



Trends in *Mycobacterium tuberculosis* Transmission During a 10-year Period (2006-2016) in the Northwest of Iran by MIRU-VNTR Molecular Typing

Mohammad Hossein Soroush Barhaghi ^{1,2}, Sepehr Taghizadeh ³, Peyvand Kashi¹, Mohammad Asgharzadeh ⁴, Poursya Gholizadeh ⁵, Khudaverdi Ganbarov ⁶, Asghar Tanomand ⁷, Milad Bastami ⁴, Seyyed Reza Moaddab ⁸, Behrooz Shokouhi ⁹ and Hossein Samadi Kafil ^{10,*}

¹Department of Medical Microbiology, Faculty of Medicine, Tabriz University of Medical Sciences, Tabriz, Iran

²Hematology and Oncology Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

³Department of Infectious Diseases, Tabriz University of Medical Sciences, Tabriz, Iran

⁴Biotechnology Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

⁵Student Research Committee, Tabriz University of Medical Sciences, Tabriz, Iran

⁶Department of Microbiology, Baku State University, Baku, Azerbaijan

⁷Department of Microbiology, Maragheh University of Medical Sciences, Maragheh, Iran

⁸Faculty of Paramedical, Tabriz University of Medical Sciences, Tabriz, Iran

⁹Stem Cell Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

¹⁰Drug Applied Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

*Corresponding author: Drug Applied Research Center, Tabriz University of Medical Sciences, Tabriz, Iran. Email: kafilhs@tbzmed.ac.ir

Received 2019 July 01; Revised 2020 February 08; Accepted 2020 February 10.

Abstract

Background: *Mycobacterium tuberculosis* is one of the biggest health challenges all over the world. The Caucasian region is one of the places with a high prevalence of drug-resistant *M. tuberculosis*.

Objectives: In this study, we aimed to investigate the trends in *M. tuberculosis* transmission during a 10-year period in the northwest of Iran.

Methods: We collected 166 *M. tuberculosis* isolates from 2005 to 2006 and 119 *M. tuberculosis* isolates from 2015 to 2016 and subjected them to MIRU-VNTR and ETR-VNTR typing by a polymerase chain reaction and compared them by phylogenetic tools.

Results: In the 2006 isolates, 104 different patterns were observed including 75 unique patterns and 91 isolates were clustered in 29 different clusters. In the 2016 isolates, 98 different patterns were observed with 86 unique patterns and 33 isolates were clustered in 12 different clusters. One cluster had a shared member from the 2006 and 2016 isolates, indicating the transmission of a single isolate during these years. The minimum estimate for the tuberculosis proportion, which is due to the recent transmission of tuberculosis, was 36.7% for the 2006 isolates and 17.6% for the 2016 isolates.

Conclusions: In spite of the reduced number of tuberculosis patients, the isolates were more resistant and had a close relationship with worldwide strains. Cross-border immigration for treatment from Republic of Azerbaijan had significant participation in the recent transmission of tuberculosis in this region. In conclusion, the strict control of patients commuting and developing new tuberculosis clinics inside Republic of Azerbaijan can play a key role in the control of tuberculosis transmission in the northwest of Iran.

Keywords: *Mycobacterium tuberculosis*, Genotyping, Transmission, Infection

1. Background

Tuberculosis is a bacterial disease caused by *Mycobacterium tuberculosis*, which often affects the lungs but can affect other parts of the body (1). It is one of the biggest health challenges all over the world, particularly in developing countries, which have large unregulated private sectors and weak health systems (2). The number of 10.4 million new cases was estimated according to the WHO report in 2016 around the world, 90% of which were in adults, 65% in males, 35% in females, and 56% in five countries includ-

ing India, Indonesia, China, Philippines, and Pakistan (3). Tabriz is located in the northwest of Iran neighboring with the Republic of Azerbaijan that is one of the countries with a high rate of multidrug-resistant tuberculosis (4). The transmission route of tuberculosis is from person to person through the air. The identification of the transmission routes and sources of *M. tuberculosis* is associated with prohibiting the generation of infectious particles, prevention programs, controlling the disease, and reducing or eliminating tuberculosis (5).

The low rate of tuberculosis transmission among the population is the characteristic of most low-incidence countries, but cross-border immigration from high-incidence countries can affect the incidence and features of tuberculosis in the low-incidence countries (6). The increased migration of high-burden tuberculosis countries such as Afghanistan and the Republic of Azerbaijan has resulted in the increased frequency and resistance of tuberculosis in low-burden cities of Iran as a low-incidence country (7, 8). Genotyping approaches of *M. tuberculosis* are used for epidemiological study, which include DNA fingerprinting methods such as Insertion Sequence 6110-restriction fragment length polymorphism (IS6110-RFLP), variable number of tandem repeats (VNTRs) based on mycobacterial interspersed repetitive units (MIRU-VNTR), exact tandem repeat loci-VNTR (ETR-VNTR), whole genome sequencing (WGS), and single nucleotide polymorphisms (SNPs) (9-13). These genotyping approaches are widely used in the control programs of tuberculosis to determine possible epidemiological links between patients, virulence patterns, and dynamics of transmission and to distinguish between relapse and re-infection (13).

Among the various genotyping approaches, IS6110-RFLP is the “Gold standard” approach, because it has high discriminatory power. However, IS6110-RFLP is a less reliable indicator of clonality for low-copy-number strains, particularly those with fewer than six copies of IS6110, and its profiles are difficult to compare across the results of laboratories (14, 15). The MIRU-VNTR approach investigates the number of repeated mycobacterial interspersed repetitive units based on polymerase chain reaction (PCR) at 12, 15, and 24 loci (11, 16). Mycobacterial interspersed repetitive units are composed of 40 - 100 bp scattered repetitive sequences over 41 locations of entire the chromosome of *M. tuberculosis* (11). The discriminating power of the MIRU-VNTR typing approach is generally associated with the increasing number and sets of loci used (17, 18). Moreover, the MIRU-VNTR approach is useful for studying at the relatively high resolution of clonal expansion and diversity of particular strains or lineages (19, 20). In addition, ETRs are three or five tandem repeats that are combined by 12 and 24 loci MIRU-VNTR for genotype family classification of *M. tuberculosis* strains and have less discriminatory power than IS6110 RFLP (21, 22).

2. Objectives

In the present study, we aimed to compare the association of two groups of isolates collected from tuberculosis patients referring to the East-Azarbayjan Center for Tuberculosis in 2006 and 2016 by MIRU-VNTR and ETR-VNTR

typing methods. Moreover, we assessed the historical connection among strains and predicted future outbreaks or cluster investigation.

