Published online 2016 August 30.

Research Article

Healing Effect of Hawthorn (*Crataegus pontica* C. Koch) Leaf Extract in Dermal Toxicity Induced by T-2 Toxin in Rabbit

Heibatullah Kalantari,¹ Ali Asghar Hemmati,¹ Mehdi Goudarzi,¹ Hossein Forouzandeh,^{1,2,*} Mojtaba

Kalantar,¹ Nasrin Aghel,³ Moslem Kiyani Aslani,¹ and Tahere Shamsi Ehsan¹

¹Department of Pharmacology and Toxicology, Pharmacy School, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, IR Iran ²Iranian Blood Transfusion Organization (IBTO), Shiraz, IR Iran

³Department of Pharmacogenosy, Pharmacy School, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, IR Iran

Corresponding author: Hossein Forouzandeh, Department of Pharmacology and Toxicology, Pharmacy School, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, IR Iran. Tel: +98-9379873029, Fax: +98-7152247777, E-mail: hosainforuozandeh@yahoo.com

Received 2015 December 21; Revised 2016 May 12; Accepted 2016 May 30.

Abstract

Background: T-2 toxin is a mycotoxin, exposure to which causes dermal and systemic toxicity. *Crataegus pontica* (a member of the Rosaceous family) is a small tree with caduceus foliage that is widely distributed throughout the western and central areas of Iran. **Objectives:** This study was performed to examine the healing effects of creams prepared using *Crataegus pontica* leaf extract on dermal toxicity induced by T-2 toxin.

Methods: Iranian rabbits were used for this study. The back left flanks of the animals were shaved and to induce toxicity, a solution of 100 μ g/12 μ l of T-2 toxin in ethanol was applied to the skin for 2 successive days. Creams were prepared using *Crataegus pontica* leaf extract in a eucerin base at concentrations of 5%, 10%, and 15% (w/w). Beginning on the third day, the creams were applied to the skin lesions twice per day until complete healing occurred. The positive control group received the toxin only without treatment and the negative control group received solvent (ethanol) only. Healing was defined as a decreased wound margin, as well as treatment of the erythema and blisters.

Results: Our findings indicated that wound healing occurred 15, 15, 12, 10, and 10 days after initial wounding in the negative control, eucerin, 5%, 10%, and 15% *C. pontica* extract groups, respectively. The most effective treatments were obtained with the creams containing 10% and 15% *C. pontica* extract. Histological findings confirmed wound reduction in the affected areas.

Conclusions: The results obtained in this study indicate that *C. pontica* extract has a healing effect on dermal toxicity caused by T-2 toxin and is effective for its treatment.

Keywords: Crataegus Pontica, Dermal Toxicity, T-2 Toxin, Rabbit

1. Background

Tricothecene is classified as a mycotoxin, which is produce by various species of fungi including Fusarium, Trichoderma, Myrothecium, Verticimonisporium, and Stachynotris (1). T-2 toxin, which belongs to the tricothecene group, is mainly produce by *Fusarium sporotrichoedes* (2), and has the highest toxicity among the toxins produced by Fusarium. In the case of oral administration in rats, T-2 toxin is mainly absorbed in the intestine. T-2 toxin causes necrotic lesions and bleeding in the stomach, intestine, liver, and kidney (3). T-2 toxin absorbs through the skin and causes local inflammation that is potentially lethal. Macroscopic evaluation indicates that in humans, severe dermal irritation caused by small amounts of T-2 toxin can potentially last for two weeks. A variety of mechanisms of action have been proposed for T-2 toxin, which

reacts with the thiol groups of sulfhydryl enzymes and inhibits protein and DNA synthesis (4), as well as impairs the production of antibodies, alters membrane functions, reduces lymphocyte proliferation, and alters the maturation process of dendritic cells (5-7). Currently, the demand for herbal therapy is increasing and the amount of research being conducted on traditional herbal drugs indicates that many people around the world trust such remedies to alleviate various skin diseases (8). The genus Crataegus belongs to the family Rosaceous comprising a complex group of trees and shrubs that are native to Northern temperate zones, mostly between latitudes 30° and 50° N. Hawthorn refers to the plant Crataegus and is widely distributed throughout the Northern temperate regions of the world having approximately 280 species. Crataegus pontica is a small tree or large spiny shrub, mostly growing 6 - 10 m tall, with a dense crown. The bark is

