Published online 2018 December 8.

Research Article

Curcumin Inhibited Endothelin-1 mRNA Expression Induced by TGF- β in Bovine Aortic Endothelial Cell

Zahra Keshavarz^{1,2}, Alireza Kheirollah¹, Mohammad-Ali Ghaffari¹ and Hossein Babaahmadi-Rezaei^{1,*}

¹Cellular and Molecular Research Center, Department of Clinical Biochemistry, Faculty of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran ²Student Research Committee, Department of Clinical Biochemistry, Faculty of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

^{*} *Corresponding author*: Cellular and Molecular Research Center, Department of Clinical Biochemistry, Faculty of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran. Tel: +98-6133367543-2591, Fax: +98-6133738632, Email: babaahmadi-h@ajums.ac.ir

Received 2017 September 28; Revised 2018 April 11; Accepted 2018 May 07.

Abstract

Objectives: Curcumin is a plant polyphenol compound used as a traditional supplement in many countries. The potential therapeutic or preventive effects of curcumin may be related to its antioxidant and anti-inflammatory properties. The current in vitro study aimed to investigate the effect of curcumin on the expression of endothelin-1 (ET-1) stimulated by transforming growth factorbeta (TGF- β) in bovine aortic endothelial cells (BAECs).

Methods: Bovine aortic endothelial cells derived from bovine aorta were maintained in low glucose DMEM (Dulbecco's modified Eagle's medium). The time-course and different concentrations of TGF- β (2 ng/mL and 10 ng/mL) were used to evaluate ET-1 mRNA expression. Also, the BAECs were treated with 10 μ M of SB431542 (chemical inhibitor of TGF- β receptor) as positive control and different doses of curcumin (5 μ M, 10 μ M, and 15 μ M). The expression of ET-1 mRNA was quantified by quantitative real-time polymerase chain reaction (qPCR). Statistical analysis was performed by one-way ANOVA with SPSS software.

Results: ET-1 mRNA expression significantly increased in six hours in the TGF- β -treated group (2.79 ± 0.9). SB431542, as well as curcumin 10 μ M and 15 μ M significantly decreased the expression of ET-1 mRNA by 2.3 ± 0.15, 1.5 ± 0.16, and 1.02 ± 0.01, respectively. **Conclusions:** Curcumin downregulated ET-1 mRNA; this result suggested a possible underlying molecular mechanism mediated through ET-1 to exert its anti-inflammatory and antioxidant properties.

Keywords: Endothelin-1, Curcumin, Atherosclerosis, Transforming Growth Factor-eta

1. Background

Atherosclerosis is a chronic inflammatory disease that annually takes the lives of many people worldwide (1, 2). Atherosclerosis is a complex endothelial dysfunction that leads to an inflammatory response, and is induced by elevated and modified low-density lipoproteins (LDL), free radicals, oxidative stress, hypertension, or combinations of these and other factors (3, 4). A major reason for extracellular lipid accumulation in atherosclerotic plaques is binding of LDL to proteoglycan of artery wall into the subendothelial space (5, 6).

TGF- β plays a key role in the maintenance of normal blood vessel wall structure. Specific defects in the genes encoding TGF- β are linked to a range of cardiovascular syndromes including atherosclerosis (7). Studies show that TGF- β enhances proteoglycans expression in vascular smooth muscle cells and results in the formation and development of atherosclerotic plaques (6, 8).

ET-1 is a peptide consisting of 21 amino acids normally produced in physiologic conditions in small amounts by

endothelial cells and is abundantly produced in pathophysiologic conditions by different cell types including endothelial cells, vascular muscle cells, and macrophages (9, 10). ET-1 is a potent vasoconstrictor and pro-inflammatory peptide that plays a role to control and maintain vascular homeostasis. Disturbance in endothelin regulation alters the normal function of vascular cells, which is correlated with the development of atherosclerotic plaques (10). ET-1 is increased in atherosclerotic plaques, as well as elevated in the serum of diabetic patients that have accelerated atherosclerosis (11). ET-1 causes an increase in LDL oxidation through stimulating reactive oxygen species (ROS) production in endothelial cells and vascular smooth muscles, on one hand, and promotes proteoglycans synthesis on the other hand, all of which ultimately lead to atherosclerotic plaques induction (5, 12).

