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Anticholinesterase, Antioxidative and Neuroprotective Effects of Hydroethanolic Extracts of Guiera Senegalensis J. F. Gmel. Leaves on Scopolamine Induced Alzheimer's Disease in Wistar Rat Models

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Abstract

Background: Medicinal plants like Guiera senegalensis J. F. Gmel. (GS) have gained attention for their potential neuroprotective effects due to their rich composition of bioactive compounds, including flavonoids and polyphenolic acids. While previous studies have highlighted the antioxidant, antiinflammatory, and neuroplasticity-enhancing properties of GS extract, its efficacy in mitigating dementia-related pathology remains to be fully understood.

Objectives: This study aimed to investigate the neuroprotective effects of hydroethanolic GS extract against scopolamine (Sco)-induced dementia in Wistar rats, focusing on its impact on cholinergic function, oxidative stress, neuroinflammation, neurodegeneration, and memory impairment.

Methods: Fresh leaves were processed for extraction using standard methods. Antioxidant activity was evaluated using FRAP and DPPH assays. Adult male Wistar rats were used for behavioral tests (Y-maze, NOR, MWM) and biochemical analyses, including ELISA for cholinergic activity, oxidative stress markers (Aβ1- 42, phosphorylated Tau), pro-inflammatory cytokines (IL-1β, TNF-α, IL-6, IFN-γ), GFAP, BDNF, and IL-10. Brain tissues underwent histopathological examination. Data were analyzed using GraphPad Prism software.

Results: The study assessed the antioxidant potential of GS extract and found that it significantly scavenged DPPH radicals (70.29%) and reduced Fe $^{3+}$ (49.69%). Behavioral tests showed that GS extract (100 - 400 mg/kg) improved spatial memory and learning in Sco-treated rats. Y-maze results indicated increased spontaneous alternation with GS extract (P < 0.01). NOR and MWM tests showed improved memory and learning, with GS extract-treated rats spending more time in the target quadrant. Biochemical analysis revealed that GS extract increased acetylcholine (ACh), Superoxide Dismutase (SOD), Catalase (CAT), Glutathione (GSH), IL-10, and BDNF levels, while decreasing acetylcholinesterase (AChE), malondialdehyde (MDA), nitrite, Aβ1-42, phosphorylated Tau, IL-1β, TNFα, IL-6, IFN-γ, and GFAP levels in the hippocampus. Histological analysis confirmed restored hippocampal tissue architecture in AD rats treated with GS extract.

Conclusions: Our findings suggest that GS modulates cholinergic, antioxidant, anti-inflammatory, and neuroplasticity pathways, exerting beneficial effects on cognitive functions and biochemical markers associated with Alzheimer's disease (AD) pathology. These results indicate the potential of GS as a neuroprotective agent for the treatment of AD.

Keywords: Guiera Senegalensis (GS), Alzheimer's Disease (AD), Neuroprotective, Cholinesterase Inhibitors, Antioxidants, Antiinflammatory

1. Background

Dementia currently affects 57.4 million people worldwide, with estimates reaching 152.8 million by 2050 ([1\)](#page-13-2). Alzheimer's disease (AD), prevalent in the elderly, is characterized by cognitive dysfunction, including impairments in learning and memory ([2,](#page-13-0) [3](#page-13-1)). Alzheimer's disease pathophysiology involves amyloid-β

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(Aβ) accumulation, cholinergic dysfunction, Tau hyperphosphorylation, oxidative stress, and neuroinflammation [\(4](#page-13-3)-[6\)](#page-13-4). Key factors include microglial activation and elevated levels of proinflammatory cytokines (IL-1β, TNF-α, IL-6, IFN-γ), which disrupt cholinergic function and lead to neuronal loss [\(7-](#page-13-5)[9\)](#page-13-6). Increased levels of glial fibrillary acidic protein (GFAP), resulting from astrocyte activation, are indicative of neuronal damage [\(10](#page-13-7), [11](#page-13-8)), while reduced brain-derived neurotrophic factor (BDNF) levels are associated with cognitive decline [\(12,](#page-13-9) [13](#page-13-10)).

Scopolamine (Sco) induces amnesia in rodents by blocking muscarinic receptors, resulting in cognitive deficits, increased acetylcholinesterase (AChE) activity, decreased acetylcholine, neuroinflammation, and elevated GFAP levels [\(9,](#page-13-6) [14-](#page-13-11)[21\)](#page-13-12). It also contributes to neurodegeneration through amyloid peptide accumulation and Tau hyperphosphorylation ([9](#page-13-6), [14,](#page-13-11) [22-](#page-13-13) [24](#page-14-0)).

Current AD treatments, such as AChE inhibitors like galantamine and donepezil, can cause side effects ([25\)](#page-14-1). Therefore, research is focusing on safer, multitarget plant-based therapies ([26](#page-14-2), [27\)](#page-14-3). Guiera senegalensis (GS), native to West and Central Africa, has shown promise in treating neurological conditions due to its antimicrobial, anticancer, antioxidant, and antiinflammatory properties ([28-](#page-14-4)[36](#page-14-5)). Its neuroprotective effects have been demonstrated in zebrafish models, and further research in rodent models is ongoing to evaluate its efficacy in AD [\(37,](#page-14-6) [38](#page-14-7)).

2. Objectives

This research aimed to explore the anticholinesterase, antioxidant, and anti-inflammatory activities of the hydroethanolic extract of Guiera senegalensis J. F. Gmel. leaves in rats with Sco-induced dementia. The effectiveness of the treatment was assessed through behavioral tests, including the Y-maze, novel object recognition (NOR), and Morris Water Maze (MWM), which evaluate learning and memory impairments. Additionally, factors such as cholinergic dysfunction, oxidative stress, neuroinflammation, and neurodegeneration were examined using biochemical and histopathological analyses in the disease model.

3. Methods

3.1. Chemicals and Antibodies

Thiobarbituric acid (TBA), Sco, hydrogen peroxide $(H₂O₂)$, adrenaline, and 5,5-dithiobis(2-nitro-benzoic acid) (DTNB) were obtained from Sigma-Aldrich, USA. Donepezil (Aricept®) was acquired from Pfizer, USA. ELISA kits for acetylcholine (ACh) and AChE were from BioVision, India, and Elabscience, China, respectively. ELISA kits for BDNF, amyloid-β peptide (Aβ1-42), and inflammatory markers (TNF-α, IL-1β, IL-10, IL-6, IFN-γ) were from Cusabio, China. ELISA kits for Tau phospho protein and GFAP were from Biomatik, Canada.

3.2. Plant Collection and Processing

Fresh GS leaves were collected in June 2021 from Touloum, Far North Cameroon, and deposited at the Herbier National du Cameroun as specimen No. 49837/HNC, corresponding to Sabatié B. No. 699 (G. senegalensis J. F. Gmel) ([29](#page-14-8), [39\)](#page-14-9). The leaves were shadedried for one week, ground into powder, and macerated for 72 hours in 80% ethanol and 20% distilled water. After filtering through Whatman paper, the extract was evaporated using a rotavapor at 80°C to remove ethanol, followed by drying in an oven at 50°C for 48 hours. The hydroethanolic extract yielded 7.67% (v/v) ([29](#page-14-8), [39\)](#page-14-9).

3.3. In Vitro Antioxidant Activity of Hydroethanolic G. Senegalensis J. F. Gmel Leaves Extract

The in vitro antioxidant activity of GS leaves extract was evaluated by determining its ferric reducing potential and radical scavenging activity using the Ferric Reducing Antioxidant Power (FRAP) and 2,2- Diphenyl-1-picrylhydrazyl (DPPH) assays, respectively $(40, 41)$ $(40, 41)$ $(40, 41)$ $(40, 41)$.

3.4. Animal Models

Adult male Wistar rats (2 - 3 months old) were obtained from the Biophysics and Biochemistry Laboratory at Cameroon University. They were housed in six polyacrylic cages, with 5 rats per cage, and acclimated for 7 days in a controlled environment (25 \pm 2°C, 12-hour light/dark cycles). The rats were provided with standard food and water, with a 12-hour fast before treatment and 7 hours after treatment, while water was continuously available.