3. Methods

From February 2005 to February 2006, 166 *M. tuberculosis* isolates were collected from the East Azarbaijan Center for Tuberculosis. From 2015 to 2016, 119 *M. tuberculosis* isolates were collected from patients with tuberculosis in the East Azarbaijan Tuberculosis Center. Demographic data of patients were collected by questionnaires filled out by patient interviews or based on their documents. All samples were approved as *M. tuberculosis* isolates by conventional biochemical tests based on catalase, niacin, nitrate reduction, and pigment production of isolates. All isolates were analyzed for antibiotic sensitivity patterns by the proportional method using isoniazid, streptomycin, rifampin, and ethambutol (Hortel, Spain) (23).

We conducted DNA extraction by the Phenol-Chloroform method as previously described (24). All the extracted DNAs were stored at -70°C for further analysis. For molecular analysis, we used the 12-loci MIRU-VNTR method plus ETR-VNTR (22, 25). The Supply et al. protocol was used for MIRU-VNTR typing of the isolates (26). In this regard, 12 selected loci were amplified by the primer-specific PCR method. All reactions were done in 20 μ L volumes using the Takara master mix (Takara, Japan). The thermal protocol included 94°C for 7 min, followed by 35 cycles of 94°C for 45 s, and different temperatures for annealing at 45°C and 72°C for 55 s. The annealing temperatures were as follows: loci 2 (65°C), 4 (63°C), 10 (68°C), 16 (65°C), 20 (59°C), 23 (67°C), 24 (59°C), 26 (65°C), 27 (64°C), 31 (63°C), 39 (68°C), 40 (65°C), ETR-A (66°C), ETR-B (68°C), and ETR-C (69°C). Electrophoresis of the PCR products was done on the 1.5% agarose gel, stained with ethidium bromide (Merck, Germany), and visualized by UV light in a Manual Gel Documentation System (inGenius3, UK). All sizes of fragments were determined by comparing them with a 50-bp DNA ladder size marker (Yekta Tajhiz, Iran) (26). *Mycobacterium tuberculosis* PT71 was provided by the Tabriz reference tuberculosis lab as the reference strain and used as external quality control for consistency of the results of MIRU and ETR.

3.1. Statistical Analysis

The phylogenic analysis of the isolates was done using Multi-variable Statistical Package (MVSP) software and MIRU-VNTRplus free web-based software (available at <https://www.miru-vntrplus.org/MIRU/index.faces>). The Neighbor-joining (NJ) algorithm was used for all comparisons. The categorical distance was used for the analysis of

our data (27). A cluster in this study was defined as two or more isolates with identical MIRU-VNTR and ETR patterns while isolates with different patterns in any loci were considered non-clustered isolates. Clusters were assumed to be the result of recent transmission. The chi-square test was used for statistical analysis and p values of < 0.05 were considered significant. The minimum estimation of recent transmission of tuberculosis was calculated by the following formula: (number of clustered isolates-number of clusters)/number of all isolates.

4. Results

4.1. Sample Collection in 2006

In 2006, 197 isolates of *M. tuberculosis* were collected. Thirty-one isolates had not acceptable DNA for molecular analysis; thus, 166 isolates were analyzed by the MIRU-VNTR method. Ninety isolates (54.27%) were from males and 76 (45.78%) from females and the age of the patients ranged from 2 to 88 years. There was no significant difference in risk factors between clustered and non-clustered patients ($P > 0.05$). In the MIRU-ETR analysis of these isolates, 104 different patterns were observed including 75 unique patterns and 29 shared patterns with several isolates; 91 isolates were clustered in 29 different clusters. There were 29 isolates from Urmia and 61 isolates from Tabriz in the clusters while no patient isolate from Republic of Azerbaijan was in the clusters. We observed 17 clusters with two members, three clusters with three members, three clusters with four members, one cluster with five members, four clusters with six members, and one cluster with seven members. The biggest cluster included six isolates from Tabriz and one from Urmia.

4.2. Sample Collection in 2016

In 2016, 119 isolates of *M. tuberculosis* were isolated. All the isolates had enough DNA for further molecular typing. Fifty-eight (48.73%) isolates were from males and 61 (51.26%) from females and the age of patients ranged from 15 to 86 years. Risk factors had no difference between clustered and non-clustered patients ($P > 0.05$). Twenty-nine isolates were from Republic of Azerbaijan. Eight isolates from Azerbaijan were resistant to at least two antibiotics (five isolates to three antibiotics and three isolates to two antibiotics) and only one isolate from Tabriz was resistant to at least two antibiotics. Only these isolates had resistance to antibiotics. No resistance to streptomycin was observed (Appendix 2 in Supplementary File). Ninety-eight different patterns were observed with 86 unique patterns and 12 shared patterns clustered in 12 different clusters including 33 isolates. Nine clusters had two members, one

cluster had four members, one cluster had five members, and one cluster had six members. Patients from Republic of Azerbaijan were included in four clusters that indicated the transmission of tuberculosis among these patients. The biggest cluster included one patient from Republic of Azerbaijan and five patients from Tabriz. Other clusters with patients from Azerbaijan were a cluster with three patients from Republic of Azerbaijan and one from Tabriz and a cluster with one patient from Republic of Azerbaijan and one from Tabriz. In addition, a cluster with only two patients from Republic of Azerbaijan was observed.

In addition, only three isolates from the 2016 isolates had a close relation and there was a cluster with shared patients from the 2006 and 2016 isolates. It included a patient from Tabriz from the 2016 isolates and a patient from Urmia from the 2006 isolates (Figures 1 and 2). The minimum spanning tree of the isolates showed a close relation among clustered and non-clustered isolates of each sample group from 2006 and 2016 (Figure 3). The comparison among patterns of the present study and other identified international patterns showed that the 2016 isolates had a closer relationship with the international pattern than the 2006 isolates. In the 2006 isolates, the patterns were close to "S" and "LAM" strains and in the 2016 isolates, they were close to "Beijing", "New Delhi", and "Uganda" strains (Figure 4). The minimum estimate for the tuberculosis proportion, which is due to the recent transmission of tuberculosis, was 36.7% for the 2006 isolates and 17.6% for the 2016 isolates.