Copyright © 2016, School of Pharmacy, Ahvaz Jundishapur University of Medical Sciences. This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (http://creativecommons.org/licenses/by-nc/4.0/) which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited.

dull brown with vertical orange cracks. Leaves grow in clusters on spur shoots and most species have lobed or serrated margins. They are somewhat variable in shape, with the lobes spreading at a wide angle. Blossoms are white; petals are oval or approximately round. The fruit is berry-like in appearance, having a pome-like structure, with a golden yellow skin color. Crataegus pontica C. Koch (Persian name: zalzalak) is a member of this genus and is widely distributed throughout the western, northwestern, and central areas of Iran (9). Almost all parts of these plants are reported to possess various pharmacological properties including anti-inflammatory, gastro-protective, antimicrobial, cardio tonic, hypotensive, and hepatoprotective effects. In Iranian traditional medicine, the leaves and flowers of some Crataegus species have been used to produce decoctions for various purposes such as tonics for cardiovascular diseases, sedatives, and antianxiety agents. Hawthorn fruit has been reported to possess cardio tonic, coronary vasodilator, and hypotensive properties. Traditionally, it has been used for cardiac failure, myocardial weakness, paroxysmal tachycardia, hypertension, arteriosclerosis, and Buerger's disease. Hawthorn leaves, flowers, and berries contain a variety of bioflavonoid-like complexes that appear to be primarily responsible for the plant's actions on the cardiac system (9, 10).

2. Objectives

The experiments described in the present study were performed to examine the healing effects of creams prepared using *Crataegus pontica* leaf extract on dermal toxicity induced by T-2 toxin.

3. Methods

3.1. Animals

Male and female Iranian rabbits (1.8 - 2.2 kg) were obtained from the animal house at Ahvaz Jundishapur University of Medical Sciences. Animals were housed individually in cages and allowed to feed on a standard commercial pellet diet supplemented with fresh vegetables, carrots, and tap water ad libitum. The animals were maintained under controlled conditions at 20 ± 2 °C with a 12 hours light: 12 hours dark cycle. The investigation was performed according to the Local Animal Ethics Committee guidelines for the use of experimental animals.

3.2. Extract Preparation

Fresh *C. pontica* leaves were collected from the Khuzestan region of Iran in September 2013. The leaves were identified at the herbarium in the department of pharmacognosy, school of pharmacy, Ahvaz, Iran. The leaves were dried in shade at room temperature, crushed into small pieces, and soaked in a 70% aqueous-ethanol solution in a large container for 3 days with occasional shaking (11). The extract was filtered through a clean cotton cloth, dried using a rotary evaporator at 40°C, and then freeze-dried. The extract yield (dry powder) was 16% w/w. The powdered extract and eucerin were then mixed at proportions of 5 to 95 m/m, 10 to 90 m/m and 15 to 85 m/m, to respectively produce *C. pontica* creams at concentrations of 5%, 10%, and 15%.

3.3. Experimental Schedule

T-2 toxin was supplied in a 5 mg vial, which was solubilized using ethanol (as a solvent) to a final volume of 0.6 ml before use. The hair on the animals' lower backs around the backbone was shaved so that the areas were fully clear. A 1.5 \times 1.5 cm square was drawn on the desired area using a marker. The toxin solution was topically administered to the marked region using a Hamilton micro syringe. To induce dermal toxicity, a solution of 100 μ g/12 μ l T-2 toxin was applied to the marked skin surface area for 2 successive days. The experiment was performed on six groups, each containing 5 rabbits. The groups were treated as follows: group1(positive control) was treated with toxin only; group 2 (negative control) was treated with eucerin only; group 3 was treated with 5% C. pontica cream in a eucerin base; group 4 was treated with 10% C. pontica cream; group 5 was treated with 15% C. pontica cream; and group 6 was treated with ethanol only (to evaluate the safety of the T-2 toxin solvent). All treatments commenced 48 hours after toxin administration; treatment was applied topically to the surface of the damaged areas twice per day for 7 consecutive days. During treatment of the various groups, damages due to toxicity, including blistering, inflammation, and erythematic (redness) reactions, were evaluated by means of a score ranging from +1 to +4, according to severity.