3.5. Experimental Design

The six cages ($n = 5$) represented six groups of rats [\(Figure](#page-2-0) 1) Two of these groups were randomly selected as controls, one as a model, and the other three as experimental groups. The treatment protocol for each group is described below.

- Group I (control): Rats received distilled water (10 mL/kg, p.o.) via oral gavage and 0.9% NaCl (10 mL/kg, i.p.).

- Group II (model): Rats received distilled water (10 mL/kg, p.o.) via oral gavage and Sco (1 mg/kg, i.p.).

- Group III (positive control): Rats received donepezil (2 mg/kg, p.o.) via oral gavage and Sco (1 mg/kg, i.p.).

- Groups IV-VI (experimental): Sco (1 mg/kg, i.p.) induced rats were treated with 100, 200, and 400 mg/kg of GS leaves extract, respectively, administered via oral gavage.

For administration, Sco was dissolved in saline (0.9% NaCl), and donepezil and the plant extract were dissolved in distilled water, with each rat receiving 0.1 mL/10 g. Distilled water, plant extract, or donepezil were administered orally once daily for 14 days, while Sco or saline was given intraperitoneally for 7 days (starting from day 8). Doses were based on the literature: Sco at 1 mg/kg ([4,](#page-13-3) [14,](#page-13-11) [42\)](#page-14-12); donepezil at 2 mg/kg ([42\)](#page-14-12). Cognitive functions were assessed 30 minutes after Sco or saline and 1 hour after the other treatments. The study adhered to the Cameroon Bioethics Committee and NIH guidelines, with approval from the Ethics Consultative Commission of Maroua University (Ref. No. 14/0261/Uma/D/FS/VD-RC).

3.6. Behavioral Studies

3.6.1. Y-maze Test

The Y-maze test assesses spatial short-term memory in rodents by measuring spontaneous alternations in a three-arm maze ([43,](#page-14-13) [44\)](#page-14-14). Each arm measured 33 cm \times 11

 $cm \times 12$ cm and was arranged at 120 $^{\circ}$ angles. Rats were placed at the end of one arm and allowed 8 minutes to explore. To minimize olfactory cues, the maze was cleaned with 70% ethanol between trials. Arm visit sequences were recorded during the final 5 minutes of the test. Locomotor activity was measured by counting the number of arm entries, and spontaneous alternations (consecutive entries into all three arms) were calculated as a percentage [\(20\)](#page-13-14).

$$
\%~alternation = \left[\frac{Number~of~alternations}{Total~number~of~arms~entries-2}\right]\!\!\!\left(\!\check{\gamma}\right)\!100
$$

3.6.2. Test of Novel Object Recognition

The NOR test followed a standard protocol in a 72 cm x 72 cm x 36 cm open-field setup. It was conducted over three days, with each day dedicated to the habituation, training, and retention phases ([2](#page-13-0)). On day 1 (habituation), rats were allowed to explore the empty field for 10 minutes. On day 2 (training), two identical objects were placed in the field, and the rats explored for 5 minutes. On day 3 (retention), one object was replaced with a novel object, while all other conditions remained the same. The setup and objects were cleaned with 70% ethanol between sessions. Exploratory behavior was recorded as the time spent exploring the novel (N) and familiar (F) objects. The Discrimination Index (DI) was calculated as follows (47):

$$
DI = \frac{N - F}{N + F}
$$
 (2)

Figure 2. Effect of hydroethanolic GS leaves extract on spontaneous alternation in the Y-maze [tes](#page-14-16)t (A). Correlation analysis between spontaneous alternation and number of arm entries (Pear-son's correlation) (B). Results are presented as mean ± SEM, (n = 5). 88 P < 0.01 significant difference from control group; * P < 0.05 and ** P < 0.01 significant
difference from model group; Sco: Sco-polami

3.6.3. Test of Morris Water Maze

Standard MWM protocols were used to assess hippocampal-dependent spatial learning and memory in rats [\(45,](#page-14-15) [46\)](#page-14-16). The maze, a metallic cylinder with a 79 cm diameter, was divided into four quadrants (North, South, East, West) and filled with opaque water mixed with non-toxic dye. The experiment, spanning eight days, included four phases:

- Phase 1 (days 1 - 3): Rats underwent four trials each day, with the escape platform moved to different quadrants every 60 seconds. Rats that did not find the platform within 60 seconds were guided to it and left there for 20 seconds to learn its location.

- Phase 2 (days 4 - 6): The escape platform was relocated to a different quadrant, and the same protocol was followed.

- Phase 3 (day 7): Rats were placed randomly in any quadrant with its designated escape platform, and the time spent in each quadrant was recorded.

- Phase 4 (day 8): The escape platform was placed in one of the two previously unused quadrants, and the escape latency was recorded for each trial.

3.7. Biochemical Studies

Rats were euthanized using intraperitoneal injections of ketamine/diazepam (50 mg/kg / 10 mg/kg) ([46\)](#page-14-16). After decapitation, the brains were removed, and the hemispheres were separated. One hemisphere was fixed in 10% formaldehyde for hippocampal histology,

while the other hemisphere was homogenized in 10% (w/v) ice-cold phosphate buffer (0.2 M; pH 7.4), centrifuged at 3000 rpm for 15 minutes, and the supernatant was stored at -20°C. The supernatant was used to measure Superoxide Dismutase (SOD) and Catalase (CAT) activities, as well as levels of ACh, AChE, Glutathione (GSH), Malondialdehyde (MDA), nitrite, Interleukin-1 beta (IL-1β), Tumor Necrosis Factor-alpha (TNF-α), BDNF, Interferon-gamma (INF-γ), Interleukin-10 (IL-10), Interleukin-6 (IL-6), Amyloid-beta 1-42 (Aβ1-42), Tau phospho protein, and GFAP.

3.7.1. Determination of Cholinergic Activity

Acetylcholine and AChE levels were quantified using commercially available ELISA kits, following the manufacturer's protocol, and the results were expressed in pg/mL.

3.7.2. Determination of Oxidative and Nitrosative Stress

Oxidative stress was evaluated by measuring SOD [\(47](#page-14-17)) and CAT activities, as well as GSH (48) (48) and MDA (49) (49) levels. The nitrite level was determined using the Griess method (50) (50) .

3.7.3. Determination of Neurological Inflammation and Damage

The expression of markers for inflammation (IL-1β, INF-γ, IL-6, IL-10, and TNF-α), neuronal damage (BDNF, GFAP), and AD (Aβ1-42, tau phospho protein) were assessed using commercially available ELISA kits,

Figure 3. Effect of hydroethanolic extract of GS leaves on the exploratory time (A) and the discrimination index (B) in the novel object recognition (NOR) test. Results are represented as mean ± SEM, (n = 5). 888 P < 0.001 significant difference from control group; * P < 0.05 and *** P < 0.001 significant difference from the model group; Sco:
Scopolamine1mg/kg,Don:Donepezil2mg/kg,GS:Concentra

following the manufacturer's protocol, and their concentrations were expressed in pg/mL.

3.8. Histopathological Study and Statistical Analysis

Rat brain tissues were fixed in 10% paraformaldehyde, embedded in paraffin, and sectioned at 5 μm thickness. The sections were stained with hematoxylin and eosin ([51](#page-14-21)) and examined under a microscope at 400X magnification. The number of healthy CA1, CA2, CA3, and dentate gyrus cells was quantified using ImageJ software.

Statistical analysis was performed using GraphPad Prism version 8.0.1. One-way ANOVA followed by Tukey post-hoc tests was used for Y-maze (alternation percentage), MWM (time spent in the target quadrant, latency), NOR (DI), and biochemical parameters. Twoway ANOVA with Bonferroni post-hoc tests was applied for object recognition test exploration time and hippocampal healthy cell counts. Data are presented as mean \pm SEM, and a P-value \leq 0.05 was considered statistically significant.

