



Preparation, Characterization, and Skin Permeation Evaluation of Naproxen Microemulsions for Transdermal Delivery

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Abstract

Microemulsions (MEs) are considered for preparing drug delivery carriers, especially transdermal vehicles. Nonsteroidal anti-inflammatory drugs (NSAIDs) are commonly used to manage chronic and acute pain and inflammatory diseases. However, NSAIDs have drawbacks such as gastrointestinal tract disorders and poor pharmacokinetic properties for oral administration. To address these issues, we evaluated the potential of ME as a transdermal system for locally delivering naproxen (NPX) as an NSAIDs model (NPX-MEs). Phase diagrams were constructed for MEs composed of tween 80, span 80, and propylene glycol (PG) as surfactant (S)/cosurfactant (CS), transcuto[®] P (TRC-P), and Labrafac[™] PG as oil. The final concentration of NPX in MEs was 1% (w/v). The MEs were analyzed for particle size, refractive index, and viscosity. In vitro permeability studies of NPX-MEs were conducted using Franz diffusion cells on rat skin samples. Additionally, the effects of Eucalyptus oil (EU oil), oleic acid (OLA), and TRC-P as enhancers on the skin permeation of NPX were investigated. The particle size and viscosity values of the NPX-MEs ranged from 7.05 ± 0.03 to 79.56 ± 0.58 nm and 222.4 ± 0.87 to 681.13 ± 1.97 , respectively. The optimal formulation, ME-3, consisted of 20% oil, 10% water, and 70% S/C phases. The skin permeation rates of NPX from ME-3 were higher than those of other formulations ($D_{app} = 1.36 \pm 0.616$, $ER_D = 527.989 \pm 313.627$) with a lower lag time. Additionally, OLA-treated skin showed the highest transdermal permeation rate ($ER_D = 75.55 \pm 23.532$). Based on these results, the formulated NPX-ME may be a desirable carrier for transdermal delivery compared to traditional formulations, potentially reducing side effects and improving the therapeutic efficacy of NPX.

Keywords: Naproxen, Microemulsion, Transdermal Drug Delivery, Skin Permeation

1. Background

As one of the popular NSAIDs, NPX has been widely used to treat pain, arthritis, and inflammatory disorders. However, side effects of traditional administration of NSAIDs, particularly NPX, including poor aqueous solubility and permeability properties, might limit their clinical applications (1-3). Many studies have focused on addressing these problems through the development of novel, highly efficient vehicles for optimized NSAIDs delivery. Hence, to overcome the aforementioned drawbacks, drug delivery nanocarriers such as MEs, lipid-based, polymeric-based materials, and others prove to be effective solutions. These carriers can address various pharmaceutical limitations, such as

solubility, permeability, bioavailability, and reduced side effects, which are among the main challenges of NSAIDs (4, 5).

One of the most recent attempts in NPX delivery is the development of transdermal-based formulations to optimize its solubility and skin permeation, which may influence clinical translation. Moreover, transdermally administered NPX, compared to other routes, remarkably improves therapeutic effects (2, 3, 6). Overall, transdermal delivery of drugs, compared to oral and intravenous routes, offers several benefits, including efficient penetration into deeper layers of the skin, non-invasive application, reduced systemic side effects (such as gastric disorders), avoidance of pre-systemic metabolism in the liver, targeted delivery,

prolonged drug release profile, and ease of use. Nevertheless, the permeability issue of drug molecules through the skin barrier poses a significant challenge (4, 5, 7).

ME-based carriers have recently attracted attention in preparing transdermal formulations for drug delivery due to their tunable physicochemical properties, such as droplet size and morphology, homogeneity, surface area, stability, and permeability. MEs, thermodynamically stable liquids, are composed of aqueous nanodroplets encapsulated within a mixture of surfactant and co-surfactant, dispersed in an oil phase. Additionally, by adjusting the concentration and nature of the organic and aqueous phases, the physicochemical properties of the droplets can be easily modified. In this context, MEs are recognized as promising materials for application in the fabrication of nanocarriers, particularly for skin vehicles, due to their unique structural properties. Moreover, the relatively straightforward preparation and scalability of MEs make them attractive for biomedical applications (8-11).