The authors' previous study revealed that (S)-[6]gingerol inhibits proteoglycans synthesis and has a high structural similarity with curcumin (13). In that study, the role of (S)-[6]-gingerol to inhibit TGF- β -induced

Copyright © 2018, Jundishapur Journal of Natural Pharmaceutical Products. This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (http://creativecommons.org/licenses/by-nc/4.0/) which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited.

biglycan synthesis. Curcumin [1,7-bis- (4-hydroxy-3methoxyphenyl)-1,6-heptadiene-3,5-dione] is the major constituent of turmeric powder extracted from the rhizomes of Curcuma longa (13, 14). Curcumin possesses anti-inflammatory, antioxidant, anti-mutagenic, and anti-carcinogenic activities (15, 16).

2. Objectives

The current study aimed to investigate the expression of ET-1 in BAECs induced by TGF- β .

The studies show that curcumin may has a potent protective effect against cardiovascular disease, specifically atherosclerosis. We show that curcumin inhibit ET-1 mRNA expression induced by TGF- β .

3. Methods

Dulbecco modified Eagle's medium (DMEM) and penecilin/streptomycin and fetal bovine serum (FBS) were obtained from Gibco (Invitrogen, Carlsbad, CA, USA). Recombinant TGF- β was purchased from Cell Signaling (Boston, MA, USA); SB431542 and curcumin were purchase from Sigma Aldrich (MO, USA).

Bovine aortic endothelial cells (BAECs) derived from bovine aorta were kindly donated by professor Peter J Little (School of Pharmacy, The University of Queensland, Australia). BAECs were maintained in DMEM with 1 mM glucose, 10% FBS (Invitrogen, Carlsbad, CA, USA), and 1% penicillin-streptomycin solution at 37°C in 5% CO₂. For the experiments, BAECs were seeded into a 70-mm petri dish and maintained until confluence. Then the cells became quiescent by serum starvation for 16 hours prior to treatment. To check the time dependency of TGF- β , BECs were treated with TGF- β (2 ng/mL and 10 ng/mL) for one, two, four, and six hours. To study the curcumin effect on the expression of ET-1 mRNA, BAECs were pretreated with SB431542 (10 μ M) and curcumin (5 μ M, 10 μ M, and 15 μ M) for 30 minutes; then TGF- β (10 ng/mL) was added and the cells were harvested after six hours.

Total cellular RNA was isolated from the cells using the RNeasy[®] Plus Mini Kit (Qiagene, Valencia, CA). The RNA concentration was quantified using a NanoDrop-2000 (Thermo Fischer Scientific). Also, to assess sample quality, 260/280 or 260/230 ratios were measured and considered for each sample. One microgram of total RNA was reverse transcribed with a PrimeScript[™] RT Reagent Kit (TaKaRa, Dalian, China). The mRNA expression of ET-1 and GAPDH (glyceraldehyde-3-phosphate dehydrogenase; as the internal control) were quantified by quantitative real-time polymerase chain reaction (qPCR) (SYBR Green) using the ABI 7500 Fast Real-Time PCR System (Applied Biosystems). The amplification protocol consisted of 30 seconds at 95°C followed by 40 cycles of PCR steps (five seconds denaturation at 95°C, 30 seconds annealing and extension at 57°C). After amplification, a melting curve was acquired to confirm the absence of non-specific products and also determine Tm of each product to calculate accuracy of the study product. Real-time PCR was performed in a final volume of 20 μ L of reaction mixture consisting of 10 μ L SYBR Green Master Mix (Takara), 0.8 μ M of each primer, and 3 μ L of cDNA.