4. Results

4.1. Effect of Hydroethanolic Extract of Guiera Senegalensis J. F. Gmel. Leaves on Spatial Short-Term Memory in the Y-maze Test

Spontaneous alternations in the Y-maze test are represented in [Figure](#page-3-0) 2A. A significant difference between groups was observed $[F(5, 24) = 5.01; P = 0.003]$. Tukey's post-hoc analysis revealed a noteworthy increase in spontaneous alternation in the control group compared to the Sco-treated (model) group ($P < 0.01$). Additionally, rats pre-treated with the hydroethanolic GS leaves extract (100, 200, and 400 mg/kg) exhibited a significant increase in spontaneous alternation compared to the Sco-treated group ($P < 0.05$, $P < 0.01$, and $P < 0.05$, respectively). Notably, spontaneous alternation significantly increased in the group administered donepezil (P < 0.01). Pearson's correlation $(r = 0.321; P = 0.099)$ indicated no significant correlation between spontaneous alternation and the number of arm entries [\(Figure](#page-3-0) 2B).

4.2. Effect of Hydroethanolic Guiera Senegalensis J. F. Gmel. Leaves Extract on Long-Term Memory in the Novel Object Recognition Test

The results of the two-way analysis of variance, followed by Bonferroni post-hoc analysis, revealed a significant decrease in the time spent exploring the novel object in the Sco-treated group (model group) compared to the control group [F (5, 48) = 6.07; P < 0.001]. Conversely, the donepezil-treated group showed a significant increase ($P < 0.001$) in the time spent exploring the novel object [\(Figure](#page-4-0) 3A). The groups treated with hydroethanolic GS leaves extract at 100 and 200 mg/kg exhibited a significant increase (P < 0.05 and $P < 0.001$, respectively) in the time spent exploring the novel object compared to the Sco-treated group.

Figure 4. Effect of hydroethanolic GS leaves extract on the escape latency (A) and the time spent in target quadrant (B) in the MWM test. Results are represented as mean ± SEM, (n = 5). δ P < 0.05 and δδδ P < 0.001 significant difference from control group; * P < 0.05, ** P < 0.01 and *** P < 0.001 significant difference from model group; Sco: Scopolamine 1 mg/kg, Don: Donepezil 2 mg/kg, GS: Concentration of Guiera senegalensis leaves extract in mg/kg.

[Figure](#page-4-0) 3B demonstrates poor object recognition in the Sco-treated group compared to the control group, as indicated by a significant ($P < 0.001$) decrease in the DI. This impairment was significantly reversed $[F (5, 48) =$ 6.07; $P < 0.001$ by all doses of the hydroethanolic GS leaves extract, as well as in the donepezil-treated group.

4.3. Effect of Hydroethanolic Guiera Senegalensis J. F. Gmel. Leaves Extract on Visuospatial Learning and Memory in the Morris Water Maze Test

[Figure](#page-5-0) 4A shows the escape latency in the MWM test. Scopolamine significantly increased escape latency in the model group compared to controls $[F(5, 24) = 18.8; P$ < 0.001]. Pre-treatment with GS extract (100 and 200 mg/kg) and donepezil significantly reduced escape latency ($P < 0.001$).

[Figure](#page-5-0) 4B indicates that the time spent in the target quadrant was significantly reduced in the Sco group compared to controls [F (5, 24) = 4.69; P < 0.05]. GS extract (100, 200, and 400 mg/kg) significantly increased the time spent in the target quadrant ($P <$ 0.01, $P \le 0.05$, and $P \le 0.01$, respectively), as did donepezil ($P < 0.01$).

4.4. Effect of Hydroethanolic Guiera Senegalensis J. F. Gmel. Leaves Extract on Cholinergic Activity

To investigate the mechanism of GS in reducing Scoinduced memory impairment, cholinergic markers were measured. Scopolamine significantly reduced brain ACh levels in the model group compared to

controls [F $(5, 24) = 23.6$; $P < 0.001$]. Donepezil significantly increased ACh levels ($P < 0.001$). Guiera senegalensis extract pre-treatment (100, 200, 400 mg/kg) significantly increased ACh levels compared to the model group ($P < 0.01$, $P < 0.001$, and $P < 0.001$, respectively) ([Figure](#page-6-0) 5A).

Acetylcholinesterase levels were elevated in the model group [F (5, 24) = 21.9; $P < 0.001$], and donepezil also increased AChE levels (P < 0.001). GS extract (200 and 400 mg/kg) significantly reduced AChE levels ($P <$ 0.001, $P < 0.01$) compared to the model group. No significant difference was observed with the 100 mg/kg dose ([Figure](#page-6-0) 5B).

4.4.1. Malondialdehyde Levels

The MDA levels were notably higher [F $(5, 24) = 25$; P < 0.001] in rats treated with Sco (model group) compared to the control group. Conversely, the donepezil-treated group exhibited a significant reduction in MDA levels (P < 0.001). Treatment with GS extract at doses of 100 and 200 mg/kg mitigated Sco-induced lipid peroxidation, as evidenced by a significant reduction in MDA levels (P < 0.001) in these groups compared to the model group. However, no significant difference was observed between the rats receiving the extract at a dose of 100 mg/kg and the model group ([Figure](#page-7-0) 6A).

4.4.2. Nitrite Levels

As shown in [Figure](#page-7-0) 6B, nitrite levels significantly increased in the model group $[F(5, 24) = 28.9; P < 0.001]$

F**igure 5.** Effect of hydroethanolic GS leaves extract on hippocampus acetylcholine (A) and acetylcholinesterase (B) levels. Results are represented as mean ± SEM, (n = 5). 888 P <
0.001 significant difference from control Concentration of Guiera senegalensis leaves extract in mg/kg.

compared to controls. Donepezil treatment significantly reduced nitrite levels (P < 0.001). Pre-treatment with GS extract at 200 and 400 mg/kg also significantly decreased nitrite levels ($P < 0.001$), though the 100 mg/kg dose did not show a significant difference from the model group.

The elevated nitrite levels in the model group emphasize the role of oxidative and nitrosative stress in Sco-induced cognitive impairment. Donepezil and GS extract (200 and 400 mg/kg) effectively reduced nitrite levels, demonstrating their antioxidative properties and potential therapeutic benefits for neuroinflammation and neurodegeneration. These findings support the potential of GS extract in treating AD and related disorders.

4.4.3. Superoxide Dismutase Activity

Superoxide dismutase activity was significantly lower in the model group [F (5, 24) = 51.4; $P < 0.001$] compared to controls [\(Figure](#page-7-0) 6C). Administration of GS extract significantly increased SOD activity ($P < 0.01$, $P <$ 0.001, and $P < 0.001$ for the 100, 200, and 400 mg/kg doses, respectively), as did donepezil $(P < 0.01)$.

The decrease in SOD activity in the model group underscores the role of oxidative stress in Sco-induced cognitive impairment. Guiera senegalensis extract and donepezil effectively restored SOD activity, enhancing antioxidant defenses and reducing oxidative damage. These findings highlight the potential of GS extract in addressing oxidative stress-related neurodegenerative diseases, such as AD, by boosting SOD activity and protecting cognitive function.

4.4.4. Catalase Activity

[Figure](#page-7-0) 6D shows a significant decrease in CAT activity $[F(5, 24) = 35.7; P < 0.001]$ in the model group compared to controls. Pre-treatment with GS extract at 100, 200, and 400 mg/kg significantly increased CAT activity ($P <$ 0.01, $P < 0.001$, and $P < 0.001$, respectively). Donepezil also significantly increased CAT activity ($P < 0.01$). The decrease in CAT activity in the model group highlights the role of oxidative stress in Sco-induced cognitive impairment. Both GS extract and donepezil effectively restored CAT activity, enhancing antioxidant defenses and reducing oxidative damage. These results suggest the potential of GS extract in treating oxidative stressrelated neurodegenerative diseases, such as AD, by boosting CAT activity and protecting cognitive function.