5. Discussion

Phylogenetic analysis plays an important role in the assessment of the evolutionary history of various bacterial strains, such as *M. tuberculosis*. In addition, the DNA sequences play a key role in the futures of genomic evolution, including adaptation to different host environments, enhanced virulence factors, resistance to antibacterial agents, and other changes in important properties. Repeated sequences are possibly related to the molecular basis of widespread clonal variants and the polymorphism of *M. tuberculosis* structural genes. We identified 284 *M. tuberculosis* isolates in both 2006 and 2016 years by using the combination of MIRU-VNTR and ETR-VNTR methods in the northwest of Iran. In the present study, we determined the dynamics of transmission in this region. In the 2006 isolates, we had no Azeri patients referring to the East-Azerbaijan center for tuberculosis, but in the 2016 isolates, we had more admissions of Azeri tuberculosis patients, which caused more MDR isolates. In addition, in 30% of the clusters, we had Azeri patients. Because of the close culture and distance, language similarity, appropriate treatment

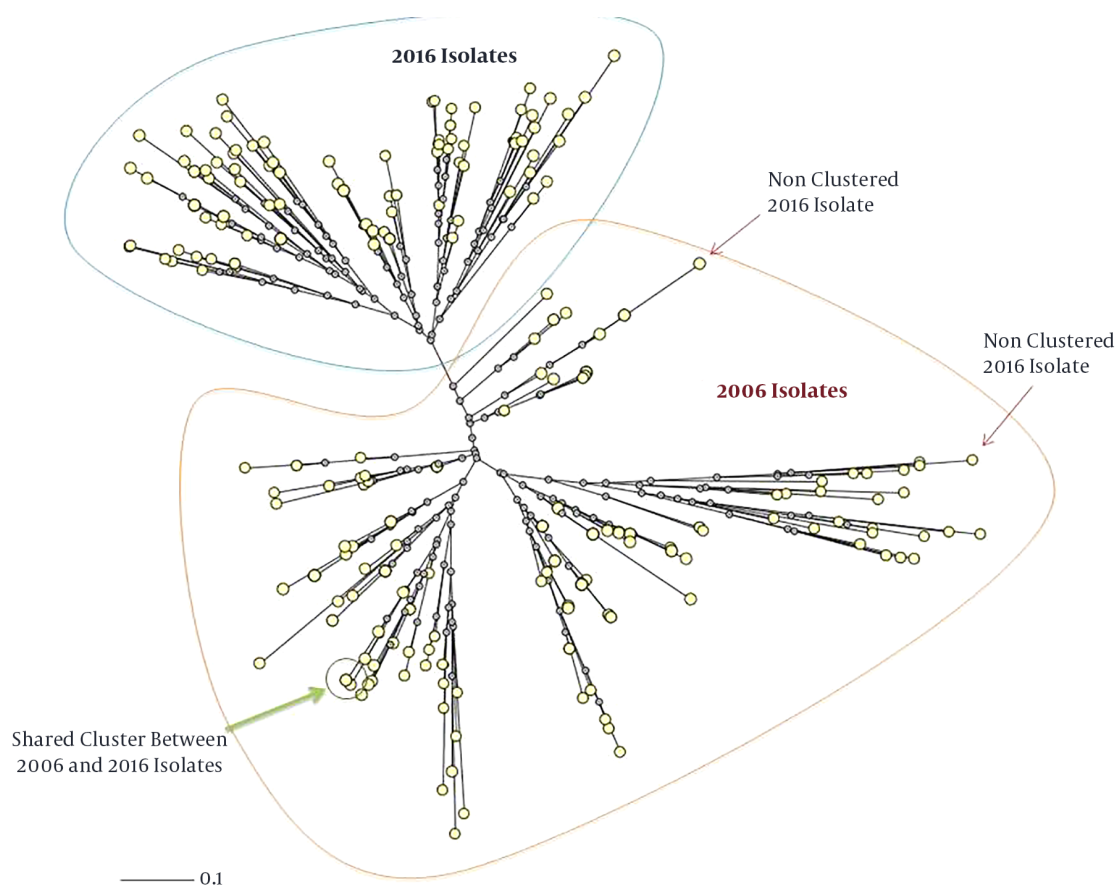


Figure 1. Phylogenetic comparison of isolates indicating complete changes in the molecular pattern of isolates during 10 years (neighbor-joining tree provided by MIRU-VNTRplus). Only three isolates from the 2016 isolates had a close relationship with the 2006 isolates, which one of them was in a shared cluster with one from the 2006 isolates.

facilities, and low medical cost, Azeri patients prefer to refer to Tabriz (the capital city of East Azerbaijan province) for the treatment of tuberculosis.

The presence of patterns related to “Beijing”, “S”, and “LAM” strains, which are famous MDR patterns of tuberculosis (28, 29), was directly related to Azeri patients referring to this region. Beijing, S, and LAM strains are more influenced by the worldwide transmission of tuberculosis. Increased cross-border immigration of Azeri patients with more MDR isolates can contribute to the increased risk of tuberculosis among the Tabriz population. The Republic of Azerbaijan is a traditionally prevalent center for Beijing genotypes of tuberculosis (30). However, recent studies indicated an increasing prevalence of Beijing and New Delhi strains in Iran by immigration from Afghanistan to the west and northwest of Iran (31, 32). The increase in the number of genotypes with resistance to antibiotics can cause hardship in infection control that needs attention for con-

trol of infection transmission from Republic of Azerbaijan to Iran. These patients prefer treatment in Iran because of free drug and treatment courses in Iran. Providing clinics in borders or inside Republic of Azerbaijan and restricted admission of these patients can control the transmission of infection to the northwest of Iran.

According to the obtained results, three isolates from 2016 had a close relation to the isolates from 2006 and there was a cluster with shared patients from the 2006 and 2016 isolates. These results demonstrated that there were three patients with relapse infection in 2016 who had tuberculosis in 2006. These patients were from different cities in this region; this transmission can indicate the Ice Mountain and show the visible part of the transmission of tuberculosis and the need for tracing possible infected patients untreated and unidentified. Using whole-genome sequencing can help us better understand the changes in the tuberculosis genome during transmission in this time