MEs have great potential to be used as transdermal carriers to increase the solubility of both hydrophobic and hydrophilic drugs, while simultaneously enhancing skin permeability (12-14). The main advantages of ME-based transdermal carriers, compared to other conventional formulations of NSAIDs, include local and controllable drug release, as well as sufficient distribution of drugs into various layers of the skin, resulting in reduced side effects. Hence, MEs, as transdermal carriers, can offer favorable physicochemical properties, especially in terms of skin permeability (15, 16). However, the use of large amounts of S/CS, phase inversion, and the selection of safe excipients for pharmaceutical application pose major disadvantages from both formulation and biocompatibility perspectives (17). Therefore, new assessments are necessary for MEs to guide their fabrication towards optimal formulations in pharmaceutical applications, particularly in transdermal formulations.

There are very few studies on ME formulation for NPX. In prior studies, Salimi et al. characterized and evaluated the skin penetration of NPX on various components (e.g., Tween 80, Span 80, PG, TRC-P, and +LabrafacTM PG) of prepared MEs.

2. Objectives

Herein, we prepared a series of MEs (W/O) containing NPX, which could improve the permeability of the corresponding NPX in a rat skin model (18-20). Meanwhile, the permeation parameters such as steady-state flux (J_{ss}), permeability coefficient (P), lag time (T_{lag}), apparent diffusion coefficient (D_{app}), and enhancement ratio (ER_{flux} , ER_D , and ER_p) of NPX permeated were investigated. The permeation analysis was evaluated using a Franz diffusion cell set with rat skin. Furthermore, the skin permeability potential of MEs was evaluated with different enhancers including EU oil, OLA, and TRC-P.

3. Methods

3.1. Materials

Naproxen ($w \geq 99\%$) was purchased from Pars Darou Co, IRAN. Span 80, Tween 80, Oleic acid, and Propylene glycol (PG) were supplied by Merck Co, Germany. Propylene glycol dicaprylocaprate (LabrafacTM PG) and Diethylene glycol monoethyl ether (Transcutol P) were received as gift samples from GATTEFOSSÉ, France. EU oil was purchased from Barij Essence Co, IRAN. All materials were of reagent grade and applied without further purification.

3.2. Animals

The skin permeation of NPX-MEs was investigated on Wistar rats. The male rats weighing 200-250 g (8 - 10 weeks old) were supplied by the Animals Laboratory, Ahvaz Jundishapur University of Medical Sciences. The experimental protocol was approved by the Animal Ethics Committee of Jundishapur University of Medical Sciences, Faculty of Pharmacy (Approval No. N 75).

3.3. MEs Region Identification

The concentration of the different constituents in the MEs can be chosen according to the water titration method using pseudo-ternary phase diagrams. Here, two phase diagrams were first prepared by accurately weighing Span 80 / Tween 80 (1.1) as surfactant and PG as co-surfactant. One contained a 4: 1 weight ratio, and the second contained a 6: 1 weight ratio of S/C mixture. EU oil, OLA, and TRC-P (+ LabrafacTM PG) (1.10) were selected as oil phases and were accurately weighed in the vial.

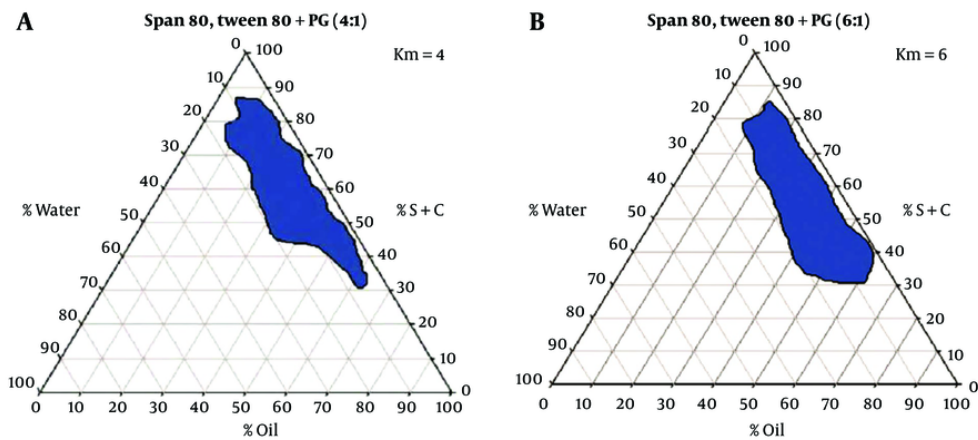


Figure 1. The pseudo-ternary phase diagrams of the water-oil-S/C mixture system at the 4: 1 and 6: 1 weight ratio of Span 80 /Tween 80/ PEG at ambient temperature, dark area show MEs zone