4.4.5. Glutathione Levels

A significant decrease in GSH levels [F (5, 24) = 50.3; P < 0.001] was observed in the Sco group (model group) compared to the control group. In contrast, GSH levels in GS-treated rats (100, 200, and 400 mg/kg) were significantly higher ($P < 0.001$) compared to the model group, indicating the amelioration of Sco-induced oxidative stress [\(Figure](#page-7-0) 6E). The standard drug donepezil also significantly ($P < 0.001$) reversed the effect of Sco on GSH levels.

Figure 6. Effect of hydroethanolic GS leaves extract on hippocampus malondialdehyde (MDA) (A), nitrite (B), Superoxide Dismutase (SOD) (C), Catalase (CAT) (D) and Glutathione (GSH) (E) levels. Each point represents the mean ± SEM, (n = 5). δδδ P < 0.001 significant difference from control group; * P < 0.05, ** P < 0.01 and *** P < 0.001 significant
difference from model group; Sco: Scopolamine

4.5. Effect of Hydroethanolic Guiera Senegalensis J. F. Gmel. Leaves Extract on Proinflammatory and Anti-inflammatory Cytokines Levels

[Figure](#page-8-0) 7A shows a significant increase in hippocampal TNF- α levels [F (5, 24) = 116; P < 0.001] in the Sco-treated model group compared to controls. GS extract (100, 200, and 400 mg/kg) and donepezil significantly reduced TNF-α levels (P < 0.001).

[Figure](#page-8-0) 7B indicates higher hippocampal IL-1β levels [F $(5, 24) = 82.7$; $P < 0.001$ in the model group. The GS

Figure 7. Effect of hydroethanolic GS leaves extract on hippocampus proinflammatory cytokine (TNF-α, IL-1β, IL-6, IFN-γ) and anti-inflammatory cytokine (IL-10) levels. Each point represents the mean ± SEM, (n = 5). 888 P < 0.001 significant difference from control group; *** P < 0.001 significant difference from model group; Sco: Scopolamine 1 mg/kg, Don:
Donepezil 2 mg/kg, GS: Concentration of Gui

extract (100, 200, and 400 mg/kg) and donepezil significantly decreased IL-1β levels (P < 0.001).

[Figure](#page-8-0) 7C shows increased IL-6 levels [F $(5, 24) = 25$; P < 0.001] in Sco-treated rats. The GS extract (100, 200, and 400 mg/kg) and donepezil significantly reduced IL-6 levels (P < 0.001).

[Figure](#page-8-0) 7D reveals a significant increase in IFN-γ [F (5, 24) = 190; P < 0.001] in the Sco group. The GS extract (100, 200, and 400 mg/kg) and donepezil significantly lowered IFN-γ levels (P < 0.001).

[Figure](#page-8-0) 7E shows lower IL-10 levels [F (5, 24) = 39.1; P < 0.001] in the model group. The GS extract (200 and 400 mg/kg) and donepezil significantly increased IL-10 levels

F**igure 8.** Effect of hydroethanolic GS leaves extract on effect on hippocampus Aβ1-42 (A) and phospho Tau protein (B) levels. Results are represented as mean ± SEM, (n = 5). δδδ P
< 0.001 significant difference from contr senegalensis and Guiera senegalensis leaves extract in mg/kg.
 ≤ 0.001 significant difference from control group; $\gamma P \lt 0.05$ and Concentration of Guiera senegalensis leaves extract in mg/kg.

 $(P < 0.001)$. No significant difference was found between the 100 mg/kg GS dose and the model group.

4.6. Effect of Hydroethanolic Guiera Senegalensis J. F. Gmel. Leaves Extract on Amyloid-beta 1-42 (A*β*1-42) and Tau Phospho Protein Levels

4.6.1. Amyloid-beta 1-42 (A*β*1-42) Level Estimation

[Figure](#page-9-0) 8A shows that Aβ1-42 levels were significantly higher [F (5, 24) = 31.0; $P < 0.001$] in the model group compared to controls. Guiera senegalensis extract (100, 200, and 400 mg/kg) and donepezil significantly reduced Aβ1-42 levels ($P < 0.001$). The elevated Aβ1-42 levels in the model group underscore the role of amyloid beta peptides in Sco-induced cognitive impairment and neurodegeneration. The effectiveness of GS extract and donepezil in reducing Aβ1-42 levels suggests their potential in mitigating AD pathology, offering neuroprotective effects and aiding in the preservation of cognitive function. These results emphasize the importance of targeting amyloid beta peptides in AD treatment strategies.

4.6.2. Tau Phospho Protein Level Estimation

[Figure](#page-9-0) 8B shows significantly increased phospho Tau protein levels [F (5, 24) = 38.2; $P < 0.001$] in the model group compared to controls. Pre-treatment with GS extract (100, 200, and 400 mg/kg) significantly reduced these levels ($P < 0.05$, $p < 0.001$, and $P < 0.001$, respectively), and donepezil also significantly reduced

phospho Tau protein levels ($P < 0.001$). The elevated phospho Tau in the model group underscores the role of tau pathology in Sco-induced cognitive impairment. The GS extract and donepezil effectively reduced phospho Tau levels, suggesting their potential in mitigating tau pathology and preserving neuronal function in AD. These findings highlight the importance of targeting tau pathology for AD treatment.

4.7. Effect of Hydroethanolic Guiera Senegalensis J. F. Gmel Leaves Extract on Glial Fibrillary Acidic Protein Level

[Figure](#page-10-0) 9 shows significantly elevated GFAP levels [F (5, 24) = 56.2; P < 0.001] in the model group compared to controls. Treatment with GS extract (100, 200, and 400 mg/kg) and donepezil significantly reduced GFAP levels $(P < 0.001)$. The increased GFAP in the model group indicates neuroinflammation and neuronal damage associated with Sco-induced dementia. Both GS extract and donepezil effectively decreased GFAP levels, suggesting their potential to attenuate neuroinflammation and protect against neuronal damage. These results highlight the value of targeting neuroinflammation as a therapeutic strategy for AD and related disorders.

4.8. Effect of Hydroethanolic Guiera Senegalensis J. F. Gmel. Leaves Extract on Brain-Derived Neurotrophic Factor Level

[Figure](#page-11-0) 10 shows a significant reduction in BDNF levels [F (5, 24) = 11.5; $P < 0.001$] in the model group compared to controls. Treatment with GS extract (200

Figure 9. Effect of hydroethanolic GS leaves extract on hippocampus GFAP level. Results are represented as mean ± SEM, (n = 5). δδδ P < 0.001 significant difference from control
group; **P < 0.001 significant difference f group; ***P < 0.001 significant difference from model group; Sco: Scopolamine 1 mg/kg, Don: Donepezil 2 mg/kg, GS: Concentration of Guiera senegalensis leaves extract in mg/kg.

and 400 mg/kg) and donepezil significantly increased BDNF levels ($P < 0.001$), though the 100 mg/kg dose of GS did not show a significant difference from the model group. The decreased BDNF in the model group highlights impaired neurotrophic support and synaptic plasticity in Sco-induced dementia. The GS extract and donepezil effectively increased BDNF levels, suggesting their potential to enhance neurotrophic signaling, promote neuronal survival, and improve cognitive function. These findings underscore the importance of boosting neurotrophic support as a therapeutic approach for AD and similar neurodegenerative conditions.

4.9. Effect of Hydroethanolic Guiera Senegalensis J. F. Gmel. Leaves Extract on Histological Sections

[Figure](#page-12-0) 11 shows that the control group exhibited normal hippocampal microarchitecture in the CA1, CA2, CA3 regions, and the dentate gyrus (DG). Scopolamine treatment for 7 days caused a marked decrease in cell density in these areas. Pre-treatment with GS extract and

donepezil reversed these changes, indicating their potential neuroprotective effects. These findings suggest that both GS extract and donepezil can mitigate neuronal damage and preserve hippocampal integrity, offering promise for therapeutic interventions in AD and similar neurodegenerative disorders.