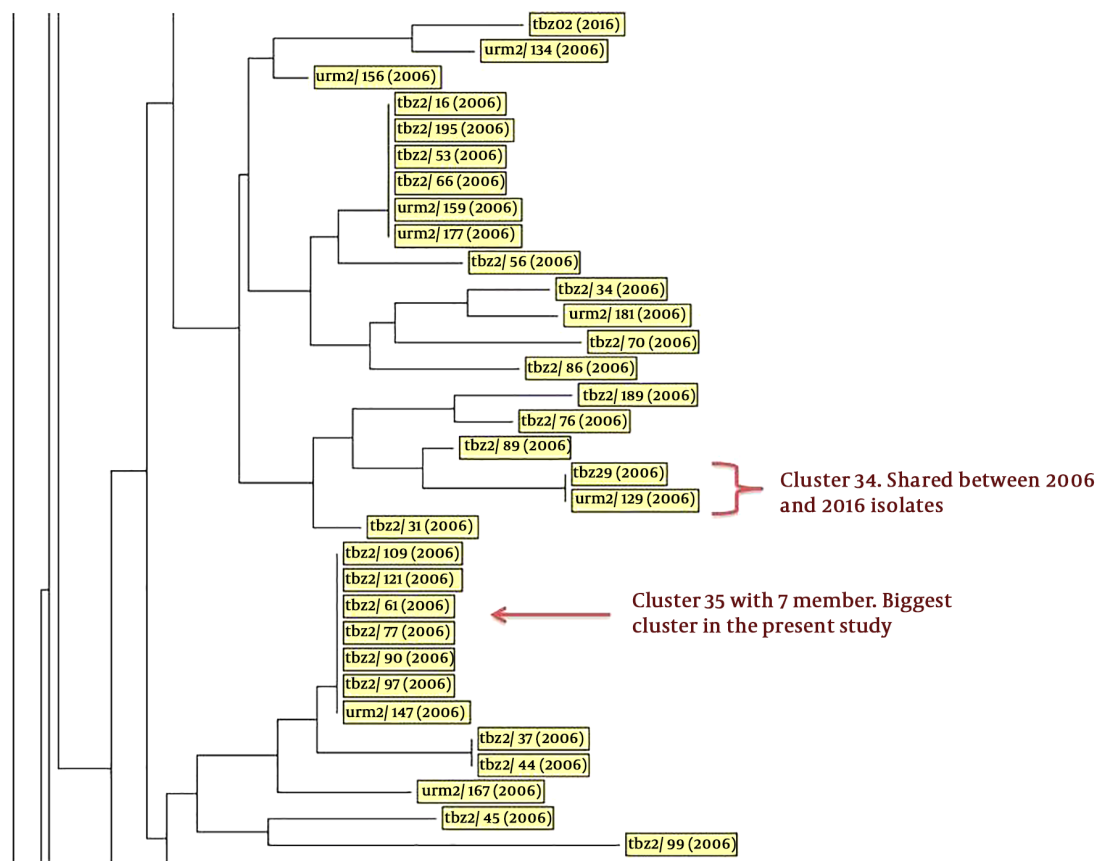


Figure 2. Part of the neighbor-joining phylogenetic tree of isolates. This part reveals a cluster with shared isolates from the years 2006 and 2016. One of these isolates was from Tabriz city and another one from Urmia, which indicates the transmission of isolates between these two patients during this time. Also, a cluster with identity 35 had seven members and was the biggest cluster in this study (the complete tree is provided in Appendix 1 in Supplementary File). Abbreviations indicate the origin of isolates; tbz: Tabriz; urm: Urmia.

period (33). Figure 2 shows that in cluster 34, we had two patients with an exact similar pattern but from different isolates, indicating the circulation of this isolate in patients and its transmission in this region. The early patient in this cluster had no background of tuberculosis in her family and transmission probably happened in the community.

The minimum estimation was 17% for the 2016 isolates and 36% for the 2006 isolates. The reduced number of isolates in the same period in 2016 and 2006 and less minimum estimate for the tuberculosis proportion indicated less transmission of tuberculosis and the success of tuberculosis transmission control in our region. In our region, the increase in prescribing vitamin D (34), free treatment of tuberculosis, and controlling the treatment trends according to the WHO guidelines (35) have led to the reduc-

tion in tuberculosis in 2016 compared to 2006. In an Iranian study from Tehran, it was reported as 24% (36); in other regions worldwide, it was reported from 14.6% (37) in French Guiana to 32% in South African gold miners (38). In a study, the minimum estimation for tuberculosis transmission was 5.1% in patients with negative results on nucleic acid amplification in the United States and it was 11.2% in smear-negative tuberculosis patients (39). These results indicate a reduced rate in our region compared to previous studies.

5.1. Conclusion

The results of the present study indicate tuberculosis controlling in the northwest of Iran was satisfactory but in spite of the reduced number of tuberculosis patients, the isolates were more resistant and had a close relationship with worldwide strains. Cross-border immigration

