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Research Article

*PNPLA*3 and *TM6SF2*, but Not *MBOAT7*, Are Associated with Steatosis and HBV Viral Persistence in Pakistani Population

Ismail Jalil ^[]^{1, 2, *}, Muhammad Arshad ^[]², Shahtaj Khan ³ and Javid Iqbal Dasti²

¹School of Biotechnology & Biomolecular Sciences, University of New South Wales, Sydney, Australia
²Department of Microbiology, Quaid-i-Azam University, Islamabad, Pakistan
³Department of Pathology, Hayatabad Medical Complex, Peshawar, Pakistan

^{*} *Corresponding author*: School of Biotechnology & Biomolecular Sciences, University of New South Wales, Kensington, Sydney, Australia. Tel: +61-0293852029, Email: babs@unsw.edu.au

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Abstract

Background: Hepatitis B infection has an intimate relationship with lipids. The role of lipid-related variants remains unknown in the risk of hepatitis B infection persistence and steatosis in the Pakistani population. Recently, three GWAS-based polymorphisms in the *TM6SF2*, *PNPLA3*, and *MB0AT7* genes have suggested being associated with steatosis and/or liver injury. However, the role of these variants is unknown in Hepatitis B virus (HBV) persistence and steatosis in the Pakistani population.

Objectives: We determined whether *TM6SF2*, *PNPLA3*, and *MBOAT7* genetic variations are associated with HBV chronicity and hepatic steatosis in the Pakistani population.

Methods: A total of 297 patients visiting the Hayat Abad Medical Complex in Peshawar were included in this study. Clinical analysis, along with genotyping of SNPs in the *PNPLA3*, *TM6SF2*, and *MBOAT* genes, was performed using the TaqMan genotyping assay. Logistic regression analysis, along with other tests as appropriate, was used to determine the association of the analyzed SNPs with HBV persistence, chronicity, and hepatic steatosis in the analyzed set of patients.

Results: In 297 subjects (240 HBV patients and 57 healthy controls), *PNPLA*3 rs738409 (OR: 0.43, 95% CI: 0.23 - 0.81, P = 0.009) and *TM6SF2* rs58542926 (P = 0.018) genotypes were independently associated with the risk of chronic HBV infection, but not *MBOAT* rs641738 (OR: 1.3, 95% CI: 0.64 - 2.62, P = 0.454). We also observed that the *PNPLA3* rs738409 GG genotype was associated with 2.97-fold and *TM6SF2* rs58542926 genotype T allele with 1.54-fold increased risk of steatosis.

Conclusions: *PNPLA*3 rs738409 and *TM6SF2* rs58542926, but not *MBOAT* rs641738, were the risk variants for HBV persistence and steatosis in the Pakistani population.

Keywords: CHB, TM6SF2, PNPLA3, MBOAT7, Steatosis, Persistence

1. Background

Hepatitis B virus (HBV) is highly prevalent, as one-third of the world's population has been exposed to it and 350 -400 million have developed a chronic infection, with > 1 million deaths per year from cirrhosis and liver cancer (1). In Pakistan, almost more than 9 to 12 million people are estimated to be living with HBV or Hepatitis C virus (HCV), with a carrier rate of 3% - 5% (2, 3). Hepatitis B virus is responsible for more than half of all new cases of liver cancer and it is among the "top ten" causes of cancer death. On the other hand, non-alcoholic fatty liver disease (NAFLD) is currently on a trajectory to become the most common liver disease, as it has affected around 20-30% of the global population (4). This renders that the co-occurrence of both diseases is not infrequent and studying this relationship is of pivotal importance.

Hepatitis B has an intimate link with hepatic lipid metabolism. An opposite association is observed between positive Hepatitis B surface antigen (HBsAg) status and the prevalence of fatty liver in humans. It has been suggested that HBsAg seroclearance is more than three-fold higher in those with moderate-to-severe hepatic steatosis than in those without hepatic steatosis (5). A similar observation was reported in mouse models and *in vitro* (6). Multiple reports have identified variants in patatin-like phospholipase domain containing 3 (PNPLA3) (I148M) and transmembrane 6 superfamily member 2 (TM6SF2)(E167K) as risk variants for steatosis in patients with NAFLD. However, the data are scarce on the impact of these variants on hepatic steatosis in patients with viral hepatitis, especially in Asian patients with hepatitis B. Notably, it is proposed that these polymorphisms are associated with HBV-DNA levels, sug-

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gesting a potential role in HBV persistence (7-9).