5. Discussion

The Sco-induced dementia model is commonly used to study cognitive impairments and therapies for AD ([9](#page-13-6), [14,](#page-13-11) [15\)](#page-13-15). This study evaluated the effects of the hydroethanolic extract of GS leaves on cholinergic dysfunction, oxidative stress, neuroinflammation, neurodegeneration, and memory impairment in Wistar rats with Sco-induced dementia.

Behavioral tests (Y-maze, NOR, MWM) demonstrated that GS extract improved short-term, long-term, and spatial memory. Specifically, GS extract and donepezil reversed Sco-induced memory deficits in the Y-maze [\(20](#page-13-14), [44](#page-14-14)), enhanced long-term memory in the NOR test [\(43](#page-14-13), [52\)](#page-14-22), and improved spatial memory in the MWM test by

Figure 10. Effect of hydroethanolic GS leaves extract on on hippocampus brain-derived neurotrophic factor (BDNF) level. Results are represented as mean ± SEM, (n = 5). δδδ P < 0.001 significant difference from control group; *** P < 0.001 significant difference from model group; Sco: Scopolamine 1 mg/kg, Don: Donepezil 2 mg/kg, GS: Con-centration of Guiera senegalensis leaves extract in mg/kg.

reducing escape latency and increasing the time spent in the target quadrant [\(45,](#page-14-15) [53\)](#page-14-23).

Biochemical analyses revealed that GS extract reversed Sco-induced decreases in ACh levels and increases in AChE, thereby enhancing cholinergic function ([3,](#page-13-1) [14,](#page-13-11) [15](#page-13-15), [54](#page-15-0)). The GS also reduced oxidative stress by lowering MDA levels and restoring SOD, CAT, and GSH levels ([5](#page-13-16), [19,](#page-13-17) [21](#page-13-12), [54\)](#page-15-0).

Inflammatory markers (TNF-α, IL-1β, IL-6, IFN-γ) were elevated by Sco, while IL-10 and BDNF levels were reduced, indicating neuroinflammation. The GS extract reversed these changes, suggesting anti-inflammatory and neuroplasticity benefits ([4](#page-13-3), [10](#page-13-7), [11,](#page-13-8) [55\)](#page-15-1). Additionally, GS reduced Aβ1-42 and phospho Tau levels, indicating potential therapeutic benefits by inhibiting amyloid plaque formation and Tau hyperphosphorylation [\(56-](#page-15-2) [58\)](#page-15-3).

The antioxidant and anti-inflammatory effects of GS are likely attributed to its bioactive compounds, such as flavonoids and polyphenolic acids ([19,](#page-13-17) [46,](#page-14-16) [59](#page-15-4), [60\)](#page-15-5). Compounds like quercetin, apigenin, and catechin have shown promise in improving memory and cognitive function (9) .

Histological analysis confirmed that GS extract reversed the Sco-induced decrease in cell density in the hippocampus, aligning with the behavioral and biochemical findings. Overall, hydroethanolic GS extract effectively mitigates Sco-induced cognitive impairments, highlighting its therapeutic potential for AD and similar conditions.

5.1. Conclusions

Our results suggest that the hydroethanolic extract of GS holds promise as a potential treatment for AD. Guiera senegalensis exhibited significant anticholinesterase, antioxidant, and anti-inflammatory effects, improving cognitive function while reducing oxidative stress and neuroinflammation in a Scoinduced rat model. The reduction in anticholinergic activity coincided with decreased levels of pro-

Figure 11. Effect of hydroethanolic GS leaves extract on scopolamine (Sco)-induced hippocampus histological changes. Coronal sections - hematoxylin and eosin - magnification
250X. CAI = Cornu ammonis 1, CA2 = Cornu ammon

inflammatory cytokines and oxidative stress markers, indicating potential interactions between these pathways. These findings highlight GS's multifaceted neuroprotective effects and its potential as a therapeutic agent for AD, warranting further exploration of its mechanisms and clinical applications.

Footnotes

Conflict of Interests Statement: The authors declare no conflict of interest.

Data Availability: No new data were created or analyzed in this study. Data sharing does not apply to article.

Ethical Approval: This study was approved by the Ethic committee of the Faculty of Science of the University of Maroua (Ref. N"14/02161 UMa/D/FS/VD-RC (https://tabletadefrumusete.ro/ethical-declarations-ofprofessor-simplice-harqlin-foyet/)

