* protein, affecting biological mechanisms such as cell proliferation, transcription, apoptosis, and signal transduction, ultimately contributing to disease progression (11). The *HBX* protein is also able to create a cellular environment conducive to HBV replication by activating host genes associated with cell proliferation and inflammation (12).

In hepatic inflammation, *HBX* stimulates the transcription of numerous pro-inflammatory cytokines. The protease inhibitor-like structure of *HBX* may also cause the accumulation of toxic factors, resulting in severe hepatocellular injury (8, 13). Specific mutations such as AG1762/1764TA, C1653T, C1485T, T1753C, A1383C, and G1613A have been associated with HCC survival (14). The nonsynonymous C1653T mutation in the EnH II region may alter binding affinity, leading to an amino acid substitution in the *HBX* gene (15). The occurrence of the double mutation K130M + V131I increases as liver disease progresses and may contribute to HCC by affecting NF- κ B activity (16, 17).

2. Objectives

Recent studies have reported that *HBX* plays a crucial role in the progression and pathogenesis of HBV-related complications. Therefore, the aim of this study, similar to our previous research on S-gene mutations in HBV, was to analyze and detect *X*-gene mutational patterns in patients across three generations.

3. Methods

3.1. Baseline Demographic Features of the Study Population

In this cross-sectional study conducted in the northeastern region of Iran, from September 2020 to January 2021, at the Research Center of Gastroenterology and Hepatology, Golestan University of Medical Sciences, 3,250 HBV-infected patients were investigated based on the inclusion criteria. Samples were collected from patients with CHB, and plasma was separated from the blood and stored. All participants were tested for alanine transaminase (ALT), aspartate transaminase (AST), and alkaline phosphatase (ALP) levels (18). Ninety patients with confirmed HBV infection (positive *HBs-Ag*, history of HBV infection for more than six months) were included in the study. Patients were divided into three groups: Three-generations group (30 cases), twogenerations group (40 cases), and intrafamilial group (20 cases), based on the inclusion criteria (*HBs-Ag*: Positive, *HBs-Ab*: Negative, *HBc-Ab*: Positive, *HBe-Ag* and *HBe-Ab*: Positive/negative). Informed consent was obtained from each participant after a thorough interview explaining the study goals and answering any questions. Participation in the study was voluntary and had no impact on the treatment course of patients.

3.2. Extraction of Hepatitis B Virus DNA and Semi-Nested PCR

The HBV-DNA extraction was performed from 200 µL of serum following the manufacturer's instructions (Viral Nucleic Acid Extraction Kit, Yektatajhiz). Amplification of HBV sequences covering nucleotides 1365 - 2078 was conducted using F1/R1 primers. In the second round, a 713 bp template was used to amplify nucleotides 1365 - 1881 with F1/R2 primers (Table 1) (4).

3.3. DNA Sequencing and Mutation Analysis

PCR products that yielded positive results were subjected to automated directional sequencing (3130 Genetic Analyzer, *ABI/HITACHI*, Tehran University of Medical Sciences). Mutation identification and analysis were performed by aligning the nucleotide sequences with the standard sequence of hepatitis B (accession number: AB033559) from the GenBank database. Statistical analysis was conducted using SPSS version 20. The student's *t*-test was used to compare the collected data, with a P-value of less than 0.05 considered statistically significant for differences between categories.

4. Results

Investigation of the *HBx* gene through sequencing of 90 samples from patients (accession numbers: ON346437-ON346526) was conducted. In contrast to the other groups, liver function test results in the threegeneration group showed a significant association with values above the normal range. The study found that in the intra-familial group and the two-generation group, liver function test levels were elevated in *HBe*-Ag-positive patients, with statistical significance observed in the two-generation group (P-value = 0.08). In the third