3.4. Statistical Analysis

Statistical evaluation was performed, using the Kruskal-Wallis test. P-values less than 0.05 were considered significant.

4. Results

4.1. Macroscopic Inspection

The skins of the animals in the different groups were inspected daily and the degree of healing of the lesions was recorded. In the group that was treated with ethanol, no abnormalities were observed on the initial day; therefore histological studies were not performed in this group. No animal mortality occurred during the study. In the first 48 hours after toxin application, edema, stiffness, and erythema were observed in the epidermis; furthermore, seemingly deep blisters formed in some areas (Figure 1A). On the 9th day after T-2 toxin administration, central necrosis surrounded by an erythematic reaction was observed (Figure 1B).

The macroscopic healing pattern of the lesions and skin damages began with scab formation over the wound on the 8th to 12th day after initiation of the treatment. Thereafter, the scab was removed from the lateral part of the wound or simultaneously completely removed at the end of the healing stage (Figure 2).

4.2. Inspecting the Inflammation, Erythema, and Blisters

On the 3rd day of treatment, inflammation was observed in all groups that had received the toxin. There were no significant differences in inflammation, erythema, or blister healing between the treatment and non-treatment groups. Compared to the other groups, on the 10th day of the study, there was less inflammation, erythema, and blister damage in the groups treated with the 10% and 15% *C. pontica* creams (Figure 2).

There were no significant differences in inflammation, erythema, and blister healing between the groups treated with the 10% and 15% *C. pontica* creams, but a significant difference was observed compared to the eucerin group (P < 0.05) (Figure 3).

4.3. Histopathological Findings

Almost 48 hours after the primary application of the toxin, primary destruction (necrosis and wound lesions) of the epidermis and vessel dilation with lymphocyte aggregation was observed in the group without epidermal treatment, indicating the initiation of the inflammation reactions (Figure 4B). Seven days after primary application of the toxin, parts of the wound lesions, in addition to dispersed granulation aggregation and inflammatory reactions, remained.

On the 10th day of treatment in the group treated with the 5% cream, no wound lesions had concisely formed in the epidermis. Furthermore, a slight layer of epidermis and dermal granulation tissue had formed, and collagen precipitation and dispersed lymphocyte aggregation was observed (Figure 4C). In the group treated with the 10% cream on the 10th day of treatment, we observed reformation of the epidermis, complete dermal granulation, tissue formation, collagen precipitation, and no inflammation (Figure 4D). In the group treated with the 15% cream, the epidermis reached the normal level and complete granulation tissue and intact fibroblast strands were observed on the 10th day of treatment (Figure 4E).

5. Discussion

A wound is a physical injury that arises from an opening or break in the skin. Wound healing is a collaborative process involving a variety of cells and matrix components interacting continually toward a common goal (12, 13). This process is characterized by three stages: inflammation, proliferation, and remodeling. The proliferative phase typically demonstrates angiogenesis, collagen deposition, granulation tissue formation, epithelialization, and wound contraction (14, 15). Trichothecenes are secondary metabolites produced by some species of fungi. More than one hundred trichothecenes have been identified in the laboratory, but only about a dozen of these compounds have been found under natural conditions (16, 17). Tricothecenes are classified according to their chemical structures, based on a ring system denominated 12, 13-epoxytrichothec-9-ene with several possible functional groups attached. Deoxynivalenol (DON) and nivalenol (NIV) are the most frequently studied trichothecenes, and diacetoxyscirpenol (DAS) and T-2 toxin are the most toxic among the non-cyclic compounds of the family (18, 19).