For the preparation of each phase diagram, the mixture of S/C was then added to the oil phase at weight ratios of 1: 9, 2: 8, 3: 7, 4:6, 5: 5, 6:4, 7: 3, 8: 2, and 9: 1. The resulting S/C-oil mixture was homogeneously mixed, and then deionized water was added under continuous titration until a transparent dispersion was formed. The transparent monophasic liquids were classified as MEs. The MEs were formed spontaneously at room temperature. Pseudo-ternary phase diagrams were investigated to study the region of MEs existence as well as the quaternary system of water/ Labrafac™ PG, Transcutol P (10: 1)/ Span 80-Tween 80/ PG. These diagrams were previously reported in our studies, and the results are shown in Figure 1 (21).

3.4. Preparation of NPX-MEs

NPX-MEs were constructed following the same formulations used in our previous works (21). A full factorial design, as a convenient method to prepare MEs, was used with three variables at two levels. The percentages of oil and water phases and S/C ratio have been proposed as the main variables, and eight different formulations of MEs were selected from the phase diagrams with low and high levels of oil (20% and 40%), water (5%, 10%), and S/C ratio (4: 1, 6: 1). NPX-MEs formulations were prepared as follows in Table 1. The oil phase contains 1% of NPX, and the S/C mixture was separately prepared and then mixed. Afterward, a

certain amount of deionized water was added dropwise into the previous mixture under stirring at room temperature until a uniform phase was formed.

Table 1. Components of the Selected NPX-MEs

Batch No.	Factorial	S/C	% Oil	%(S + C)	% Water
ME-1	+++	6:1	40	50	10
ME-2	++-	6:1	40	55	5
ME-3	++	6:1	20	70	10
ME-4	+--	6:1	20	74	5
ME-5	--	4:1	20	75	5
ME-6	-+-	4:1	20	70	10
ME-7	+-	4:1	40	55	5
ME-8	-++	4:1	40	50	10

3.5. Particle Size

All prepared NPX-MEs were characterized by dynamic light scattering (DLS) using SCATTER SCOPE 1 QUIDIX (South Korea) to measure the particle size of droplets.

3.6. Refractive Index

The refractive index measurements were conducted using a refractometer instrument (Quarts, Ceti, Belgium).

3.7. Viscosity

The viscosity of NPX-MEs was measured using a Brookfield viscometer (DV-II + Pro, Brookfield, USA). For

the viscosity evaluation of ME-NPX formulations, 10 mL of sample was poured into a beaker, and the viscosity was estimated using Spindle number 34 at room temperature with a shear rate of 100 rpm.

3.8. Analytical Method of NPX

A UV assay method for NPX was validated, and the quantitative analysis of NPX was estimated using a UV-Visible spectrophotometer (BioWaveII, WPA, England) at $\lambda_{\text{max}} = 271$ nm.

3.9. Ex vivo Permeation Study

Skin samples were obtained from male rats. Immediately after shaving the hair, abdominal skin was excised, and the subcutaneous fatty tissue was separated. The hairless skin was then cut into smaller pieces, wrapped in aluminum foil, and stored at -20°C until use. All permeation experiments were performed using Franz cells with a diffusional area of 4.9 cm^2 . All skin samples were hydrated in distilled water before permeation studies and checked for integrity. The thickness of skin samples was 0.055 ± 0.005 cm. Phosphate buffer saline (PBS; pH 7.4) was used as the receptor medium. Each skin sample was placed between the donor and receptor chambers so that the stratum corneum of the skin surface was exposed to the donor solution. Then, 23 mL of PBS was poured into the receptor chamber. The donor chamber was filled with the MEs formulation or a saturated solution of NPX as the control sample. The Franz diffusion cell was kept in an oil bath setup at 37°C under stirring. At specific interval times, 2 mL of samples were extracted from the receptor chamber up to 56 h for UV assay analysis at 271 nm. To maintain sink conditions after every sample withdrawal, the receptor chamber was filled with 2 mL of fresh PBS. Additionally, EU oil, OLA, and TRC-P were used as enhancers to increase skin permeability. The effect of these enhancers was also evaluated through rat skin. For this purpose, the hydrated skin samples were treated with 1 mL of each enhancer in the donor chamber before the addition of the MEs formulation. After 2 h, both the donor and acceptor chambers were washed with distilled water, and the experiment was performed following the same procedure as described above.