Table 1. Oligonucleotide Primers Used for Semi-nested PCR and Sequencing of X-region of Hepatitis B						
Primer Name	Sequence (5' to 3')	Target Sequence	PCR Program			
F1	ATCGTATCCATGGCTGCTAGGCT	1365 - 1387	Step 1: 94°C 5 min, 35 cycles (94°C 1 min, 55°C 1 min and 72°C 1 min), 72°C 7 min; Step 2: 94°C 5 min, 35 cycles (94°C 30 Sec, 57.5°C 30 Sec and 72°C 45 Sec), 72°C 5 min.			
R1	CAGAATAGCTTGCCTGAGTGC	2058 - 2078				
R2	CACAGCTTGGAGGCTTGAACA	1861 - 1881				

 Table 2. Main Characteristics of Chronic Hepatitis B Virus Patients in Three Groups ^a

Basic Characteristics	Three Generations; (Grandmother-Mother and Child)	Two Generations; (Mother and Child)	Intrafamilial Members
Age	45.23 ± 22.8	35.83±17.7	35.5 ± 15.2
Gender			
Male	4 (13.4)	11 (27.5)	8(40)
Female	26 (86.6)	29 (72.5)	12 (60)
Total	30	40	20
HBe -Ag			
+	0(0)	7 (17.5)	13 (65)
-	30 (100)	33 (82.5)	7(35)
Total	30	40	20
Anti- <i>HBe</i>			
+	30 (100)	4 (10)	16 (80)
-	0(0)	36 (90)	4 (20)
Total	30	40	20
ALT (IU/L)	24.6±14.37	24 ± 8.9	22 ± 7.9
AST (IU/L)	25.46±13.67	26±11.2	23.3 ± 10.2
ALP (IU/L)	419.567±259.78	464.75 ± 274.1	337.6 ± 172.2

Abbreviations: IU/L, international unit/litre; ALT, alanine transaminase; AST, aspartate transaminase; ALP, alkaline phosphatase. ^a Values are expressed as mean ± SD or No. (%).

group, there was no significant correlation between liver function test levels, gender (P-value = 0.5), and age (P-value = 0.06) (Table 2).

4.1. HBx Mutations Concerning the Status of Hepatitis B Virus Infection

When the sequences of the HBV partial genome isolated from CHB patients were compared with the reference sequence, it was found that 9.47% of CHB patients did not have a mutation in the X-gene (3.4% in the first group, 15% in the second group, and 10% in the third group). The sequence analysis of the X-gene in these patients demonstrated the highest degree of homology (79.3%) and the lowest mutation rate (20.7%) in mothers and children, respectively. Additionally, the results showed that grandmothers in the three-generation group had the most mutations in the X-gene region, which was statistically significant (P-value = 0.06).

This study identified the co-occurrence of A1762T/G1764A mutations in 6.7% of the three-generation group, 20% of the two-generation group, and 20% of the intra-familial group. Statistical analysis revealed a significant difference (P-value = 0.03) in the cooccurrence of G1764A/A1762T mutations among these distinct groups. Co-occurrence of C1766G/G1764T mutations was observed in 13.4%, 15%, and 10% of CHB patients in the three groups, respectively (P-value = 0.4). The presence of T1464C was detected in 30% of CHB patients, with a higher prevalence in the intra-familial group (35%) compared to the two-generation (25%) and three-generation groups (33.4%) (P-value = 0.02). Additionally, the double mutation G1479A/C1481T was found in 6.7%, 10%, and 15% of CHB patients in the three groups, respectively (P-value = 0.04).

The C1500T point mutation was detected in 40% of CHB patients, with the highest frequency observed in the three-generation group (43.4%) compared to the

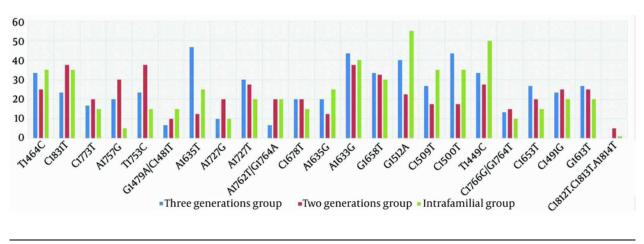


Figure 1. The frequency of nucleotide points mutations in the X gene in chronic hepatitis B virus (CHB) patients

other two groups (P-value = 0.03). The three-generation group also exhibited significantly higher frequencies of A1635T (46.7%) and A1635G (20%) point mutations compared to the other two groups (P-value = 0.01 and P-value = 0.04, respectively). Statistical analysis did not reveal a significant relationship between *HBx* mutations and elevated LFT or age (P-value = 0.1) (Figure 1).