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References

- 1. G. B. D. Dementia Forecasting Collaborators. Estimation of the global prevalence of dementia in 2019 and forecasted prevalence in 2050: an analysis for the Global Burden of Disease Study 2019. Lancet Public Health. 2022;7(2):e105-25. [PubMed ID: [34998485](http://www.ncbi.nlm.nih.gov/pubmed/34998485)]. [PubMed Central ID: [PMC8810394](https://www.ncbi.nlm.nih.gov/pmc/PMC8810394)]. [https://doi.org/10.1016/S2468-2667\(21\)00249-8](https://doi.org/10.1016/S2468-2667(21)00249-8).
- 2. Foyet HS, Keugong Wado E, Ngatanko Abaissou HH, Assongalem EA, Eyong OK. Anticholinesterase and Antioxidant Potential of Hydromethanolic Extract of Ziziphus mucronata (Rhamnaceae) Leaves on Scopolamine-Induced Memory and Cognitive Dysfunctions in Mice. Evid Based Complement Alternat Med. 2019;2019:4568401. [PubMed ID: [31781268\]](http://www.ncbi.nlm.nih.gov/pubmed/31781268). [PubMed Central ID: [PMC6855091\]](https://www.ncbi.nlm.nih.gov/pmc/PMC6855091). <https://doi.org/10.1155/2019/4568401>.
- 3. Nazir N, Zahoor M, Nisar M, Karim N, Latif A, Ahmad S, et al. Evaluation of neuroprotective and anti-amnesic effects of Elaeagnus umbellata Thunb. On scopolamine-induced memory impairment in mice. BMC Complement Med Ther. 2020;20(1):143. [PubMed ID: [32397979\]](http://www.ncbi.nlm.nih.gov/pubmed/32397979). [PubMed Central ID: [PMC7216467](https://www.ncbi.nlm.nih.gov/pmc/PMC7216467)]. <https://doi.org/10.1186/s12906-020-02942-3>.
- 4. Muhammad T, Ali T, Ikram M, Khan A, Alam SI, Kim M. Melatonin Rescue Oxidative Stress-Mediated Neuroinflammation/ Neurodegeneration and Memory Impairment in Scopolamine-Induced Amnesia Mice Model. J Neuroimmune Pharmacol. 2019;14(2):278-94. <https://doi.org/10.1007/s11481-018-9824-3>.
- 5. Al Omairi NE, Al-Brakati AY, Kassab RB, Lokman MS, Elmahallawy EK, Amin HK, et al. Soursop fruit extract mitigates scopolamine-induced amnesia and oxidative stress via activating cholinergic and Nrf2/HO-1 pathways. Metab Brain Dis. 2019;34(3):853-64. [PubMed ID: [30919246](http://www.ncbi.nlm.nih.gov/pubmed/30919246)]. [https://doi.org/10.1007/s11011-019-00407-2.](https://doi.org/10.1007/s11011-019-00407-2)
- 6. Tao G, Min-Hua C, Feng-Chan X, Yan C, Ting S, Wei-Qin L, et al. Changes of plasma acetylcholine and inflammatory markers in critically ill patients during early enteral nutrition: A prospective observational study. J Crit Care. 2019;52:219-26. [PubMed ID: [31108325](http://www.ncbi.nlm.nih.gov/pubmed/31108325)]. [https://doi.org/10.1016/j.jcrc.2019.05.008.](https://doi.org/10.1016/j.jcrc.2019.05.008)
- 7. Xian YF, Ip SP, Mao QQ, Su ZR, Chen JN, Lai XP, et al. Honokiol improves learning and memory impairments induced by scopolamine in mice. Eur J Pharmacol. 2015;760:88-95. [PubMed ID: [25912802\]](http://www.ncbi.nlm.nih.gov/pubmed/25912802). <https://doi.org/10.1016/j.ejphar.2015.04.013>.
- 8. Roy ER, Wang B, Wan YW, Chiu G, Cole A, Yin Z, et al. Type I interferon response drives neuroinflammation and synapse loss in Alzheimer disease. J Clin Invest. 2020;130(4):1912-30. [PubMed Central ID: [PMC7108898](https://www.ncbi.nlm.nih.gov/pmc/PMC7108898)]. <https://doi.org/10.1172/JCI133737>.
- 9. Olayinka J, Eduviere A, Adeoluwa O, Fafure A, Adebanjo A, Ozolua R. Quercetin mitigates memory deficits in scopolamine mice model via protection against neuroinflammation and neurodegeneration. Life
Sci. 2022:292:120326. [PubMed ID: 35031260]. Sci. 2022;292:120326. [https://doi.org/10.1016/j.lfs.2022.120326.](https://doi.org/10.1016/j.lfs.2022.120326)
- Konar A, Shah N, Singh R, Saxena N, Kaul SC, Wadhwa R, et al. Protective role of Ashwagandha leaf extract and its component withanone on scopolamine-induced changes in the brain and brainderived cells. PLoS One. 2011;6(11). e27265. [PubMed ID: [22096544](http://www.ncbi.nlm.nih.gov/pubmed/22096544)]. [PubMed Central ID: [PMC3214041](https://www.ncbi.nlm.nih.gov/pmc/PMC3214041)]. <https://doi.org/10.1371/journal.pone.0027265>.
- 11. Hazzaa SM, Abdelaziz SAM, Abd Eldaim MA, Abdel-Daim MM, Elgarawany GE. Neuroprotective Potential of Allium sativum against Monosodium Glutamate-Induced Excitotoxicity: Impact on Short-Term Memory, Gliosis, and Oxidative Stress. Nutrients. 2020;12(4). [PubMed ID: [32290031](http://www.ncbi.nlm.nih.gov/pubmed/32290031)]. [PubMed Central ID: [PMC7230314](https://www.ncbi.nlm.nih.gov/pmc/PMC7230314)]. [https://doi.org/10.3390/nu12041028.](https://doi.org/10.3390/nu12041028)
- 12. Zhang J, Wang J, Zhou GS, Tan YJ, Tao HJ, Chen JQ, et al. Studies of the Anti-amnesic Effects and Mechanisms of Single and Combined Use of Donepezil and Ginkgo Ketoester Tablet on Scopolamine-Induced Memory Impairment in Mice. Oxid Med Cell Longev. 2019;2019:8636835. [PubMed ID: [30911351\]](http://www.ncbi.nlm.nih.gov/pubmed/30911351). [PubMed Central ID: [PMC6398023](https://www.ncbi.nlm.nih.gov/pmc/PMC6398023)]. <https://doi.org/10.1155/2019/8636835>.
- 13. Woo Y, Lim JS, Oh J, Lee JS, Kim JS. Neuroprotective Effects of Euonymus alatus Extract on Scopolamine-Induced Memory Deficits in Mice. Antioxidants (Basel). 2020;9(5). [PubMed ID: [32456069\]](http://www.ncbi.nlm.nih.gov/pubmed/32456069).
[PubMed Central ID: PMC7278771]. Central ID: [PMC7278771\]](https://www.ncbi.nlm.nih.gov/pmc/PMC7278771). <https://doi.org/10.3390/antiox9050449>.
- 14. Choi JH, Lee EB, Jang HH, Cha YS, Park YS, Lee SH. Allium hookeri Extracts Improve Scopolamine-Induced Cognitive Impairment via Activation of the Cholinergic System and Anti-Neuroinflammation in Mice. Nutrients. 2021;13(8). [PubMed ID: [34445062](http://www.ncbi.nlm.nih.gov/pubmed/34445062)]. [PubMed Central ID: [PMC8400157\]](https://www.ncbi.nlm.nih.gov/pmc/PMC8400157). <https://doi.org/10.3390/nu13082890>.
- 15. Ionita R, Postu PA, Beppe GJ, Mihasan M, Petre BA, Hancianu M, et al. Cognitive-enhancing and antioxidant activities of the aqueous extract from Markhamia tomentosa (Benth.) K. Schum. stem bark in a rat model of scopolamine. Behav Brain Funct. 2017;13(1):5. [PubMed ID: 28351401]. [PubMed Central ID: PMC5371259]. ID: [28351401](http://www.ncbi.nlm.nih.gov/pubmed/28351401)]. [PubMed Central ID: [PMC5371259\]](https://www.ncbi.nlm.nih.gov/pmc/PMC5371259). [https://doi.org/10.1186/s12993-017-0123-6.](https://doi.org/10.1186/s12993-017-0123-6)
- 16. Navarro NM, Krawczyk MC, Boccia MM, Blake MG. Extinction and recovery of an avoidance memory impaired by scopolamine. *Physiol*
Behav. 2017;171:192-8. [PubMed ID: 28069463]. 2017;171:192-8. [https://doi.org/10.1016/j.physbeh.2016.12.042.](https://doi.org/10.1016/j.physbeh.2016.12.042)
- 17. Guo C, Shen J, Meng Z, Yang X, Li F. Neuroprotective effects of polygalacic acid on scopolamine-induced memory deficits in mice.
Phytomedicine. 2016;23(2):149-55. [PubMed ID: 26926176]. Phytomedicine. 2016;23(2):149-55. [PubMed [https://doi.org/10.1016/j.phymed.2015.12.009.](https://doi.org/10.1016/j.phymed.2015.12.009)