3.10. Statistical Data Analysis

To calculate the permeation parameters of NPX, the cumulative mass of released NPX per unit surface area of the rat skin versus time was first plotted. The J_{ss} ($\text{mg}/\text{cm}^2\cdot\text{h}$) value through the rat skin into the acceptor medium was determined by the slope of the linear portion of the mentioned plot. The permeability coefficient includes P (cm/h), D_{app} (cm^2/h), and ER parameters (denoted as the enhancer effect on the enhancement of NPX permeation) were calculated using the following Equations ((22, 23):

$$J_{ss} = P \cdot C \quad (1)$$

$$D = \frac{h^2}{6T_{lag}} \quad (2)$$

$$ER = \frac{J_{ss}P, D \text{ after treatment}}{J_{ss}P, D \text{ before treatment}} \quad (3)$$

Where C , T_{lag} , and h were the total amount of permeated drug in the donor phase, the needed time to reach a steady state, and the length of the diffusion pathway, respectively.

All data are reported as the mean \pm standard deviation (SD) from triplicate independent experiments for each sample. The statistical differences between experimental formulations were obtained by Analysis of Variance (ANOVA) using GraphPad Software. P-values less than < 0.05 were considered significant.

4. Results and Discussion

4.1. Characterization Studies

The results of the characterization of MEs formulations are listed in Table 2. The findings show that the average particle size values of the NPX-MEs range from 7.05 ± 0.03 to 79.56 ± 0.58 nm, which were in the nanoscale range. Also, the refractive index for the NPX-MEs was about 1.45, near to the oil phase, suggesting water-in-oil structures of formulations. The obtained viscosity values were in the range of 222.4 ± 0.87 to 681.13 ± 1.97 cp. Moreover, the correlation between the independent variables such as %w, % oil, and S/C, and particle size, refractive index, and viscosity values for NPX-MEs were analyzed and expressed using Equations 4 (Table 3).

Table 2. Particle Size, Refractive Index and Viscosity of NPX-MEs (n = 3)^a

Batch No.	Particle Size, nm	Refractive Index	Viscosity, cp
ME-1	7.05 ± 0.03	1.445 ± 0.14	251.3 ± 1.91
ME-2	79.56 ± 0.58	1.451 ± 0.15	222.4 ± 0.87
ME-3	53.63 ± 0.42	1.453 ± 0.2	652.63 ± 1.22
ME-4	10.26 ± 0.15	1.452 ± 0.18	681.13 ± 1.97
ME-5	21.96 ± 0.31	1.456 ± 0.16	319.4 ± 1.37
ME-6	34.4 ± 0.41	1.452 ± 0.21	341.8 ± 0.61
ME-7	10.43 ± 0.12	1.451 ± 0.19	364.16 ± 1.60
ME-8	11.6 ± 0.3	1.444 ± 0.17	475.9 ± 1.83

^a Values are expressed as mean ± SD.