The Hepatitis B virus DNA sequence analysis of the three-generation and two-generation groups revealed silent mutations A1727G (10% and 20%, respectively) and missense mutation A1727T (30% and 27.5%, respectively) (P-value = 0.04, P-value = 0.03, respectively).Additionally, a triplet mutation, A1762T/G1764A/C1773T, was detected in CHB patients. This study also identified specifically another triplet mutation, C1812T/C1813T/A1814T, which showed lower prevalence at 5% and 1% in the two-generation and intra-familial groups, respectively. However, no statistically significant difference was observed (P-value = 0.3) in the frequency of these mutations among the groups.

The B-cell epitope of the *HBx* protein showed two types of mutations: C1491G and C1500T. In the two- and three-generation groups, the prevalence of these mutations was 25% and 43.4%, respectively. The three-generation group also exhibited the highest abundance of mutations overall (33.4%) compared to the other groups. The G1613T mutation, which causes the substitution of Glu80 with Asp, was reported in 23.9% of the study population. Despite the comparable mutation frequency in both groups, statistical analysis did not reveal any significant difference (P-value = 0.2).

According to our findings, the A1633G (43.4%), C1500T (43.4%), and A1635T (46.7%) mutations were the most abundant in the three-generation group compared to the other two groups.

4.2. Mutations in X Region and its Effect on HBe-Ag Expression

There was a statistically significant difference between CHB patients who tested positive for *HBe*-Ag and those who tested negative (P-value = 0.03). Among the 90 patients, only 11 (12.3%) tested positive for *HBe*-Ag. All CHB patients in the three-generation cohort tested negative for *HBe*-Ag. Additionally, it was observed that these patients had the highest number of mutations in the X region of the HBV genome.

5. Discussion

Detection of *HBx* in patients with HCC is a welldocumented phenomenon often linked to mutations that contribute to HBV infection development (19, 20). *HBx* proteins lack the C domain, which is crucial for their suppressive effects on cell proliferation, growth, transactivation activity, and transformation (21, 22). In 2.9% of CHB cases, 8bp deletions or insertions in the Cterminus of *HBx* were found in the cirrhotic group. Moreover, 15 different deletions, such as 1769 - 1773, 1762 -1768, 1763 - 1770, and T1771/A1775, were identified in cirrhotic patients (23). In contrast to previous research, this study found that CHB patients did not demonstrate any deletion-related mutations, except for C1773T, which had a 20% prevalence in the two-generation group. Our results align with previous studies (23, 24), showing that *HBe*-Ag-negative patients are more likely to have deletion and insertion mutations in the C-terminus region of *HBx*.

Another study demonstrated that the A1762T/G1764A mutation in the C-terminal overlap region of the X-gene with BCP contributes to the progression of the disease from the chronic phase to cirrhosis. Salarnia et al. showed that the A1762T/G1764A mutation was more common in patients with cirrhosis than in those with CHB (4). The presence of this mutation plays a significant role in the advancement of liver disease to more critical stages (4). Our findings are consistent with those of previous studies, including Chen et al. (16), Vazjalali et al., and Maleki et al. (25, 26), which showed that an increased prevalence of A1762T/G1764A mutations contributes to disease progression. This mutation accelerates viral replication, and our results indicate it is more frequent in *HBe*-Ag-negative patients in the two-generation group. We also found that the C1773T point mutation coincided with the A1762T/G1764A double mutation, though neither was statistically significant.