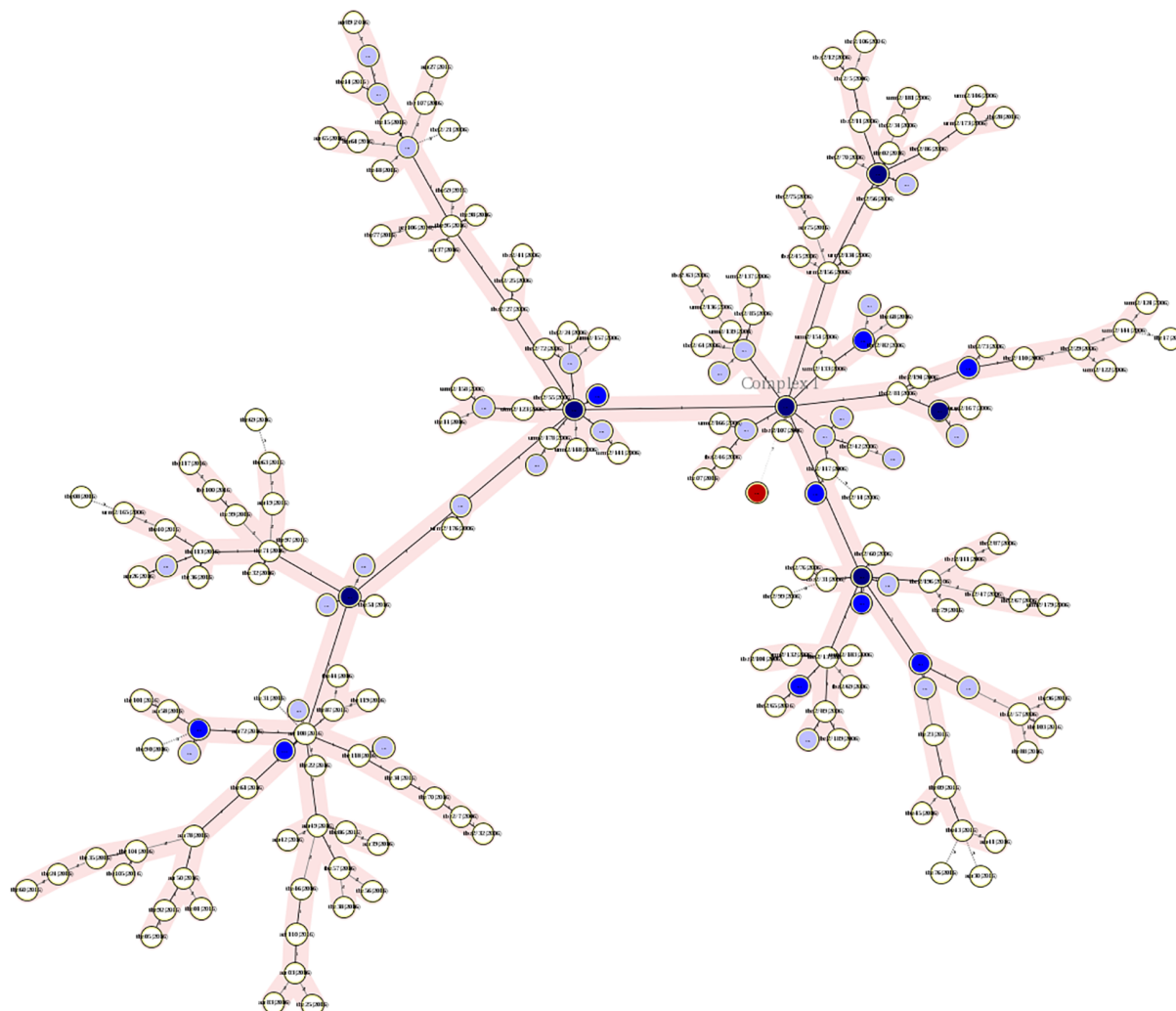


Figure 3. Minimum spanning tree of isolates indicating the number of clusters and their relation with other single isolates. This figure shows how a cluster had a close MIRU-VNTR pattern with non-clustered isolates or other clusters. Red circles are isolates that had no MIRU-VNTR pattern. The darkness of circles indicates the number of isolates in a cluster. Two right branches belong to the 2006 isolates and the left branch belongs to the 2016 isolates.

for treatment from Republic of Azerbaijan had significant participation in the recent transmission of tuberculosis in this region. In conclusion, the strict control of patients commuting and developing new tuberculosis clinics inside Republic of Azerbaijan can play a key role in the control of tuberculosis transmission in the northwest of Iran.

Supplementary Material

Supplementary material(s) is available [here](#) [To read supplementary materials, please refer to the journal website and open PDF/HTML].

Acknowledgments

This study had financial support from the Faculty of Medicine, Tabriz University of Medical Sciences, and was done in the DARC- Drug Applied Research Center, Tabriz University of Medical Sciences, Tabriz, Iran.

Footnotes

Authors' Contribution: Mohammad Hossein Soroush Barhaghi: Study design, data collection, data analysis, and manuscript preparation; Sepehr Taghizadeh: Study design, data collection, data analysis, and manuscript preparation.

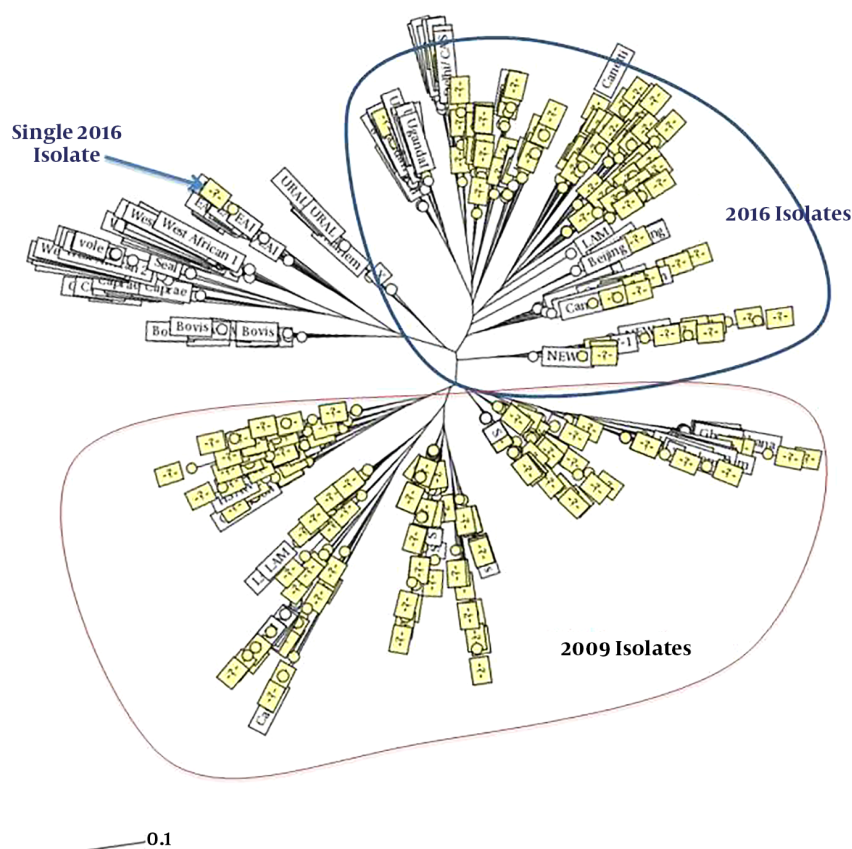


Figure 4. Comparing patterns with other identified international patterns indicating the closer pattern of the 2016 isolates with the 2006 isolates. The presence of patterns close to the Beijing pattern, which is one of the famous MDR patterns of tuberculosis, indicates that isolates in this region are more the result of the worldwide transmission of tuberculosis, as well as the result of more patients from Republic of Azerbaijan admitted in our region. In the 2006 isolates, the patterns were close to “S” and “LAM” strains and in the 2016 isolates they were close to “Beijing”, “New Delhi”, and “Uganda” strains.

ration; Peyvand Kashi: Study design, data collection, data analysis, and manuscript preparation; Mohammad Asgharzadeh: Study design, manuscript preparation, and final draft preparation; Pourya Gholizadeh: Data collection, manuscript preparation, and final draft preparation; Khudaverdi Ganbarov: Data collection, manuscript preparation, and final draft preparation; Asghar Tanomand: Study design, manuscript preparation, and final draft preparation; Milad Bastami: Study design, manuscript preparation, and final draft preparation; Seyyed Reza Moadab: Manuscript preparation and final draft preparation; Behrooz Shokouhi: Manuscript preparation and final draft preparation; Hossein Samadi Kafil: Study design, data collection, data analysis, manuscript preparation, final draft preparation, and the head of the research group.