In the present study, in comparison to the control and eucerin-treated groups, topical application of *C. pontica* cream caused acceleration in wound healing as indicated by the decreased time to complete epithelization. However, the best results were obtained with the 10% and 15% *C. pontica* extract creams.

To further confirm the quality and maturity of the healing tissue on different days and in the different groups, we microscopically examined histopathological wound sections stained by hematoxylin and eosin (H&E). In many previous studies evaluating wound healing, parameters such as edema, fibrin clots, infiltration of white blood cells and inflammatory cells, intensity of fibroblasts, angiogenesis, texture of granulation, re-epithelization, and collagen intensity have been examined histopathologically (20, 21). In the present study, the histological findings corresponded with the observed reductions in the damaged areas. The H&E stained sections revealed that in the group treated with the 5% cream, on the 12th day of treatment, there were no wound lesions in the epidermis, epidermis and dermal granulation tissue had formed, and collagen precipitation and dispersed lymphocyte aggregation was observed. In the group treated with the 10% cream on the 10th day of treatment, there was no inflammation, and reformation of the epidermis, complete dermal granulation tissue, and collagen precipitation were observed. In the group treated with the 15% cream, normal levels of epidermis, complete granulation tissue, and intact fibroblast strands were observed on the 10th day of treatment.

There is increasing interest in the use of herbal

Figure 1. Photographs of Rabbit Skin Lesions 48 Hours After T-2 Toxin Administration Without any Treatment



(A) and 9 days after T-2 toxin administration and treatment with eucerin (B). Erythema, in [U+FB02] ammation, and tissue necrosis are the characteristic features of the effect of T-2 toxin (A). A central necrosis surrounded by an erythematic reaction is observed (B).

Figure 2. Photographs of Rabbit Skin Lesions in the Groups Treated with 5%, 10%, and 15% Crataegus pontica Cream



(A-C), on the 10th day of treatment. Erythema, inflammation, and partial necrosis are seen in the 5% treatment group. Erythema and inflammation are not seen in the 10% treatment group. In the 15% treatment group, there is no erythema or inflammation and normal skin and hair regrowth are observed.

medicines for wound healing. Flavonoids have significant impacts on wound healing and dermal protection. These components prevent blood and lymph accumulation by regulating micro vessels around the wound lesion area and thus reduce local swelling. They also improve blood circulation and enhance continuous perfusion to facilitate the healing of wound lesions (22, 23). Since the most important class of active ingredients present in *Crataegus* sp. is flavonoids (including hyperoside, vitexin, isoquercetin, luteolin 7-glucoside, and quercetin), it is suggested that the flavonoids present in the available extract must play a crucial role in the treatment of damage caused by T-2 toxin.

A similar study investigated the healing effects of quince seed mucilage on dermal toxicity induced by T-2 toxin. In this study, to induce dermal toxicity, 100 μ g of T-

2 toxin was applied to the shaved skin of rabbits. The obtained results indicate that quince seed mucilage has remarkable healing effects on dermal toxicity caused by T-2 toxin (1).

In the present study, it was observed that doses of both 10% and 15% *C. pontica* cream had similar effects on wound healing, therefore 10% *C. pontica* cream can be suggested as a potential treatment for T-2 toxin-induced dermal toxicity.

Finally, this study confirmed that *C. pontica* cream accelerated wound healing and the time taken for complete healing of damaged skin when compared to the negative control and eucerin only groups. The beneficial effects of the *C. pontica* preparation in the healing of superficial skin wounds may be attributed to its flavonoid content. Furthermore, the present study indicates that this prepara-



Figure 3. Comparisons of The Effect of C. pontica Extract Cream on Erythema, Blisters, and Inflammation and the Time Required for Complete Healing in the Rabbit Groups

Significant differences between the treatment groups (n = 5) and the eucerin group are indicated as (P < 0.05).

tion could be applied for the treatment of skin damage and may be an alternative to the use of other routine treatments.