Table 3. The Statistical Relationship Between Independent Variables with Particle Size, Refractive Index and Viscosity

Dependent Variables	Independent Variable	P-Value	Significant
Particle Size	% oil	0.880	No
	% w	0.770	No
	S/C	0.697	No
Refractive Index	% oil	0.970	No
	% w	0.001	Yes
	S/C	0.001	Yes
Viscosity	% oil	0.210	No
	% w	0.009	Yes
	S/C	0.576	No

According to Equation 4, there were no significant differences between the independent variables and average particle size values of the NPX-MEs. In the case of refractive index, both the S/C ratio and % w phase indicated a significant effect (P-value = 0.001); thereby, the refractive index of NPX-MEs decreases with the increase in % water phase or S/C ratio. The viscosity is affected by % w phase (P-value = 0.009), so the viscosity increases with the decrease in % w phase. Through the change of various parameters, including the oil phase, S/CS mixture, and weight ratio of excipients, MEs with different physicochemical properties can be formulated. In this context, Salimi et al. (24) prepared MEs formulations via a pseudo-ternary phase diagram-assisted full factorial method, using Tween-80 and Span 20 as S, PG as CS, TRC-P, and OLA as the oil phase to enhance transdermal delivery of meloxicam. In this study, the droplet size (5.24 to 16.75 nm) and viscosity (132 to 418 cp) of MEs were smaller compared to our study. They seemed to be influenced by the oil phase type (mixture of TRC-P/OLA) and the ratio of S/CS (1:1 and 3:1). Additionally, they found that the particle size was not dependent on the independent variables, coherent

with the present study. For another example, Moghimipour et al. (21) fabricated quercetin-loaded MEs composed of Tween-80, Span 20, PG, TRC-P, and OLA with a size range of 5.31 - 26.07 nm and viscosity of 115 - 361 cp for transcorneal delivery application. It seems that the decreased particle size resulted in increased surface area due to the different weight ratio of MEs components. Similarly, the relationship between the droplet size and independent variables was not significant. In another study, Salimi et al. (25) developed an ME formulation for transfollicular delivery of adapalene. MEs were fabricated using TRC-P, OLA, PG, Tween-80, and Span 20. These MEs had a droplet size of 13.86 to 56.16 nm, and the viscosity ranged from 117 to 240 cp. Furthermore, statistical differences were observed between the viscosity parameter and % w, S/CS, and oil%. These findings suggest that the physicochemical properties of MEs can be affected by constituents and their weight ratio in the ME (26).

$$\text{Particle size} = 15.8 + 0.908 (\% \text{ oil}) + 0.6 (\% w) - 1.98 (S/C) \quad (4)$$

$$\text{Refractive Index} = 1.47 - 0.000001 (\% \text{ oil}) - 0.00112 (\% w) - 0.00204 (S/C) \quad (5)$$

$$\text{Viscosity} = 470 - 3.83 (\% \text{ oil}) - 34.1 (\% w) - 16.8 (S/C) \quad (6)$$

4.2. Ex-vivo Permeation Studies of NPX-MEs

Permeation of NPX from MEs carriers containing 1% NPX was evaluated using Franz diffusion cells with rat full-thickness skin. A saturated NPX solution served as the control sample. The permeation parameters and enhancement ratios of pre-prepared NPX-MEs and the control are shown in Table 4. Transdermal delivery revealed a significant increase in permeated NPX with all MEs carriers. Among the various MEs, MEs-3, composed of 20% oil, 10% water phases, and 70% S/C ratio, exhibited the highest permeation rate. The initial time point of NPX detection ranged from 11.72 ± 1.532 to 20.42 ± 15.521 hours across different MEs formulations. In ME-3 formulation, ER_D (527.989 ± 313.627) notably increased compared to the control sample. These findings suggest that the higher percentage of S and C components in the prepared MEs may contribute to the increased permeation rate of NPX on the rat skin model. Surfactants, particularly, may enhance drug penetration into the intercellular lipid matrix of the stratum

corneum, thereby enhancing skin permeation. Additionally, the use of TRC-P and Labrafac™ PG can modify MEs formulations. Similar results were observed in previous studies on transdermal drug delivery of NSAIDs based on MEs carriers. Ustundag Okur et al. demonstrated that incorporating different components like Labrafil, Tween 80, and Transcutol P favored the formation of MEs as carriers for drug delivery of NSAIDs such as NPX, promoting skin permeation (19). Hu et al. reported MEs-based vehicles for the transdermal delivery of ibuprofen, which also increased ibuprofen permeation through rat skin without associated skin irritation risk (13). Overall, the observed results in skin permeation studies could be attributed to various effects: (1) enhanced local NPX availability incorporated into the hydrophobic domain of MEs carriers; (2) increased distribution of drug molecules into skin layers; (3) the nonionic and hydrophobic components of MEs interacting with lipidic components and disrupting the skin structure, particularly the stratum corneum layer (20, 27, 28). Additionally, ex vivo skin permeation profiles of NPX from ME formulations are shown in Figure 2. We observed that NPX release from all MEs was approximately 6-fold higher compared to the control sample after 56 hours. NPX, being a hydrophobic drug, benefits from the nanocarriers of developed MEs, which promote controlled local release into skin layers, providing wide drug distribution and increased solubility (29-31).