Conflict of Interests: None to declare.

Ethical Approval: This study was a cross-sectional study conducted in Tabriz University of Medical Sciences sup-

ported by the Faculty of Medicine with Local Ethics Committee approval number IR.TBZMED.REC.1397.353.

Funding/Support: The Faculty of Medicine, Tabriz University of Medical Sciences, Tabriz, Iran, funded the study.

References

- Asgharzadeh M, Samadi Kafil H, Pourostadi M, Asadi Faezi N, Rashedi J, Mahdavi-pour B. Strain differentiation of *Mycobacterium tuberculosis* for epidemiology in northwest of Iran. *Cell Mol Biol (Noisy-le-grand)*. 2016;**62**(8):15–20. [PubMed: 27545209].
- Zahedi Bialvaei A, Asgharzadeh M, Aghazadeh M, Nourazarian M, Samadi Kafil H. Challenges of tuberculosis in Iran. *Jundishapur J Microbiol*. 2017;**10**(3). doi: 10.5812/jjm.37866.
- Pourostadi M, Rashedi J, Mahdavi Poor B, Samadi Kafil H, Shirazi S, Asgharzadeh M. Molecular diversity of *Mycobacterium tuberculosis* strains in Northwestern Iran. *Jundishapur J Microbiol*. 2016;**9**(9). e35520. doi: 10.5812/jjm.35520. [PubMed: 27800145]. [PubMed Central: PMC5086081].
- Asgharzadeh M, Khakpour M, Salehi TZ, Kafil HS. Use of mycobacterial interspersed repetitive unit-variable-number tandem repeat