Acknowledgments

This work was a part of the thesis of Moslem Kiani Aslani which was supported by grant number CH-12 provided by the deputy of research of Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran.

Footnotes

Authors' Contribution: Study concept and design, Heibatullah Kalantari and Ali Asghar Hemmati; analysis and interpretation of data, Hossein Forouzandeh, Mojtaba Kalantar, and Mehdi Goudarzi; drafting of the manuscript, Nasrin Aghel; critical revision of the manuscript for important intellectual content, Moslem Kiyani Aslani; statistical analysis, Mehrnush Musavi, Tahere Shamsi Ehsan.

Conflict of Interest: All authors declare no conflict of interest related to the present work.

Funding/Support: This study was supported by the deputy of research of Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran.



Figure 4. Histopathological Observations (Skin Sections Stained with Hematoxylin and Eosin, Magnification × 400) for the Skin Healing Sites

Intact rabbit skin (A), rabbit skin from the non-treatment group 48 hours after primary T-2 toxin administration (B), rabbit skin from the groups treated with 5%, 10%, and 15% *C. pontica* cream on the 10th day of treatment (C, D, and E, respectively).

References

- Hemmati AA, Kalantari H, Jalali A, Rezai S, Zadeh HH. Healing effect of quince seed mucilage on T-2 toxin-induced dermal toxicity in rabbit. *Exp Toxicol Pathol.* 2012;64(3):181–6. doi: 10.1016/j.etp.2010.08.004. [PubMed: 20832267].
- Chaudhari M, Jayaraj R, Bhaskar AS, Lakshmana Rao PV. Oxidative stress induction by T-2 toxin causes DNA damage and triggers apoptosis via caspase pathway in human cervical cancer cells. *Toxicol*ogy. 2009;262(2):153–61. doi: 10.1016/j.tox.2009.06.002. [PubMed: 19524637].
- Bouaziz C, Abid-Essefi S, Bouslimi A, El Golli E, Bacha H. Cytotoxicity and related effects of T-2 toxin on cultured Vero cells. *Toxi*con. 2006;48(3):343–52. doi: 10.1016/j.toxicon.2006.06.004. [PubMed: 16884754].
- Rizzo AF, Atroshi F, Ahotupa M, Sankari S, Elovaara E. Protective effect of antioxidants against free radical-mediated lipid peroxidation induced by DON or T-2 toxin. *Zentralbl Veterinarmed A*. 1994;**41**(2):81–90. [PubMed: 8091893].
- Li M, Harkema JR, Islam Z, Cuff CF, Pestka JJ. T-2 toxin impairs murine immune response to respiratory reovirus and exacerbates viral bronchiolitis. *Toxicol Appl Pharmacol.* 2006;217(1):76–85. doi: 10.1016/j.taap.2006.08.007. [PubMed: 17005225].
- Kamalavenkatesh P, Vairamuthu S, Balachandran C, Manohar BM, raj GD. Immunopathological effect of the mycotoxins cyclopiazonic acid and T-2 toxin on broiler chicken. *Mycopathologia*. 2005;**159**(2):273–9. doi:10.1007/s11046-004-7321-0. [PubMed: 15770454].
- Hymery N, Sibiril Y, Parent-Massin D. In vitro effects of trichothecenes on human dendritic cells. *Toxicol In Vitro*. 2006;20(6):899–909. doi: 10.1016/j.tiv.2006.01.015. [PubMed: 16517116].
- 8. Valizadeh R, Hemmati AA, Houshmand G, Bayat S, Bahadoram M. Wound healing potential of Althaea officinalis flower mucilage in

rabbit full thickness wounds. *Asian Pac J Trop Biomed*. 2015;**5**(11):937-43.