Table 4. The Permeation Parameters and Enhancement Ratio (ER) of NPX-MEs (n = 3)^a

Batch No.	J_{ss} , mg/cm ² .h	D_{app} , cm ² /h	P, cm/h	T_{lag} , h	ER _{flux}	ER _D	ER _P
Control	0.0033 ± 0.00011	0.0019 ± 0.00001	0.0032 ± 0.0061	17.96 ± 0.831	-	-	-
ME-1	0.011 ± 0.002	0.006 ± 0.002	0.011 ± 0.054	10.99 ± 0.368	3.293 ± 0.671	2.829 ± 0.118	3.325 ± 0.703
ME-2	0.016 ± 0.007	0.009 ± 0.007	0.016 ± 0.001	20.42 ± 0.521	4.758 ± 0.167	4.538 ± 0.371	4.763 ± 0.175
ME-3	0.018 ± 0.004	1.36 ± 0.616	0.017 ± 0.002	1.72 ± 0.532	5.414 ± 0.094	527.989 ± 0.627	5.425 ± 0.089
ME-4	0.017 ± 0.008	0.043 ± 0.032	0.017 ± 0.008	1.87 ± 0.243	5.030 ± 0.273	21.923 ± 0.149	5.035 ± 0.277
ME-5	0.010 ± 0.002	0.021 ± 0.012	0.01 ± 0.002	3.10 ± 0.691	2.909 ± 0.652	10.632 ± 0.117	2.912 ± 0.653
ME-6	0.017 ± 0.002	0.013 ± 0.0007	0.017 ± 0.0007	3.50 ± 0.194	5.071 ± 0.513	6.720 ± 0.378	5.075 ± 0.513
ME-7	0.008 ± 0.0008	0.113 ± 0.08	0.007 ± 0.0008	2.21 ± 0.809	2.346 ± 0.237	40.790 ± 0.749	2.271 ± 0.227

Batch No.	J_{ss} , mg/cm ² .h	D_{app} , cm ² /h	P, cm/h	T_{lag} , h	ER _{flux}	ER _D	ER _P
ME-8	0.017 ± 0.004	0.013 ± 0.004	0.017 ± 0.002	4.88 ± 0.380	5.232 ± 0.353	6.372 ± 0.906	5.234 ± 0.355

^a Values are expressed as mean ± SD.

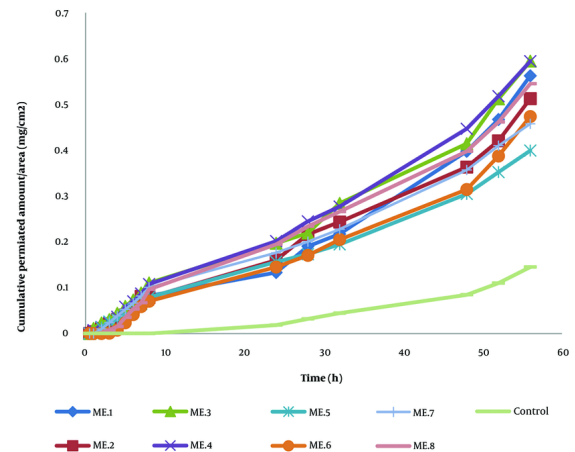


Figure 2. The graph of the cumulative amount of NPX passed through the surface unit in the whole abdominal skin of the rat by pharmaceutical MEs (mean ± SD) (n = 3).