In 2015, a single-nucleotide polymorphism (SNP) (rs641738) in membrane-bound O-acyltransferase domain containing 7 (*MBOAT7*) was identified by a genome-wide association study as a risk variant for alcohol-related cirrhosis (10). The next reports demonstrated that this polymorphism was associated with liver injury in NAFLD, hepatitis C, and hepatitis B (11-13). However, the impact of this variant on steatosis is still less clear.

2. Objectives

The role of *PNPLA3* (HGNC:18590), *TM6SF2* (HGNC:11861), and *MBOAT7* (HGNC:15505) genetic polymorphisms remains obscure in the Pakistani population. Therefore, we aimed to investigate the association of *PNPLA3*, *TM6SF2*, and *MBOAT7* polymorphisms with hepatic steatosis and hepatitis B persistence in the Pakistani population.

3. Methods

3.1. Patient Cohort

The study comprised 297 Pakistani subjects (240 HBVinfected patients and 57 healthy controls). In the period from July 2016 to May 2017, patients with positive HBsAg tests were recruited. Among 240 chronic HBV patients, 44 were found with steatosis and 196 were non-steatotic. The aspartate aminotransferase platelets ratio index and ultrasound examination were employed to detect and evaluate liver steatosis. The subjects were recruited from the outpatient Department of Hayatabad Medical Complex (HMC), Peshawar, located in the Northwest of the KPK region of Pakistan. The Hayatabad Medical Complex is a major tertiary care hospital in Peshawar that receives a large flow of subjects/patients from all populations of KPK.

3.2. Exclusion Criteria

Patients were excluded if they abused alcohol (\geq 20 g of alcohol daily), had a history of vaccination against HBV and evidence of co-infection with either human immunodeficiency virus (HIV), HCV, hepatitis delta virus (HDV), or other liver diseases.

3.3. Clinical and Laboratory Assessment

The following information was obtained at the time of blood collection: age, gender, ethnicity, alcohol intake, socioeconomic status, vaccination history, and routine laboratory tests. Hepatic steatosis was evaluated based on ultrasound and APRI. Blood samples were collected in EDTA/Heparin vacutainer tubes to avoid clotting. About 4 mL of the whole blood sample was taken from patients visiting the outpatient department and laboratory investigations were carried out at the Department of Microbiology, Quaid-i-Azam University, Islamabad, and HMC, Peshawar. The liver function, cholesterol, triglyceride, and platelets were assessed using Cobas C111 (Roche) following the manufacturer's instructions and SOPs. The samples were screened for the detection of HBsAg and positive samples were evaluated for anti-HBc-IgM, anti-HBc-IgG, anti-HDV, and anti-HCV. For this purpose, we used commercially available ELISA kits (MBS-SRL, Milano, Italy) according to the manufacturer's instructions.

3.4. DNA Extraction and Amplification

Viral DNA was extracted and purified using commercially available kits (Sacace Biotechnologies S.R.L, Italy). The genomic DNA was extracted using commercially available DNA extraction kits (GeneJET Thermo scientific) and the Phenol-Chloroform method. A proper protocol was followed for the extraction of human DNA. The extracted human DNA was stored at -80°C. Real-time PCR was done for the viral load using Cepheid Smart cycler.

3.5. Genotyping

Genotyping for *PNPLA3* (HGNC:18590) rs738409, *MBOAT7* (HGNC:15505) rs641738, and *TM6SF2* (HGNC:11861) rs58542926 was undertaken using the TaqMan SNP genotyping allelic discrimination method (Applied Biosystems, Foster city, CA, USA) (14). Genotyping was blinded to clinical variables.