- typing to study Mycobacterium tuberculosis isolates from East Azarbaijan province of Iran. *Pak J Biol Sci.* 2007;**10**(21):3769-77. doi: [10.3923/pjbs.2007.3769.3777](https://doi.org/10.3923/pjbs.2007.3769.3777). [PubMed: [19090229](https://pubmed.ncbi.nlm.nih.gov/19090229/)].
5. Addis Z, Adem E, Alemu A, Birhan W, Mathewos B, Tachebele B, et al. Prevalence of smear positive pulmonary tuberculosis in Gondar prisoners, North West Ethiopia. *Asian Pac J Trop Med.* 2015;**8**(2):127-31. doi: [10.1016/S1995-7645\(14\)60302-3](https://doi.org/10.1016/S1995-7645(14)60302-3). [PubMed: [25902026](https://pubmed.ncbi.nlm.nih.gov/25902026/)].
 6. Lonnroth K, Migliori GB, Abubakar I, D'Ambrosio L, de Vries G, Diel R, et al. Towards tuberculosis elimination: An action framework for low-incidence countries. *Eur Respir J.* 2015;**45**(4):928-52. doi: [10.1183/09031936.00214014](https://doi.org/10.1183/09031936.00214014). [PubMed: [25792630](https://pubmed.ncbi.nlm.nih.gov/25792630/)]. [PubMed Central: [PMC4391660](https://pubmed.ncbi.nlm.nih.gov/PMC4391660/)].
 7. Metanat M, Sharifi-Mood B, Shahreki S, Dawoudi SH. Prevalence of multidrug-resistant and extensively drug-resistant tuberculosis in patients with pulmonary tuberculosis in Zahedan, Southeastern Iran. *Iran Red Crescent Med J.* 2012;**14**(1):53-5. [PubMed: [22737557](https://pubmed.ncbi.nlm.nih.gov/22737557/)]. [PubMed Central: [PMC3372018](https://pubmed.ncbi.nlm.nih.gov/PMC3372018/)].
 8. Afaghi-Gharamaleki A, Moaddab S, Darbouy M, Ansarin K, Hanifian S. Determining the risk of intra-community transmission of tuberculosis in the Northwest of Iran through 15 Loci Miru-Vntr typing. *Eur J Microbiol Immunol (Bp).* 2017;**7**(1):46-54. doi: [10.1556/1886.2016.00033](https://doi.org/10.1556/1886.2016.00033). [PubMed: [28386470](https://pubmed.ncbi.nlm.nih.gov/28386470/)]. [PubMed Central: [PMC5372480](https://pubmed.ncbi.nlm.nih.gov/PMC5372480/)].
 9. Monteserin J, Camacho M, Barrera L, Palomino JC, Ritacco V, Martin A. Genotypes of Mycobacterium tuberculosis in patients at risk of drug resistance in Bolivia. *Infect Genet Evol.* 2013;**17**:195-201. doi: [10.1016/j.meegid.2013.04.010](https://doi.org/10.1016/j.meegid.2013.04.010). [PubMed: [23603419](https://pubmed.ncbi.nlm.nih.gov/23603419/)].
 10. Nakamura Y, Carlson M, Krapcho K, Kanamori M, White R. New approach for isolation of VNTR markers. *Am J Hum Genet.* 1988;**43**(6):854-9. [PubMed: [2904220](https://pubmed.ncbi.nlm.nih.gov/2904220/)]. [PubMed Central: [PMC1715598](https://pubmed.ncbi.nlm.nih.gov/PMC1715598/)].
 11. Supply P, Allix C, Lesjean S, Cardoso-Oelemann M, Rusch-Gerdes S, Willery E, et al. Proposal for standardization of optimized mycobacterial interspersed repetitive unit-variable-number tandem repeat typing of Mycobacterium tuberculosis. *J Clin Microbiol.* 2006;**44**(12):4498-510. doi: [10.1128/JCM.01392-06](https://doi.org/10.1128/JCM.01392-06). [PubMed: [17005759](https://pubmed.ncbi.nlm.nih.gov/17005759/)]. [PubMed Central: [PMC1698431](https://pubmed.ncbi.nlm.nih.gov/PMC1698431/)].
 12. Quan TP, Bawa Z, Foster D, Walker T, Del Ojo Elias C, Rathod P, et al. Evaluation of whole-genome sequencing for mycobacterial species identification and drug susceptibility testing in a clinical setting: A large-scale prospective assessment of performance against line probe assays and phenotyping. *J Clin Microbiol.* 2018;**56**(2). doi: [10.1128/JCM.01480-17](https://doi.org/10.1128/JCM.01480-17). [PubMed: [29167290](https://pubmed.ncbi.nlm.nih.gov/29167290/)]. [PubMed Central: [PMC5786738](https://pubmed.ncbi.nlm.nih.gov/PMC5786738/)].
 13. Barnes PF, Cave MD. Molecular epidemiology of tuberculosis. *N Engl J Med.* 2003;**349**(12):1149-56. doi: [10.1056/NEJMr021964](https://doi.org/10.1056/NEJMr021964). [PubMed: [13679530](https://pubmed.ncbi.nlm.nih.gov/13679530/)].
 14. Zhang L, Chen J, Shen X, Gui X, Mei J, Deriemer K, et al. Highly polymorphic variable-number tandem repeats loci for differentiating Beijing genotype strains of Mycobacterium tuberculosis in Shanghai, China. *FEMS Microbiol Lett.* 2008;**282**(1):22-31. doi: [10.1111/j.1574-6968.2008.01081.x](https://doi.org/10.1111/j.1574-6968.2008.01081.x). [PubMed: [18336551](https://pubmed.ncbi.nlm.nih.gov/18336551/)].
 15. Smittipat N, Billamas P, Palittapongarnpim M, Thong-On A, Temu MM, Thanakijcharoen P, et al. Polymorphism of variable-number tandem repeats at multiple loci in Mycobacterium tuberculosis. *J Clin Microbiol.* 2005;**43**(10):5034-43. doi: [10.1128/JCM.43.10.5034-5043.2005](https://doi.org/10.1128/JCM.43.10.5034-5043.2005). [PubMed: [16207958](https://pubmed.ncbi.nlm.nih.gov/16207958/)]. [PubMed Central: [PMC1248453](https://pubmed.ncbi.nlm.nih.gov/PMC1248453/)].
 16. Mazars E, Lesjean S, Banuls AL, Gilbert M, Vincent V, Gicquel B, et al. High-resolution minisatellite-based typing as a portable approach to global analysis of Mycobacterium tuberculosis molecular epidemiology. *Proc Natl Acad Sci U S A.* 2001;**98**(4):1901-6. doi: [10.1073/pnas.98.4.1901](https://doi.org/10.1073/pnas.98.4.1901). [PubMed: [11172048](https://pubmed.ncbi.nlm.nih.gov/11172048/)]. [PubMed Central: [PMC29354](https://pubmed.ncbi.nlm.nih.gov/PMC29354/)].
 17. Allix-Beguec C, Wahl C, Hanekom M, Nikolayevskyy V, Drobniewski F, Maeda S, et al. Proposal of a consensus set of hypervariable mycobacterial interspersed repetitive-unit-variable-number tandem-repeat loci for subtyping of Mycobacterium tuberculosis Beijing isolates. *J Clin Microbiol.* 2014;**52**(1):164-72. doi: [10.1128/JCM.02519-13](https://doi.org/10.1128/JCM.02519-13). [PubMed: [24172154](https://pubmed.ncbi.nlm.nih.gov/24172154/)]. [PubMed Central: [PMC3911419](https://pubmed.ncbi.nlm.nih.gov/PMC3911419/)].
 18. Luo T, Yang C, Pang Y, Zhao Y, Mei J, Gao Q. Development of a hierarchical variable-number tandem repeat typing scheme for Mycobacterium tuberculosis in China. *PLoS One.* 2014;**9**(2). e89726. doi: [10.1371/journal.pone.0089726](https://doi.org/10.1371/journal.pone.0089726). [PubMed: [24586989](https://pubmed.ncbi.nlm.nih.gov/24586989/)]. [PubMed Central: [PMC3934936](https://pubmed.ncbi.nlm.nih.gov/PMC3934936/)].
 19. Warren RM, Victor TC, Streicher EM, Richardson M, van der Spuy GD, Johnson R, et al. Clonal expansion of a globally disseminated lineage of Mycobacterium tuberculosis with low IS6110 copy numbers. *J Clin Microbiol.* 2004;**42**(12):5774-82. doi: [10.1128/JCM.42.12.5774-5782.2004](https://doi.org/10.1128/JCM.42.12.5774-5782.2004). [PubMed: [15583312](https://pubmed.ncbi.nlm.nih.gov/15583312/)]. [PubMed Central: [PMC535222](https://pubmed.ncbi.nlm.nih.gov/PMC535222/)].
 20. Merker M, Blin C, Mona S, Duforet-Frebourg N, Lecher S, Willery E, et al. Evolutionary history and global spread of the Mycobacterium tuberculosis Beijing lineage. *Nat Genet.* 2015;**47**(3):242-9. doi: [10.1038/ng.3195](https://doi.org/10.1038/ng.3195). [PubMed: [25599400](https://pubmed.ncbi.nlm.nih.gov/25599400/)].
 21. Oelemann MC, Diel R, Vatin V, Haas W, Rusch-Gerdes S, Loch C, et al. Assessment of an optimized mycobacterial interspersed repetitive-unit-variable-number tandem-repeat typing system combined with spoligotyping for population-based molecular epidemiology studies of tuberculosis. *J Clin Microbiol.* 2007;**45**(3):691-7. doi: [10.1128/JCM.01393-06](https://doi.org/10.1128/JCM.01393-06). [PubMed: [17192416](https://pubmed.ncbi.nlm.nih.gov/17192416/)]. [PubMed Central: [PMC1829086](https://pubmed.ncbi.nlm.nih.gov/PMC1829086/)].
 22. Frothingham R, Meeker-O'Connell WA. Genetic diversity in the Mycobacterium tuberculosis complex based on variable numbers of tandem DNA repeats. *Microbiology.* 1998;**144** (Pt 5):1189-96. doi: [10.1099/00221287-144-5-1189](https://doi.org/10.1099/00221287-144-5-1189). [PubMed: [9611793](https://pubmed.ncbi.nlm.nih.gov/9611793/)].
 23. Asgharzadeh M, Samadi Kafil H, Khakpour M. Comparison of mycobacterial interspersed repetitive unit-variable number tandem repeat and IS6110-RFLP methods in identifying epidemiological links in patients with tuberculosis in Northwest of Iran. *Ann Microbiol.* 2008;**58**(2):333-9. doi: [10.1007/bf03175339](https://doi.org/10.1007/bf03175339).
 24. Asgharzadeh M, Shahbaban K, Samadi Kafil H, Rafi A. Use of DNA fingerprinting in identifying the source case of tuberculosis in East Azarbaijan Province of Iran. *J Med Sci.* 2007;**7**(3):418-21. doi: [10.3923/jms.2007.418.421](https://doi.org/10.3923/jms.2007.418.421).
 25. Asgharzadeh M, Kafil HS, Roudsary AA, Hanifi GR. Tuberculosis transmission in Northwest of Iran: Using MIRU-VNTR, ETR-VNTR and IS6110-RFLP methods. *Infect Genet Evol.* 2011;**11**(1):124-31. doi: [10.1016/j.meegid.2010.09.013](https://doi.org/10.1016/j.meegid.2010.09.013). [PubMed: [20951237](https://pubmed.ncbi.nlm.nih.gov/20951237/)].
 26. Supply P, Mazars E, Lesjean S, Vincent V, Gicquel B, Loch C. Variable human minisatellite-like regions in the Mycobacterium tuberculosis genome. *Mol Microbiol.* 2000;**36**(3):762-71. doi: [10.1046/j.1365-2958.2000.01905.x](https://doi.org/10.1046/j.1365-2958.2000.01905.x). [PubMed: [10844663](https://pubmed.ncbi.nlm.nih.gov/10844663/)].
 27. Allix-Beguec C, Harmsen D, Weniger T, Supply P, Niemann S. Evaluation and strategy for use of MIRU-VNTRplus, a multifunctional database for online analysis of genotyping data and phylogenetic identification of Mycobacterium tuberculosis complex isolates. *J Clin Microbiol.* 2008;**46**(8):2692-9. doi: [10.1128/JCM.00540-08](https://doi.org/10.1128/JCM.00540-08). [PubMed: [18550737](https://pubmed.ncbi.nlm.nih.gov/18550737/)]. [PubMed Central: [PMC2519508](https://pubmed.ncbi.nlm.nih.gov/PMC2519508/)].
 28. Bifani PJ, Mathema B, Kurepina NE, Kreiswirth BN. Global dissemination of the Mycobacterium tuberculosis W-Beijing family strains. *Trends Microbiol.* 2002;**10**(1):45-52. doi: [10.1016/s0966-842x\(01\)02277-6](https://doi.org/10.1016/s0966-842x(01)02277-6). [PubMed: [11755085](https://pubmed.ncbi.nlm.nih.gov/11755085/)].
 29. Ignatova A, Dubiley S, Stepanshina V, Shemyakin I. Predominance of multi-drug-resistant LAM and Beijing family strains among Mycobacterium tuberculosis isolates recovered from prison inmates in Tula Region, Russia. *J Med Microbiol.* 2006;**55**(Pt 10):1413-8. doi: [10.1099/jmm.0.46575-0](https://doi.org/10.1099/jmm.0.46575-0). [PubMed: [17005791](https://pubmed.ncbi.nlm.nih.gov/17005791/)].
 30. Pfyffer GE, Strassle A, van Gorkum T, Portaels F, Rigouts L, Mathieu C, et al. Multidrug-resistant tuberculosis in prison inmates, Azerbaijan. *Emerg Infect Dis.* 2001;**7**(5):855-61. doi: [10.3201/eid0705.017514](https://doi.org/10.3201/eid0705.017514). [PubMed: [11747699](https://pubmed.ncbi.nlm.nih.gov/11747699/)]. [PubMed Central: [PMC2631864](https://pubmed.ncbi.nlm.nih.gov/PMC2631864/)].
 31. Mohajeri P, Moradi S, Atashi S, Farahani A. Mycobacterium tuberculosis Beijing genotype in Western Iran: Distribution and drug resistance. *J Clin Diagn Res.* 2016;**10**(10):DC05-7. doi: [10.5455/jcd.2016.1010.1005](https://doi.org/10.5455/jcd.2016.1010.1005).