4.3. Evaluation of the Main Variables' Effects on Permeation

In this section, the independent variables of oil (TRC-P + Labrafac™ PG), water phases, and S/C ratio (Span 80/Tween 80 and PEG-400) were investigated, and their effects on the permeation parameters of MEs are discussed. Linear equations 7-10 indicated the impact of these variables on the amount of permeated NPX. Regression analysis of these equations revealed no significant differences between J_{ss} , D_{app} , and P values for the main variables ($P > 0.05$) after ANOVA analyses. However, a statistically significant difference ($P < 0.05$) was found between the T_{lag} value and water phase, while other variables showed no significant difference with the T_{lag} value (Table 5).

$$J_{ss} = 0.00369 + 0.000141 (\%Oil) - 0.000377 (\%W) + 0/00183 (S/C) \quad (7)$$

$$J_{ss} = 0.00369 + 0.000141 (\%Oil) - 0.000377 (\%W) + 0/00183 (S/C) \quad (8)$$

$$D_{app} = 0.194 + 0.0068 (\%Oil) - 0.0294 (\%W) - 0/0620 (S/C) \quad (9)$$

$$P = 0.0214 - 0.000141 (\%Oil) + 0006 (\%W) + 0/00098 (S/C) \quad (10)$$

Table 5. The Statistical Relationship between Independent Variables with J_{ss} , T_{lag} , D_{app} and P

Depended Variables	Independent Variable	P-Value	Significant
J_{ss}	% oil	0.168	No
	% w	0.350	No
	S/C	0.077	No
T_{lag}	% oil	0.073	No
	% w	0.021	Yes
	S/C	0.574	No
D_{app}	% oil	0.381	No
	% w	0.345	No
	S/C	0.424	No
P	% oil	0.611	No
	% w	0.591	No
	S/C	0.722	No

4.4. Effect of Penetration Enhancers on the Skin Permeation of NPX-MEs

The ex vivo permeation of NPX was evaluated using EU oil, OLA, and TRC-P treated rat skin samples in a Franz diffusion cell, and the results are presented in Tables 6 and 7. All enhancer-treated skins exhibited better permeation compared to samples without any treatment. Among the enhancers, OLA demonstrated faster permeation than others ($ER_{flux} = 6.193 \pm 0.587$ and $ERD_{app} = 75.5 \pm 33.53$). This increase in the skin permeation of NPX could be attributed to OLA and EU serving as active vehicles for the preparation of MEs, as reported by Abd et al. (27). Other studies have shown that the higher permeation from OLA is likely due to its attractive features, including a cis isomer structure, unsaturated bond, and solubility ability in the skin lipid bilayers, leading to disruption in the lipidic structure of the outermost layer of skin and changes in skin functions, thereby favoring enhanced drug permeation (32-35).

Table 6. The Permeation Parameters of NPX-MEs for Enhancers (n = 3)^a

Batch No.	J_{ss} , mg/cm ² .h			D_{app} , cm ² /h			T_{lag} , h		
	EU Oil	TRC-P	OLA	EU Oil	TRC-P	OLA	EU Oil	TRC-P	OLA
ME-1	0.04 ± 0.0022	0.039 ± 0.003	0.021 ± 0.0013	0.071 ± 0.0029	0.075 ± 0.0007	0.372 ± 0.006	1.209 ± 1.2	0.523 ± 0.0003	0.083 ± 0.11
ME-2	0.036 ± 0.001	0.035 ± 0.001	0.030 ± 0.005	0.113 ± 0.0016	0.092 ± 0.01	0.011 ± 0.0018	1.064 ± 1.002	0.692 ± 0.0002	2.431 ± 1.56
ME-3	0.028 ± 0.003	0.038 ± 0.002	0.023 ± 0.001	0.076 ± 0.0024	0.064 ± 0.0031	0.005 ± 0.0021	0.812 ± 0.49	0.504 ± 0.027	3.5 ± 1.1