A study by Salarnia et al. identified new mutations within the *HBx* gene. Various mutations, including C1500T, C1491G, G1658T, and G1613T, were observed in the N-terminal region, Box α, Core promoter, and Enhancer II (6). Consistent with these findings, novel mutations were detected in CHB patients across the threegeneration, two-generation, and intra-familial groups. The three-generation group exhibited the highest mutation frequency, with A1635T (46.7%), A1633G (43.4%), C1500T (43.4%), and C1491G (23.45%) being the most commonly reported. The occurrence of the A1635T mutation in the HBx protein sequence results in interaction with the DNA damage-binding complex-1 due to the overlap between the NRE and *HBx* coding sequences. A study by Ghosh et al. demonstrated a higher prevalence of A1635T among patients with cirrhosis (53.85%) compared to inactive HBV carriers and those with chronic HBV (27). In contrast to our results, this mutation was observed in CHB patients in the threegeneration group.

Several studies have demonstrated that the A1727T mutation is a novel predictive marker for the cirrhosis phase in HBV-infected patients (6, 28). Patients with cirrhosis show a higher occurrence of the A1727G

mutation, which increases the risk of HCC (29). The majority of TA1 mutations occur in the 1750-1755 nt region, which is recognized as a potential prognosticator of HCC (30). Additionally, the T1753C mutation has been identified as an important marker for cirrhosis severity and is associated with advanced liver disease (31). Consistent with these studies, we found that the A1727G mutation was present in 14.5% of CHB cases, with a statistically significant difference among the three groups (P-value = 0.07). The T1753C mutation, a key prognosticator of cirrhosis, was found in 23.4% of the three-generation group, 37.5% of the two-generation group, and 15% of the intra-family group.

It has been observed that specific patterns of HBx mutations can serve as early markers for an increased risk of HCC and predict clinical outcomes in HBVinfected patients. Prior research, such as that by Xiao et al., demonstrated that mutations in the X-region tend to emerge during the advanced phases of chronic HBV infection, leading to serious liver disorders such as HCC and cirrhosis (32). Our findings indicate that CHB patients exhibit significant occurrences of the double mutations C1481T and G1479A, as well as the point mutations A1635T, T1464C, and C1500T. Triple mutations, specifically A1762T/G1764A/C1773T and C1812/C1813T/A1814T, were observed in the two-generation and three-generation groups.

Furthermore, the absence of statistically significant differences in age and elevated LFT levels among individuals with HBx mutations is consistent with previous studies. The C1481T/G1479A mutations were found in all three patient categories, with a higher frequency observed in the two-generation and intrafamilial groups compared to the three-generation group. Additionally, these mutations were more frequent in mothers than in their children and grandmothers, with this difference approaching statistical significance (P-value = 0.06). Our ability to comprehensively investigate disease progression and potential associations with combinations of HBV mutations is limited by the relatively small sample size of cases with progressive liver disease. We acknowledge that our findings may not be generalizable to other populations, particularly those infected with different HBV genotypes.

5.1. Conclusions

In conclusion, identifying and analyzing viral genomic mutations in correlation with clinical complications is crucial for disease management, prognosis, and treatment. These mutations can be used to screen high-risk individuals for liver disease, improve diagnostic methods, and refine therapeutic approaches. Additionally, further investigation into the impact of A1762T/G1764A mutations on different phases of HBV infection is recommended.

Acknowledgements

The present study was derived from a research project for completion of Ph.D. course Shahid Beheshti University and Golestan University of Medical Sciences.

Footnotes

Authors' Contribution: A. M., S. M. H., I. Sh., and S. B.: Contributed to study conception; A. M., N. B., M. N., and V. H.: Contributed to data analysis; M. N.: Performing the experiments; A. M., S. M. H., and M. N.; revision: A. M. and M. N.: Reading and confirming the final version of the manuscript.

Conflict of Interests Statement: All authors approve of this article, and there are no conflicts of interest.