- 9. Yazdinezhad A, Najafi F, Mousavi A. Pharmacognostic and phytochemical studies of leaves of Crataegus pontica C. Koch. *J Chem Pharm Res.* 2014;6(3).
- Gundogdu M, Ozrenk K, Ercisli S, Kan T, Kodad O, Hegedus A. Organic acids, sugars, vitamin C content and some pomological characteristics of eleven hawthorn species (Crataegus spp.) from Turkey. *Biol Res.* 2014;47:21. doi: 10.1186/0717-6287-47-21. [PubMed: 25028120].
- Javad-Mousavi SA, Hemmati AA, Mehrzadi S, Hosseinzadeh A, Houshmand G, Rashidi Nooshabadi MR, et al. Protective effect of Berberis vulgaris fruit extract against Paraquat-induced pulmonary fibrosis in rats. *Biomed Pharmacother*. 2016;81:329–36. doi: 10.1016/j.biopha.2016.04.027. [PubMed: 27261610].
- Kumar S, Wong PF, Leaper DJ. What is new in wound healing?. Turk J Med Sci. 2004;34(3):147-60.
- Hemmati AA, Houshmand G, Nemati M, Bahadoram M, Dorestan N, Rashidi-Nooshabadi MR, et al. Wound Healing Effects of Persian Oak (Quercus brantii) Ointment in Rats. Jundishapur J Nat Pharm Prod. 2015;10(4).
- Azadi M, Foruozandeh H, Karami L, Khodayar MJ, Rashidi Nooshabadi M, Kalantar M, et al. Comparing the effect of visceral fat and barley seed ash (hordeum vulgare L) with silversulfadiazine on burn wound healing in rats. *Jundishapur J Nat Pharm Prod.* 2015;10(1):ee20670. [PubMed: 25866721].
- Nayak SB, Pinto Pereira L, Maharaj D. Wound healing activity of Carica papaya L. in experimentally induced diabetic rats. *Indian J Exp Biol.* 2007;45(8):739–43. [PubMed: 17877152].
- Langseth W, Rundberget T. Instrumental methods for determination of nonmacrocyclic trichothecenes in cereals, foodstuffs and cultures. *J Chrom A.* 1998;815(1):103–21.
- Etzerodt T, Maeda K, Nakajima Y, Laursen B, Fomsgaard IS, Kimura M. 2,4-dihydroxy-7-methoxy-2H-1,4-benzoxazin-3(4H)-one (DIMBOA) in-

hibits trichothecene production by Fusarium graminearum through suppression of Tri6 expression. *Int J Food Microbiol*. 2015;**214**:123–8. doi: 10.1016/j.ijfoodmicro.2015.07.014. [PubMed: 26276561].

- Oliveira A, Soares LMV, Sawazaki E. Survey of deoxynivalenol, diacetoxyscirpenol, and T2 toxin in popcorn hybrids planted in the state of São Paulo and in popcorn commercialized in the city of Campinas, SP. Food Sci Technol. 2001;21(3):330–3.
- Placinta CM, D'mello JPF, Macdonald AMC. A review of worldwide contamination of cereal grains and animal feed with Fusarium mycotoxins. Animal Feed Sci Technol. 1999;78(1):21–37.
- 20. Gurel MS, Nayci S, Turgut AV, Bozkurt ER. Comparison of the effects of topical fusidic acid and rifamycin on wound healing in rats. *Int Wound*

J. 2015;**12**(1):106–10. doi: 10.1111/iwj.12060. [PubMed: 23489386].

- 21. Diegelmann RF, Evans MC. Wound healing: an overview of acute, fibrotic and delayed healing. *Front Biosci.* 2004;9:283–9. [PubMed: 14766366].
- Budovsky A, Yarmolinsky L, Ben-Shabat S. Effect of medicinal plants on wound healing. *Wound Repair Regen.* 2015;23(2):171–83. doi: 10.1111/wrr.12274. [PubMed: 25703533].
- Vittorazzi C, Endringer DC, Andrade TU, Scherer R, Fronza M. Antioxidant, antimicrobial and wound healing properties of Struthanthus vulgaris. *Pharm Biol.* 2016;54(2):331-7. doi: 10.3109/13880209.2015.1040515. [PubMed: 25915104].