3.6. Statistical Analysis

Data are shown as mean and standard deviation (SD), median, range or number, and proportion, as appropriate. The frequency of PNPLA3 rs738409, MBOAT7 rs641738, and TM6SF2 rs58542926 genotypes was compared between different groups using Fisher's exact test. The Cochran-Armitage test was used for the assessment of trends. The student's t-test or non-parametric Wilcoxon-Mann-Whitney U-test or Kruskal-Wallis test was used to compare quantitative data, as appropriate. All tests were two-tailed and P values of < 0.05 were considered significant. Hardy-Weinberg equilibrium tests of PNPLA3 rs738409, TM6SF2 rs58542926, and MBOAT rs641738 were performed using the chi-square test. The HBV DNA levels were log-transformed before modeling. The logistic regression model was used and the results are expressed as OR and 95% CI. Statistical analyses were performed using the statistical software package SPSS for Windows, version 21 (SPSS, Chicago, IL).

4. Results

4.1. Patient Characteristics

The demographic, biochemical, and virological characteristics of the studied HBV-infected hepatic steatotic and non-steatotic patients are presented in Table 1. The median age was 29 years and 28 years in HBV patients and controls, respectively. Moreover, 72.4% and 61.4% were males in the patient and control groups, respectively, and 18.3% of the HBV patients had hepatic steatosis.

4.2. PNPLA3 rs738409 and TM6SF2 but not MBOAT rs641738 Were Associated with the Risk of HBV Chronicity

The genotype distribution of *PNPLA3* rs738409, *TM6SF2*, and *MBOAT7* in HBV-infected patients and the healthy Pakistani population is presented in Table 2. Genotype distribution was in the Hardy-Weinberg equilibrium in both groups. The minor allele frequency (MAF) (G) of *PNPLA3* rs738409 was 0.26, which was significantly higher than that noticed in our healthy cohort (MAF 0.16, P = 0.031); it was also true using the recessive model (P = 0.011). This association remained significant in multiple logistic regression analysis after adjusting for age and gender (OR= 0.41, 95% CI: 0.22 - 0.78, P = 0.006) (Table 3).

The MAF of *TM6SF2* rs58542926 (T) was 0.06, which was significantly higher than that observed in our healthy cohort (MAF 0.00 (P = 0.078); it was also true using the dominant model (P = 0.018). The T allele was significantly associated with HBV chronicity (OR= 1.09, 95% CI: 1.05 - 1.13, P = 0.018). In contrast, the genotype distribution of *MBOAT* rs641738 was not significantly different between HBV-infected patients and healthy controls (0.49 vs. 0.41, P = 0.137); this was also true using the recessive model (P =

Fable 1. Baseline Characteristics of Steatotic and Non-Steatotic Hepatitis B Cohorts ^a			
Variables	Non-Steatosis (N = 196)	Steatosis (N = 44)	P Value
Age ^b , y	26 (4 - 75)	44 (17 - 82)	0.0001 ^c
Gender ^b (male/female)	142/54 (72.4%)	27/17 (61.4%)	0.149
ALT ^b (IU/L)	33 (11 - 211)	175 (25 - 680)	0.0001 ^c
AST ^b (IU/L)	35 (15 - 103)	149 (70 - 459)	0.0001 ^c
Total bilirubin (mg/dL)	0.7 (0.3 - 3.8)	0.9 (0.4 - 6.1)	0.140
Cholesterol (mg/dL)	131 (78 - 260)	138 (78 - 280)	0.001 ^c
TG (mg/dL)	173 (92 - 351)	189 (120 - 312)	0.014 ^c
Platelets ($ imes$ 10 9 /L)	245 (150 - 415)	169 (95 - 243)	0.0001 ^c

Abbreviations: ALT, alanine aminotransferase; AST: Aspartate Aminotransferase; IU, international unit; TB, total bilirubin; TG: triglyceride.

^aContinuous variables were described as median and interquartile range. ^bThe relative difference between two groups.

^cStatistically significant (P < 0.05).