Data Availability: The nucleotide sequences of HBx gene to identify mutations in 90 CHB patient samples in this study were deposited in the GenBank database under accession numbers ON346437 - ON346526. Also, all relevant data are within the manuscript.

Ethical Approval: The ethical committee of Golestan University of Medical Sciences granted approval for the study (IR. GOUMS.REC.1399.105).

Funding/Support: This research received no specific grant during the preparation of this manuscript.

Informed Consent: All study participants completed the informed consent form.

References

 Gong DY, Chen EQ, Huang FJ, Leng XH, Cheng X, Tang H. Role and functional domain of hepatitis B virus X protein in regulating HBV transcription and replication in vitro and in vivo. *Viruses*. 2013;5(5):1261-71. [PubMed ID: 23698398]. [PubMed Central ID: PMC3712307]. https://doi.org/10.3390/v5051261.

- Naderi M, Hosseini SM, Besharat S, Behnampour N, Shahramian I, Moradi A. Clinical and virological aspects of core and pre-core mutations in three generations of chronic hepatitis B virus patients. *Future Virology.* 2023;**18**(6):349-58. [PubMed ID: 36592642]. https://doi.org/10.2217/fvl-2022-0216.
- Salehi-Vaziri M, Sadeghi F, Almasi Hashiani A, Gholami Fesharaki M, Alavian SM. Hepatitis B virus infection in the general population of Iran: An updated systematic review and meta-analysis. *Hepat Mon.* 2016;16(4). e35577. [PubMed ID: 27257428]. [PubMed Central ID: PMC4888501]. https://doi.org/10.5812/hepatmon.35577.
- Salarneia F, Zhand S, Khodabakhshi B, Tabarraei A, Vakili MA, Javid N, et al. Mutations at nucleotide 1762, 1764 and 1766 of hepatitis B virus X gene in patients with chronic hepatitis B and hepatitis B-related cirrhosis. *Med Lab J.* 2016;10(1):31-5. https://doi.org/10.18869/acadpub.mlj.10.1.31.
- Datta S, Chatterjee S, Veer V, Chakravarty R. Molecular biology of the hepatitis B virus for clinicians. J Clin Exp Hepatol. 2012;2(4):353-65. [PubMed ID: 25755457]. [PubMed Central ID: PMC3940099]. https://doi.org/10.1016/j.jceh.2012.10.003.
- Salarnia F, Behboudi E, Shahramian I, Moradi A. Novel X gene point mutations in chronic hepatitis B and HBV related cirrhotic patients. *Infect Genet Evol.* 2022;97:105186. [PubMed ID: 34920100]. https://doi.org/10.1016/j.meegid.2021.105186.
- Schollmeier A, Glitscher M, Hildt E. Relevance of HBx for hepatitis B virus-associated pathogenesis. *Int J Mol Sci.* 2023;24(5). [PubMed ID: 36902395]. [PubMed Central ID: PMC10003785]. https://doi.org/10.3390/ijms24054964.
- Cho EY, Choi CS, Cho JH, Kim HC. Association between Hepatitis B Virus X gene mutations and clinical status in patients with chronic hepatitis B infection. *Gut Liver*. 2011;5(1):70-6. [PubMed ID: 21461076]. [PubMed Central ID: PMC3065097]. https://doi.org/10.5009/gnl.2011.5.1.70.
- Song BC, Cui XJ, Kim HU, Cho YK. Sequential accumulation of the basal core promoter and the precore mutations in the progression of hepatitis B virus-related chronic liver disease. *Intervirol.* 2006;49(5):266-73. [PubMed ID: 16714855]. https://doi.org/10.1159/000093456.
- Kreutz C. Molecular, immunological and clinical properties of mutated hepatitis B viruses. J Cell Mol Med. 2002;6(1):113-43. [PubMed ID: 12003675]. [PubMed Central ID: PMC6740305]. https://doi.org/10.1111/j.1582-4934.2002.tb00317.x.
- Zhang X, Ding HG. Key role of hepatitis B virus mutation in chronic hepatitis B development to hepatocellular carcinoma. *World J Hepatol.* 2015;7(9):1282-6. [PubMed ID: 26019744]. [PubMed Central ID: PMC4438503]. https://doi.org/10.4254/wjh.v7.i9.1282.
- Lee SA, Mun HS, Kim H, Lee HK, Kim BJ, Hwang ES, et al. Naturally occurring hepatitis B virus X deletions and insertions among Korean chronic patients. *J Med Virol*. 2011;83(1):65-70. [PubMed ID: 21108340]. https://doi.org/10.1002/jmv.21938.
- Koike K. Hepatitis B virus X gene is implicated in liver carcinogenesis. *Cancer Lett.* 2009;**286**(1):60-8. [PubMed ID: 19464104]. https://doi.org/10.1016/j.canlet.2009.04.010.
- Zhang C, Xie Y, Lai R, Wu J, Guo Z. Nonsynonymous C1653T mutation of hepatitis B virus X gene enhances malignancy of hepatocellular carcinoma cells. J Hepatocell Carcinoma. 2022;9:367-77. [PubMed ID: 35535232]. [PubMed Central ID: PMC9078866]. https://doi.org/10.2147/JHC.S348690.
- 15. Li SK, Ho SF, Tsui KW, Fung KP, Waye MY. Identification of functionally important amino acid residues in the mitochondria targeting