- 18. Haider S, Tabassum S, Perveen T. Scopolamine-induced greater alterations in neurochemical profile and increased oxidative stress demonstrated a better model of dementia: A comparative study. Brain Res Bull. 2016;127:234-47. [PubMed ID: [27725168\]](http://www.ncbi.nlm.nih.gov/pubmed/27725168). <https://doi.org/10.1016/j.brainresbull.2016.10.002>.
- 19. Parfait B, Galba Jean B, Roger P, Herve Herve NA, Balbine KK, Guillaume CW, et al. Antioxidant and Anticholinesterase Properties of the Aqueous Extract of Balanites aegyptiaca L. Delile Fruit Pulp on Monosodium Glutamate-Induced Excitotoxicity in Swiss Mice. Evid Based Complement Alternat Med. 2022;2022:7576132. [PubMed ID: [35449814](http://www.ncbi.nlm.nih.gov/pubmed/35449814)]. [PubMed Central ID: [PMC9017515\]](https://www.ncbi.nlm.nih.gov/pmc/PMC9017515). <https://doi.org/10.1155/2022/7576132>.
- 20. Subedi L, Cho K, Park YU, Choi HJ, Kim SY. Sulforaphane-Enriched Broccoli Sprouts Pretreated by Pulsed Electric Fields Reduces Neuroinflammation and Ameliorates Scopolamine-Induced Amnesia in Mouse Brain through Its Antioxidant Ability via Nrf2-HO-1 Activation. Oxid Med Cell Longev. 2019;2019:3549274. [PubMed ID: 31049133]. [PubMed Central ID: PMC6458888]. PMC64588881. <https://doi.org/10.1155/2019/3549274>.
- 21. Jo SH, Kang TB, Koppula S, Cho DY, Kim JS, Kim IS, et al. Mitigating Effect of Lindera obtusiloba Blume Extract on Neuroinflammation in Microglial Cells and Scopolamine-Induced Amnesia in Mice. Mol. 2021;26(10). [PubMed ID: [34066108](http://www.ncbi.nlm.nih.gov/pubmed/34066108)]. [PubMed Central ID: [PMC8151320](https://www.ncbi.nlm.nih.gov/pmc/PMC8151320)]. [https://doi.org/10.3390/molecules26102870.](https://doi.org/10.3390/molecules26102870)
- 22. Bihaqi SW, Singh AP, Tiwari M. Supplementation of Convolvulus pluricaulis attenuates scopolamine-induced increased tau and amyloid precursor protein (AbetaPP) expression in rat brain. Indian J Pharmacol. 2012;44(5):593-8. [PubMed ID: [23112420](http://www.ncbi.nlm.nih.gov/pubmed/23112420)]. [PubMed Central ID: [PMC3480791](https://www.ncbi.nlm.nih.gov/pmc/PMC3480791)]. <https://doi.org/10.4103/0253-7613.100383>.
- 23. Baek MS, Cho H, Lee HS, Choi JY, Lee JH, Ryu YH, et al. Temporal trajectories of in vivo tau and amyloid-beta accumulation in Alzheimer's disease. Eur J Nucl Med Mol Imaging. 2020;47(12):2879-86. [PubMed ID: [32350558\]](http://www.ncbi.nlm.nih.gov/pubmed/32350558). <https://doi.org/10.1007/s00259-020-04773-3>.
- 24. Balaban H, Naziroglu M, Demirci K, Ovey IS. The Protective Role of Selenium on Scopolamine-Induced Memory Impairment, Oxidative Stress, and Apoptosis in Aged Rats: The Involvement of TRPM2 and TRPV1 Channels. Mol Neurobiol. 2017;54(4):2852-68. [PubMed ID: [27021021\]](http://www.ncbi.nlm.nih.gov/pubmed/27021021). [https://doi.org/10.1007/s12035-016-9835-0.](https://doi.org/10.1007/s12035-016-9835-0)
- 25. Mendiola-Precoma J, Berumen LC, Padilla K, Garcia-Alcocer G. Therapies for Prevention and Treatment of Alzheimer's Disease. Biomed Res Int. 2016;2016:2589276. [PubMed ID: [27547756\]](http://www.ncbi.nlm.nih.gov/pubmed/27547756). [PubMed Central ID: [PMC4980501\]](https://www.ncbi.nlm.nih.gov/pmc/PMC4980501). <https://doi.org/10.1155/2016/2589276>.
- 26. Akram M, Nawaz A. Effects of medicinal plants on Alzheimer's disease and memory deficits. Neural Regen Res. 2017;12(4):660-70. [PubMed ID: [28553349\]](http://www.ncbi.nlm.nih.gov/pubmed/28553349). [PubMed Central ID: [PMC5436367](https://www.ncbi.nlm.nih.gov/pmc/PMC5436367)]. [https://doi.org/10.4103/1673-5374.205108.](https://doi.org/10.4103/1673-5374.205108)
- 27. Keugong Wado E, Kubicki M, Ngatanko AHH, Léa Blondelle KD, Jorelle Linda D, Roland RN, et al. Anxiolytic and antidepressant effects of Ziziphus mucronata hydromethanolic extract in male rats exposed to unpredictable chronic mild stress: Possible mechanisms of actions. J Ethnopharmacol. 2020;260:112987. [PubMed ID: [32446929](http://www.ncbi.nlm.nih.gov/pubmed/32446929)]. [https://doi.org/10.1016/j.jep.2020.112987.](https://doi.org/10.1016/j.jep.2020.112987)
- 28. Abubakr M, Sirag N, Osman I, Osman M, Abakar S, Aboul-Enein AM. Anticancer and antioxidant activities of Guiera senegalensis. Sudan J Med Sci. 2013;8(3):135-40.
- 29. Damo JLK, Boiangiu RS, Brinza I, Kenko Djoumessi LB, Rebe RN, Kamleu BN, et al. Neuroprotective Potential of Guiera senegalensis (Combretaceae) Leaf Hydroethanolic Extract against Cholinergic System Dysfunctions and Oxidative Stress in Scopolamine-Induced Cognitive Impairment in Zebrafish (Danio rerio). Plants (Basel). 2022;11(9). [PubMed ID: [35567150\]](http://www.ncbi.nlm.nih.gov/pubmed/35567150). [PubMed Central ID: [PMC9100236](https://www.ncbi.nlm.nih.gov/pmc/PMC9100236)]. [https://doi.org/10.3390/plants11091149.](https://doi.org/10.3390/plants11091149)
- 30. Kinda PT, Zerbo P, Guenne S, Compaore M, Ciobica A, Kiendrebeogo M. Medicinal Plants Used for Neuropsychiatric Disorders Treatment in the Hauts Bassins Region of Burkina Faso. Med (Basel). 2017;4(2). [PubMed ID: [28930246\]](http://www.ncbi.nlm.nih.gov/pubmed/28930246). [PubMed Central ID: [PMC5590068](https://www.ncbi.nlm.nih.gov/pmc/PMC5590068)]. <https://doi.org/10.3390/medicines4020032>.
- 31. Somboro AA, Patel K, Diallo D, Sidibe L, Chalchat JC, Figueredo G, et al. An ethnobotanical and phytochemical study of the African medicinal plant Guiera senegalensis JF Gmel. J Med Plants Res. 2011;5(9):1639-51.
- 32. Hamadnalla HMY, Hamad MAB, Adam AAI. Phytochemical Investigation, Antimicrobial, Antioxidant and Anti-Diabetic Potential of Guiera Senegalensis Leaves Extracts. Cytokines Relat. Miner Dust Induc Dis. 2020;4:1-4.
- 33. Lamien CE, Meda A, Couacy-Hymann E, Ouedraogo AG, Nacoulma OG. The phytochemical composition and in vitro antiviral activity of decoctions from galls of Guiera senegalensis J.F. Gmel. (Combretaceae) and their relative non-toxicity for chickens. Onderstepoort J Vet Res. 2005;72(2):111-8. [PubMed ID: [16137128](http://www.ncbi.nlm.nih.gov/pubmed/16137128)].
- 34. Miaffo D, Zingué S, Dingamtoudji M, Kamanyi A. Preventive effects of the aqueous extract of Guiera senegalensis roots on Dexamethasone induced insulin resistance in Mice. RJPBS. 2019;10:8-18.
- 35. Sombié PAED, Hilou A, Mounier C, Coulibaly AY, Kiendrebeogo MJFM, Millogo JF, et al. Antioxidant and anti-inflammatory activities from galls of Guiera senegalensis JF Gmel (Combretaceae). Res J Med Plant. 2011;5:448-61.
- 36. Somboro AA, Diallo D, Sidibe L, Traore N, Fofana B, Bouare S, et al. Activités anticholinestérasiques des alcaloïdes totaux extraits des feuilles, fruits, écorces de racines et écorces de tronc de Guiera senegalensis, une plante médicinale Malienne. Int J Biologica Chem Sci. 2013;7(4):1723-8.
- 37. Koehler D, Williams FE. Utilizing zebrafish and okadaic acid to study Alzheimer's disease. Neural Regen Res. 2018;13(9):1538-41. [PubMed ID:
30127109]. [PubMed Central ID: PMC6126137]. [PMC6126137](https://www.ncbi.nlm.nih.gov/pmc/PMC6126137)]. <https://doi.org/10.4103/1673-5374.237111>.