Cohorts ^a			
Genotype	Healthy	Hepatitis B	P Value
PNPLA3 rs738409			
CC	40 (70.2)	122 (50.8)	0.031 ^b
CG	16 (28.1)	110 (45.8)	
GG	1 (1.8)	8 (3.3)	
CC	40 (70.2)	122 (50.8)	0.011 ^b
CG/GG	17 (29.8)	118 (49.2)	
TM6SF2 rs58542926			
CC	57 (100.0)	220 (91.7)	0.078
CT	0 (0.0)	9 (3.8)	
TT	0 (0.0)	11 (4.6)	
CC	57 (100.0)	220 (91.7)	0.018 ^b
CT/TT	0 (0.0)	20 (8.3)	
MBOAT7 rs641738			
CC	12 (21.1)	62 (25.8)	0.137
CT	23 (40.4)	117 (48.8)	
TT	22 (38.6)	61(25.4)	
CC	12 (21.1)	62 (25.8)	0.500
CT/TT	45 (78.9)	178 (74.2)	

Table 2. Genotype Distribution of Genetic Variants in the Healthy and Hepatitis B

^aValues are expressed as No. (%).

^bStatistically significant (P < 0.05).

0.500). This remained the same in multiple logistic regression analysis after accounting for the same variables mentioned above (OR: 1.3, 95% CI: 0.66 - 2.70, P = 0.417) (Table 3).

4.3. PNPLA3 rs738409 and TM6SF2 rs58542926 but not MBOAT rs641738 Were Associated with Hepatic Steatosis

Age, serum cholesterol, and triglycerides were higher in patients with hepatic steatosis though they were insignificant, while significant differences in other clinical variables like ALT, AST, and platelets were also noticed between patients with and without hepatic steatosis, as shown in Table 1. Next, we explored the association of PN-PLA3, TM6SF2, and MBOAT7 with hepatic steatosis. The genotype distribution of PNPLA3 rs738409, TM6SF2, and MBOAT7 in chronic HBV-infected patients and Pakistani population with hepatic steatosis is shown in Table 4. The rs738409 GG genotype, observed in 3% of patients, was associated with steatosis (0.27 vs. 0.21, P = 0.033) (OR: 2.79, 95% CI: 0.64 - 12.16; P = 0.063) thought it was insignificant due to the sample size. The rs58542926 TT genotype, observed in 4.6% of patients, the minor allele (T) frequency (MAF) of TM6SF2 rs58542926 was significantly higher in patients with steatosis than in those without steatosis (0.11 vs. 0.05,

Canotyna	Healthy vs. Chronic Hepatitis B		
Genotype	OR (95%CI) ^a	P Value ^a	
PNPLA3 rs738409			
СС	1		
CG	0.4 (0.23 - 0.83)	0.012 ^b	
GG	0.3 (0.04 - 3.14)	0.370	
Adjusted	0.4 (0.22 - 0.78)	0.006 ^b	
Dominant	2.2 (1.22 - 4.23)	0.009 ^b	
CC	1	1	
CG/GG	0.4 (0.23 - 0.81)	0.009^{b}	
MBOAT7 rs641738			
CC	1		
CT	1.0 (0.47 - 2.17)	0.968	
TT	1.8 (0.84 - 4.09)	0.121	
Adjusted	1.3 (0.66 - 2.70)	0.417	
Dominant	0.7 (0.38 - 1.54)	0.454	
CC	1		
CT/TT	1.3 (0.64 - 2.62)	0.454	

Table 3. Association Between PNPLA3 rs738409, TM6SF2 rs58542926, and MBOAT rs641738 Genotypes and HBV Persistence

Genotype	Non-Steatosis	Steatosis	P Value
PNPLA3 rs738409			
CC	94 (48.0)	28 (63.6)	0.033 ^b
CG	97 (49.4)	13 (29.5)	
GG	5(2.6)	3(6.9)	
TM6SF2 rs58542926			
СС	181 (92.3)	39 (88.6)	0.024 ^b
CT	9 (4.6)	0(0.0)	
TT	6 (3.1)	5 (11.4)	
MBOAT7 rs641738			
CC	53 (27.0)	9 (20.5)	0.666 ^b
CT	94 (48.0)	23 (52.3)	
TT	49 (25.0)	12 (27.2)	

MBOAT7 rs641738 and any of these clinical variables, except

In this study, for the first time, we investigated the role of functional polymorphisms in three main lipid-related

genes, namely PNPLA3, TM6SF2, and MBOAT7, in hepatic

steatosis and HBV chronicity in the Pakistani population.