sequence of hepatitis B virus X protein. Virol. 2008;**381**(1):81-8. [PubMed ID: 18805561]. https://doi.org/10.1016/j.virol.2008.07.037.

- Chen CH, Lee CM, Lu SN, Changchien CS, Eng HL, Huang CM, et al. Clinical significance of hepatitis B virus (HBV) genotypes and precore and core promoter mutations affecting HBV e antigen expression in Taiwan. *J Clin Microbiol.* 2005;43(12):6000-6. [PubMed ID: 16333089]. [PubMed Central ID: PMC1317177]. https://doi.org/10.1128/ICM.43.12.6000-6006.2005.
- Lee JH, Han KH, Lee JM, Park JH, Kim HS. Impact of hepatitis B virus (HBV) x gene mutations on hepatocellular carcinoma development in chronic HBV infection. *Clin Vaccine Immunol.* 2011;**18**(6):914-21. [PubMed ID: 21490166]. [PubMed Central ID: PMC3122615]. https://doi.org/10.1128/CVI.00474-10.
- Naderi M, Hosseini SM, Behnampour N, Shahramian I, Moradi A. Mutations in the S gene of hepatitis B virus in three generations of patients with chronic hepatitis B. Virus Genes. 2023;59(5):662-9. [PubMed ID: 37308753]. https://doi.org/10.1007/s11262-023-02012-z.
- Lazarevic I. Clinical implications of hepatitis B virus mutations: recent advances. World J Gastroenterol. 2014;20(24):7653-64. [PubMed ID: 24976703]. [PubMed Central ID: PMC4069294]. https://doi.org/10.3748/wjg.v20.i24.7653.
- Bouchard MJ, Schneider RJ. The enigmatic X gene of hepatitis B virus. *J Virol*. 2004;**78**(23):12725-34. [PubMed ID: 15542625]. [PubMed Central ID: PMC524990]. https://doi.org/10.1128/JVI.78.23.12725-12734.2004.
- Mukherji A, Janbandhu VC, Kumar V. HBx-dependent cell cycle deregulation involves interaction with cyclin E/A-cdk2 complex and destabilization of p27Kip1. *Biochem J.* 2007;**401**(1):247-56. [PubMed ID: 16939421]. [PubMed Central ID: PMC1698683]. https://doi.org/10.1042/BJ20061091.
- Ma NF, Lau SH, Hu L, Xie D, Wu J, Yang J, et al. COOH-terminal truncated HBV X protein plays key role in hepatocarcinogenesis. *Clin Cancer Res.* 2008;14(16):5061-8. [PubMed ID: 18698024]. https://doi.org/10.1158/1078-0432.CCR-07-5082.
- Salarnia F, Besharat S, Zhand S, Javid N, Khodabakhshi B, Moradi A. Mutations in hepatitis-B X-gene region: Chronic hepatitis-B versus cirrhosis. J Clin Diagn Res. 2017;11(3):OC31-4. [PubMed ID: 28511432].
 [PubMed Central ID: PMC5427358]. https://doi.org/10.7860/[CDR/2017/22570.9498.
- 24. Fujiwara K, Tanaka Y, Paulon E, Orito E, Sugiyama M, Ito K, et al. Novel type of hepatitis B virus mutation: Replacement mutation involving a hepatocyte nuclear factor 1 binding site tandem repeat in chronic hepatitis B virus genotype E. J Virol. 2005;**79**(22):14404-10. [PubMed