- 10.7860/JCDR/2016/20893.8689. [PubMed: 27891336]. [PubMed Central: PMC5121674].
32. Sahebi L, Ansarin K, Hoffner S, Mohajeri P, Mohammadi A. Beijing strains of Mycobacterium tuberculosis in smear-positive tuberculosis patients in North-West and West of Iran. *Adv Biomed Res.* 2016;5:181. doi: 10.4103/2277-9175.190982. [PubMed: 28028521]. [PubMed Central: PMC5157006].
 33. Bryant JM, Harris SR, Parkhill J, Dawson R, Diacon AH, van Helden P, et al. Whole-genome sequencing to establish relapse or reinfection with Mycobacterium tuberculosis: A retrospective observational study. *Lancet Respir Med.* 2013;1(10):786–92. doi: 10.1016/S2213-2600(13)70231-5. [PubMed: 24461758]. [PubMed Central: PMC3861685].
 34. Wilkinson RJ, Llewelyn M, Toossi Z, Patel P, Pasvol G, Lalvani A, et al. Influence of vitamin D deficiency and vitamin D receptor polymorphisms on tuberculosis among Gujarati Asians in west London: A case-control study. *Lancet.* 2000;355(9204):618–21. doi: 10.1016/S0140-6736(99)02301-6. [PubMed: 10696983].
 35. World Health Organization. *WHO treatment guidelines for drug-resistant tuberculosis (2016 update) (WHO/HTM/TB/2016.04)*. Geneva: WHO; 2016.
 36. Torkaman MR, Nasiri MJ, Farnia P, Shahhosseiny MH, Mozafari M, Velayati AA. Estimation of recent transmission of Mycobacterium tuberculosis strains among Iranian and Afghan immigrants: A cluster-based study. *J Clin Diagn Res.* 2014;8(9):DC05–8. doi: 10.7860/JCDR/2014/8886.4864. [PubMed: 25386431]. [PubMed Central: PMC4225883].
 37. Guernier V, Sola C, Brudey K, Guegan JF, Rastogi N. Use of cluster-graphs from spoligotyping data to study genotype similarities and a comparison of three indices to quantify recent tuberculosis transmission among culture positive cases in French Guiana during a eight year period. *BMC Infect Dis.* 2008;8:46. doi: 10.1186/1471-2334-8-46. [PubMed: 18410681]. [PubMed Central: PMC2375894].
 38. Mathema B, Lewis JJ, Connors J, Chihota VN, Shashkina E, van der Meulen M, et al. Molecular epidemiology of Mycobacterium tuberculosis among South African gold miners. *Ann Am Thorac Soc.* 2015;12(1):12–20. doi: 10.1513/AnnalsATS.201404-150OC. [PubMed: 25419914]. [PubMed Central: PMC4342800].
 39. Xie YL, Cronin WA, Proschan M, Oatis R, Cohn S, Curry SR, et al. Transmission of Mycobacterium tuberculosis from patients who are nucleic acid amplification test negative. *Clin Infect Dis.* 2018;67(11):1653–9. doi: 10.1093/cid/ciy365. [PubMed: 29697779]. [PubMed Central: PMC6233677].