Consistent with other data reported in different popula-

tions, we demonstrated that PNPLA3 and TM6SF2 but not

MBOAT7 were associated with hepatic steatosis in HBV pa-

tients (7, 13, 15). Interestingly, our data suggest that these

two polymorphisms may be implicated in HBV persistence.

The natural history of viral hepatitis infection including

HBV infection exhibited a marked inter-individual varia-

tion, indicating a pivotal role for genetic basis in shaping the outcome of such patients (16). The last years have witnessed multiple GWAS studies, which revealed multiple

risk variants for common complex diseases including in-

fectious diseases (8, 9, 17-20). However, to date, most stud-

ies were conducted in either Caucasian or other Asian pop-

ulations, with very limited data in the Pakistani popula-

tion, while Pakistan is a country with a very high preva-

ages and serum cholesterol and triglyceride levels, though

not significantly, but no difference was noticed in the HBV-

DNA levels. It is consistent with a suggestion that steatosis

Patients with hepatic steatosis tended to have higher

lence of HBV infection and a large population.

^aValues are expressed as No. (%).

for cholesterol (Table 5).

5. Discussion

^bStatistically significant (P < 0.05).

^aOdds ratio and P values were calculated by using binary logistic regression and multiple logistic regression adjusted for age and gender.

^bStatistically significant (P < 0.05).

P=0.024). The T allele was not significantly associated with steatosis (OR: 1.54, 95% CI: 0.53 - 4.5, P = 0.424). In contrast, the *MBOAT* rs641738 CC genotype, observed in 25.8% of patients, the genotype distribution of MBOAT rs641738 was not significantly different between chronic HBV-infected patients and steatotic cohort (0.49 vs. 0.47, P = 0.666); it was also true using the recessive model (P = 0.448). This remained the same in multiple logistic regression analysis after adjusting for age and gender (OR: 0.9, 95% CI: 0.37 - 2.49, P = 0.950).

4.4. Association of PNPLA3, TM6SF2, MBOAT7 Genotypes with Clinical Variables

Finally, we examined if baseline clinical variables differed among HBV-infected subjects according to the *PN-PLA3* rs738409 genotype; the results are depicted in Table 5. We observed that subjects with rs738409 CC genotype were significantly older than those with CG/GG genotype (P = 0.005), while here we did not observe any significant association between the rs738409 genotype (CC versus CT/TT) and any of the clinical variables (i.e., age, gender, ALT, AST, HBV-DNA, total bilirubin, cholesterol, and TG). Similarly, there was no association between *TM6SF2* rs58542926 or

	Genot	ypes ^a	P Value ^b
PNPLA3 rs738409	СС	CG/GG	
Age	31.9 ± 14.4	26.6 ± 12.1	0.005 ^c
HBV-DNA $\log IU^d$	5.1 ± 2.1	5.4 ± 2.3	0.298
ALT (IU/L)	51.9 ± 60.1	52.6 ± 85.4	0.935
AST (IU/L)	53.0 ± 48.4	49.4 ± 52.9	0.548
Total bilirubin (mg/dL)	0.79 ± 0.41	0.82 ± 0.61	0.630
Cholesterol (mg/dL)	144.6 ± 45.9	135.3 ± 42.2	0.073
TG (mg/dL)	177.5 ± 50.6	169.4 ± 48.6	0.164
Platelets (\times 10 ⁹ /L)	240.6 ± 60.3	238.3 ± 62.9	0.747
IM6SF2 rs58542926	СС	CT/TT	
Age	29.2 ± 13.6	26.8 ± 13.8	0.446
HBV-DNA log IU ^d	5.3 ± 2.2	4.8 ± 2.0	0.432
ALT (IU/L)	50.2 ± 71.1	79.0 ± 88.5	0.087
AST (IU/L)	50.5 ± 49.7	63.9 ± 59.9	0.252
Total bilirubin (mg/dL)	0.80 ± 0.51	0.84 ± 0.44	0.714
Cholesterol (mg/dL)	140.5 ± 44.2	139.2 ± 48.2	0.903
TG (mg/dL)	174.7 ± 50.0	165.3 ± 46.8	0.431
Platelets (\times 10 ⁹ /L)	240.0 ± 60.9	233.7 ± 68.8	0.660
MBOAT7 rs641738	СС	CT/TT	
Age	26.8 ± 13.4	29.8 ± 13.5	0.100
HBV-DNA log IU ^d	5.4 ± 2.2	5.2 ± 2.2	0.527
ALT (IU/L)	44.9 ± 48.4	54.6 ± 78.9	0.323
AST (IU/L)	43.8 ± 33.9	53.9 ± 54.7	0.138
Total bilirubin (mg/dl)	0.82 ± 0.50	0.79 ± 0.51	0.718
Cholesterol (mg/dL)	149.4 ± 44.8	137.4 ± 43.9	0.043 ^c
TG (mg/dL)	180.2 ± 44.0	173.8 ± 51.5	0.199
Platelets (\times 10 ⁹ /L)	233.1 + 51.9	241.7 ± 64.2	0.300