ID: 16254374]. [PubMed Central ID: PMC1280239]. https://doi.org/10.1128/JVI.79.22.14404-14410.2005.

- Vaezjalali M, Rezaee H, Goudarzi H. HBV S gene premature stop codon in strains from Middle Eastern patients. Arch Clin Infect Dis. 2012;8(1):3-7. https://doi.org/10.5812/archcid.14410.
- Malik A, Singhal DK, Albanyan A, Husain SA, Kar P. Hepatitis B virus gene mutations in liver diseases: A report from New Delhi. *PLoS One*. 2012;7(6). e39028. [PubMed ID: 22720023]. [PubMed Central ID: PMC3375258]. https://doi.org/10.1371/journal.pone.0039028.
- Ghosh S, Mondal RK, Banerjee P, Nandi M, Sarkar S, Das K, et al. Tracking the naturally occurring mutations across the full-length genome of hepatitis B virus of genotype D in different phases of chronic e-antigen-negative infection. *Clin Microbiol Infect.* 2012;18(10):E412-8. [PubMed ID: 22827722]. https://doi.org/10.1111/j.1469-0691.2012.03975.x.
- Yin J, Xie J, Liu S, Zhang H, Han L, Lu W, et al. Association between the various mutations in viral core promoter region to different stages of hepatitis B, ranging of asymptomatic carrier state to hepatocellular carcinoma. *Am J Gastroenterol.* 2011;106(1):81-92. [PubMed ID: 20959817]. https://doi.org/10.1038/ajg.2010.399.
- 29. Khan A, Al Balwi MA, Tanaka Y, Hajeer A, Sanai FM, Al Abdulkarim I, et al. Novel point mutations and mutational complexes in the enhancer II, core promoter and precore regions of hepatitis B virus genotype D1 associated with hepatocellular carcinoma in Saudi Arabia. *Int J Cancer.* 2013;**133**(12):2864-71. [PubMed ID: 23740667]. https://doi.org/10.1002/ijc.28307.
- Lee D, Lyu H, Chung YH, Kim JA, Mathews P, Jaffee E, et al. Genomic change in hepatitis B virus associated with development of hepatocellular carcinoma. *World J Gastroenterol*. 2016;**22**(23):5393-9.
 [PubMed ID: 27340355]. [PubMed Central ID: PMC4910660]. https://doi.org/10.3748/wjg.v22.i23.5393.
- Biswas A, Banerjee A, Chandra PK, Datta S, Panigrahi R, Dutta D, et al. Variations in the functional domain of basal core promoter of hepatitis B virus among Eastern Indian patients with prevalence of genotypes A, C, and D among the same ethnic population. *J Med Virol.* 2011;83(2):253-60. [PubMed ID: 21181919]. https://doi.org/10.1002/jmv.21979.
- Xiao L, Zhou B, Gao H, Ma S, Yang G, Xu M, et al. Hepatitis B virus genotype B with G1896A and A1762T/G1764A mutations is associated with hepatitis B related acute-on-chronic liver failure. *J Med Virol.* 2011;83(9):1544-50. [PubMed ID: 21739444]. https://doi.org/10.1002/jmv.22159.