Batch No.	J_{ss} , mg/cm ² .h			D_{app} , cm ² /h			T_{lag} , h		
	EU Oil	TRC-P	OLA	EU Oil	TRC-P	OLA	EU Oil	TRC-P	OLA
ME-4	0.042 ± 0.008	0.044 ± 0.022	0.043 ± 0.011	0.016 ± 0.001	0.031 ± 0.002	0.023 ± 0.0012	2.316 ± 0.54	1.691 ± 0.062	1.81 ± 0.57
ME-5	0.027 ± 0.0001	0.019 ± 0.0016	0.022 ± 0.0014	0.089 ± 0.0019	0.055 ± 0.0021	0.096 ± 0.003	1.026 ± 0.51	0.792 ± 0.41	0.507 ± 0.24
ME-6	0.036 ± 0.0002	0.017 ± 0.0012	0.021 ± 0.0011	0.423 ± 0.00023	0.074 ± 0.0011	0.029 ± 0.0011	0.132 ± 0.12	0.292 ± 0.12	0.736 ± 0.0125
ME-7	0.029 ± 0.001	0.019 ± 0.0013	0.048 ± 0.0021	0.311 ± 0.19	0.011 ± 0.0018	0.021 ± 0.001	0.326 ± 0.005	4.165 ± 1.6	2.029 ± 0.55
ME-8	0.03 ± 0.005	0.024 ± 0.0016	0.019 ± 0.0016	0.107 ± 0.072	0.017 ± 0.025	0.034 ± 0.0014	0.563 ± 0.125	2.393 ± 1.52	0.819 ± 0.0032

^a Values are expressed as mean ± SD.

Table 7. Comparison of Permeability Parameters of NPX-MEs With and Without Enhancer (n = 3)^a

Batch No.	EU Oil		TRC-P		OLA	
	ER_{flux}	ER_D	ER_{flux}	ER_D	ER_{flux}	ER_D
ME-1	3.794 ± 0.700	14.734 ± 6.448	3.284 ± 0.674	15.284 ± 6.783	1.935 ± 0.357	75.55 ± 23.532
ME-2	2.681 ± 0.344	55.009 ± 45.795	2.628 ± 1.318	45.148 ± 37.097	2.270 ± 1.138	5.403 ± 4.439
ME-3	1.594 ± 0.0364	3.127 ± 2.895	2.200 ± 0.503	2.620 ± 2.426	1.317 ± 0.301	0.199 ± 0.185
ME-4	3.077 ± 0.862	0.54 ± 0.0380	3.262 ± 1.973	1.050 ± 0.739	3.173 ± 1.92	0.782 ± 0.551
ME-5	2.953 ± 0.668	6.33 ± 5.401	2.080 ± 0.471	3.889 ± 3.319	2.317 ± 0.524	17.675 ± 16.83
ME-6	2.136 ± 0.209	32.173 ± 1.813	1.029 ± 0.101	5.612 ± 0.316	1.251 ± 0.122	2.231 ± 0.126
ME-7	3.892 ± 0.352	14.655 ± 11.417	2.503 ± 0.237	0.51 ± 0.398	6.193 ± 0.587	0.527 ± 0.813
ME-8	1.793 ± 0.497	9.055 ± 2.701	1.453 ± 0.403	1.420 ± 0.424	1.372 ± 0.325	2.906 ± 0.867

^a Values are expressed as mean ± SD.

5. Conclusions

In this study, we evaluated the skin permeation of NPX using a series of W/O NPX-MEs successfully formulated and characterized in the present work. The optimal NPX formulation was selected based on permeability parameters and permeation rate. According to the permeability studies, the best formulation (ME-3) consisted of 20% TRC-P and Labrafac™ PG as oil, 10% water, and 70% of Tween 80, Span 80, and PG as S/C phases. Moreover, the skin permeation rate of ME-3 was higher compared to others. These drug-encapsulated MEs, which are favorable materials easily distributed on the skin layer, provide a desirable enhancement of drug penetration. Additionally, oil enhancers such as OLA can increase the skin permeation rate due to disruption in the lipid

depended on the S/C ratio, amount of aqueous/organic phases, and type of enhancer. Furthermore, transdermal delivery studies on rat skin samples indicated a remarkable increase in the permeation of NPX for both MEs formulations and the selected enhancers, which is advantageous in the therapeutic functionality of a MEs-based nanocarrier for the topical delivery of drugs.

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Footnotes

Authors' Contribution: A. S. conceived and designed the evaluation and drafted the manuscript. A. S. participated in designing the evaluation, performed parts of the statistical analysis, and helped to draft the manuscript. E. M. and N. J. re-analyzed statistical data and revised the manuscript. A. S., N. H. approved the final version to be published.

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Data Availability: The dataset presented in the study is available on request from the corresponding author during submission or after publication.

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