- 38. Imam N, Abdelwahab SI, Taha MM, Mohan S, Alhazmi H, Ahmed RH. Antioxidant and hepatoprotective activities of Blepharis linariifolia pers and Guiera senegalensis JF Gmel. against CCl4-induced hepatotoxicity. World J Pharm Res. 2018;7:70-80.
- 39. Suresh R, Saravanakumar M, Suganyadevi P. Anthocyanins from Indian cassava (Manihot esculenta Crantz) and its antioxidant properties. Int J Pharmaceutical Sci Res. 2011;2(7):1819.
- 40. Kansci G, Koubala BB, Lape IM. Effect of ripening on the composition and the suitability for jam processing of different varieties of mango (Mangifera indica). African J Biotechnol. 2003;2(9):296-300.
- 41. Hasanein P, Mahtaj AK. Ameliorative effect of rosmarinic acid on scopolamine-induced memory impairment in rats. Neurosci Lett.
2015:585:23-7. [PubMed ID: 25445372]. 2015**:585:23-7.** <https://doi.org/10.1016/j.neulet.2014.11.027>.
- 42. Lee S, Park HJ, Jeon SJ, Kim E, Lee HE, Kim H, et al. Cognitive Ameliorating Effect of Acanthopanax koreanum Against Scopolamine-Induced Memory Impairment in Mice. Phytother Res.
2017;31(3):425-32. [PubMed ID: 28164395]. 2017;31(3):425-32. [PubMed ID: [28164395\]](http://www.ncbi.nlm.nih.gov/pubmed/28164395). <https://doi.org/10.1002/ptr.5764>.
- 43. Sohn E, Lim HS, Kim YJ, Kim BY, Jeong SJ. Annona atemoya Leaf Extract Improves Scopolamine-Induced Memory Impairment by Preventing Hippocampal Cholinergic Dysfunction and Neuronal Cell Death. Int J Mol Sci. 2019;20(14). [PubMed ID: [31331043](http://www.ncbi.nlm.nih.gov/pubmed/31331043)]. [PubMed Central ID: [PMC6679418](https://www.ncbi.nlm.nih.gov/pmc/PMC6679418)]. [https://doi.org/10.3390/ijms20143538.](https://doi.org/10.3390/ijms20143538)
- 44. El-Marasy SA, El-Shenawy SM, El-Khatib AS, El-Shabrawy OA, Kenawy SA. Effect of Nigella sativa and wheat germ oils on scopolamineinduced memory impairment in rats. Bulletin Faculty Pharmacy, Cairo Univ. 2012;50(2):81-8.
- 45. Lea Blondelle KD, Simplice FH, Herve Herve NA, Eglantine KW, Roland RN, Jorelle Linda DK, et al. Antidepressant, anti-amnesic and vasoprotective effect of Bombax costatum Pellegr. & Vuillet aqueous stem bark extract on chronic mild unpredictable stress induced in rat. J Ethnopharmacol. 2022;293:115315. [PubMed ID: [35487448\]](http://www.ncbi.nlm.nih.gov/pubmed/35487448). [https://doi.org/10.1016/j.jep.2022.115315.](https://doi.org/10.1016/j.jep.2022.115315)
- 46. Misra HP, Fridovich I. The generation of superoxide radical during the autoxidation of hemoglobin. *J Biol Chem.* 1972;247(21):6960-2. [PubMed ID: [4673289\]](http://www.ncbi.nlm.nih.gov/pubmed/4673289).
- 47. Sinha AK. Colorimetric assay of catalase. Anal Biochem. 1972;47(2):389-94. [PubMed ID: [4556490\]](http://www.ncbi.nlm.nih.gov/pubmed/4556490). [https://doi.org/10.1016/0003-](https://doi.org/10.1016/0003-2697(72)90132-7) [2697\(72\)90132-7.](https://doi.org/10.1016/0003-2697(72)90132-7)
- 48. Wilbur KM, Bernheim F, Shapiro OW. Determination of lipid peroxidation. Arch Biochem Biophysics. 1949;24:305-10.
- 49. Fermor B, Weinberg JB, Pisetsky DS, Misukonis MA, Banes AJ, Guilak F. The effects of static and intermittent compression on nitric oxide production in articular cartilage explants. J Orthop Res. 2001;19(4):729-37. [PubMed ID: [11518285](http://www.ncbi.nlm.nih.gov/pubmed/11518285)]. [https://doi.org/10.1016/S0736-](https://doi.org/10.1016/S0736-0266(00)00049-8) [0266\(00\)00049-8.](https://doi.org/10.1016/S0736-0266(00)00049-8)
- 50. Smith A, Bruton J. Color Atlas of Histological Staining Techniques. Maryland Heights, Missouri: Year Book Medical Publishers; 1977.
- 51. Shin K, Guo H, Cha Y, Ban YH, Seo da W, Choi Y, et al. Cereboost™, an American ginseng extract, improves cognitive function via upregulation of choline acetyltransferase expression and neuroprotection. Regul Toxicol Pharmacol. 2016;78:53-8. [PubMed ID: [27112419\]](http://www.ncbi.nlm.nih.gov/pubmed/27112419). <https://doi.org/10.1016/j.yrtph.2016.04.006>.
- 52. Abaissou HHN, Foyet HS, Wado EK, Nkwingwa BK, Kamda JLD, Djomessie LBK, et al. Cholinergic‑Enhancing and Antioxidant Effect of Vigna subterranea (L.) Verdc.(Fabaceae) Landrace Aqueous Extract on Scopolamine‑Induced Amnesia in Male Swiss Mice. Pharmacognosy Res. 2020;12(3).
- 53. Liu S, Li Y, Yi F, Liu Q, Chen N, He X, et al. Resveratrol oligomers from Paeonia suffruticosa protect mice against cognitive dysfunction by regulating cholinergic, antioxidant and anti-inflammatory
- 54. Wang X, Zhang D, Song W, Cai CF, Zhou Z, Fu Q, et al. Neuroprotective effects of the aerial parts of Polygala tenuifolia Willd extract on scopolamine-induced learning and memory impairments in mice. Biomed Rep. 2020;13(5):37. [PubMed ID: [32874571\]](http://www.ncbi.nlm.nih.gov/pubmed/32874571). [PubMed Central ID: [PMC7453304\]](https://www.ncbi.nlm.nih.gov/pmc/PMC7453304). [https://doi.org/10.3892/br.2020.1344.](https://doi.org/10.3892/br.2020.1344)
- 55. Butterfield DA, Reed T, Newman SF, Sultana R. Roles of amyloid betapeptide-associated oxidative stress and brain protein modifications in the pathogenesis of Alzheimer's disease and mild cognitive impairment. Free Radic Biol Med. 2007;43(5):658-77. [PubMed ID:
17664130]. [PubMed Central ID: PMC2031860]. [17664130\]](http://www.ncbi.nlm.nih.gov/pubmed/17664130). [PubMed Central ID: [PMC2031860](https://www.ncbi.nlm.nih.gov/pmc/PMC2031860)]. <https://doi.org/10.1016/j.freeradbiomed.2007.05.037>.
- 56. Williams TL, Serpell LC. Membrane and surface interactions of Alzheimer's Abeta peptide-insights into the mechanism of cytotoxicity. FEBS J. 2011;278(20):3905-17. [PubMed ID: [21722314](http://www.ncbi.nlm.nih.gov/pubmed/21722314)]. <https://doi.org/10.1111/j.1742-4658.2011.08228.x>.
- 57. Bachurin SO, Bovina EV, Ustyugov AA. Drugs in Clinical Trials for Alzheimer's Disease: The Major Trends. Med Res Rev. 2017;37(5):1186- 225. [PubMed ID: [28084618](http://www.ncbi.nlm.nih.gov/pubmed/28084618)]. <https://doi.org/10.1002/med.21434>.
- 58. Singsai K, Ladpala N, Dangja N, Boonchuen T, Jaikhamfu N, Fakthong P. Effect of Streblus asper Leaf Extract on Scopolamine-Induced Memory Deficits in Zebrafish: The Model of Alzheimer's Disease. Adv Pharmacol Pharm Sci. 2021;2021:6666726. [PubMed ID: [33987539\]](http://www.ncbi.nlm.nih.gov/pubmed/33987539). [PMC8093034\]](https://www.ncbi.nlm.nih.gov/pmc/PMC8093034). [https://doi.org/10.1155/2021/6666726.](https://doi.org/10.1155/2021/6666726)
- 59. Kumar S, Pandey AK. Chemistry and biological activities of flavonoids: an overview. Sci World J. 2013;2013:162750. [PubMed ID: 24470791]. [PubMed Central ID: PMC3891543]. [PMC3891543\]](https://www.ncbi.nlm.nih.gov/pmc/PMC3891543). [https://doi.org/10.1155/2013/162750.](https://doi.org/10.1155/2013/162750)
- 60. Panche AN, Diwan AD, Chandra SR. Flavonoids: an overview. J Nutr Sci. 2016;5. e47. [PubMed ID: [28620474](http://www.ncbi.nlm.nih.gov/pubmed/28620474)]. [PubMed Central ID: [PMC5465813\]](https://www.ncbi.nlm.nih.gov/pmc/PMC5465813). [https://doi.org/10.1017/jns.2016.41.](https://doi.org/10.1017/jns.2016.41)