^aValues are expressed as mean \pm SD.

^bP values were calculated by using logistic regression.

^cStatistically significant (P < 0.05).

^dHBV DNA in log 10; IU, international unit.

trin and has triacylglycerol lipase activity, has been shown robustly by multiple studies to have a strong association with liver fat content in NAFLD patients, as well as patients with viral diseases (15, 21, 22). Similarly, nonsynonymous coding variants in transmembrane 6 superfamily member 2 (*TM6SF2*) (E167K) have recently been well studied as genetic risk factors for hepatic fat accumulation and progression to NASH, fibrosis, and HCC. Similar data were reported with hepatic steatosis in patients with viral hepatitis (7, 8, 14, 23). However, the exact function of *TM6SF2* is not known, but it regulates cholesterol synthesis and the secretion of lipoproteins (24). This polymorphism in the *TM6SF2* gene (rs58542926) affects hepatic *TM6SF2* mRNA and protein expression.

In contrast, the association between polymorphisms in the MBOAT7 and hepatic steatosis is controversial; most of the evidence suggests the lack of an association or a very marginal association at the best. Though the mechanisms of MBOAT7 function are still unclear, it is likely involved in hepatic inflammation via its function in the remodeling pathway of phosphoinositides (Land's cycle) that assigns arachidonic acids (AAs) to lysophosphatidylinositol; it can also intensify the inflammatory milieu in macrophages and other immune cells (11, 12, 14). Viral hepatitis including HBV has an intimate interaction with lipids (25, 26). Our data suggest that polymorphisms in PNPLA3 and TM6SF2 but not in MBOAT7 might be implicated in HBV persistence. This is consistent with a couple of reports suggesting that these two polymorphisms modulate HBV-DNA (8, 12). Furthermore, the lack of association between MBOAT7 and HBV persistence is consistent with our current data and others that indicate the effect of MBOAT7 on liver diseases is unlikely to be in hepatic steatosis. Further studies are required to understand the functional mechanisms of these effects and if they can be exploited for therapeutic purposes.

5.1. Conclusions

In conclusion, we demonstrated for the first time that polymorphisms in *PNPLA3* and *TM6SF2*, but not in *MB0AT7*, are associated with steatosis and HBV viral persistence in the Pakistani population. Future direction will be to explore if these findings can aid in guiding efforts for the personalization of medicines and finding novel therapeutic targets.

Footnotes

Authors' Contribution: Conception and design of study: Ismail Jalil, Shahtaj Khan, and Javid Iqbal Dasti; acquisition of data (laboratory or clinical): Ismail Jalil, Muhammad Arshad, and Javid Iqbal Dasti; data analysis and/or interpretation: Ismail Jalil and Javid Iqbal Dasti; drafting of manuscript and/or critical revision and approval of final version of manuscript: Ismail Jalil, Muhammad Arshad, Shahtaj Khan, and Javid I Dasti.

Conflict of Interests: The authors declare none.

Ethical Approval: Ethical approval was obtained from the Human Research Ethics Committee of the Institution Research and Ethics Board of Postgraduate Medical Institute, Hayatabad, Peshawar, Pakistan (PGMI/19566). The study was conducted following the Declaration of Helsinki.

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Patient Consent: Written informed consent, including for genetic testing, was obtained from all participants.

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