The employed primer sequences were as follows:

ET-1 forward: 5'-TCTGGACATCATCTGGGTCA-3' and reverse 5'-CTTGGCAAAAATTCCAGCAT-3' (17);

GAPDH forward: 5'-AGCCACATCGCTCAGACAC-3', and reverse 5'GCCCAATACGACCAAA-TCC-3' (18).

All the experiments were performed in triplicate. Statistical analysis was performed by one-way ANOVA with SPSS version 19 and P < 0.05 was considered as statistically significant. Normalization of data was performed to adjust a control variation between individual experiments; data were shown as mean \pm standard error of the mean (SEM); curves were fitted using GraphPad Prism software.

4. Results

ET-1 is expressed in endothelial cells. To evaluated antiatherosclerotic properties of curcumin, expression of ET-1 in cells stimulated with TGF- β was measured.

4.1. The Effect of TGF- β on the Expression of ET-1 mRNA

Quantitative real-time PCR showed that the ET-1 mRNA expression significantly increased in two and six hours following the induction by TGF- β (Figure 1A). ET-1 mRNA expression in the TGF- β -treated group was significantly higher after six hours with 10 ng/mL TGF- β compared with that of the control group (Figure 1B). ET-1 mRNA expression showed no significant changes in either groups. This experiment showed that ET-1 mRNA elevated in a dose dependent manner after stimulation with TGF- β .

4.2. The Effect of Curcumin and SB31542 on the Expression of ET-1 mRNA

The current study investigated the effect of curcumin on the expression of ET-1 mRNA. Three different concentrations of curcumin (5 μ M, 10 μ M, and 15 μ M) were employed to identify dose dependency inhibition of curcumin on the expression of ET-1 induced by TGF- β . It was observed that TGF- β increased ET-1 expression. The impact of SB431542 (10 μ M), inhibitor of TGF- β receptor, on ET-1 expression was also examined alongside of curcumin. Quantitative realtime PCR showed that the ET-1 mRNA expression significantly decreased in a dose dependent manner after exposure to curcumin (Figure 2).

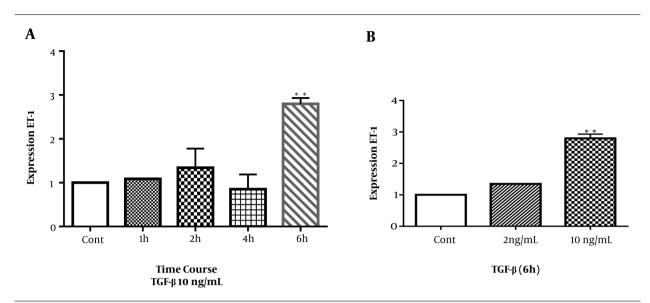


Figure 1. TGF- β stimulated mRNA ET-1 in BAECs. A, The cells were treated with TGF- β (10 ng/mL) at course-time (1 h, 2 h, 4 h, and 6 h), in which a 6-h treatment with TGF- β significantly increased the induction of ET1 mRNA compared with the control group (**P < 0.01); B, the cells were harvested after exposure to two different concentrations of TGF- β (2 ng/mL and 10 ng/mL) for 6 h. ET-1 mRNA at 10 ng/mL of TGF- β was significantly higher than that of the control group (**P < 0.01).

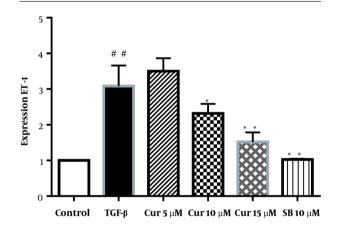


Figure 2. Curcumin inhibited expression of mRNA endothelin-1 by TGF- β . BAECs treated with TGF- β (10 ng/mL, 6 h), curcumin (5 μ M, 10 μ M, 15 μ M) and SB431542 (10 μ M) were pretreated for 30 min before adding TGF- β . ET-1 mRNA expression significantly increased in TGF- β group than the control (##P 0.01). ET-1 mRNA expression significantly decreased in the groups exposed to curcumin and SB431542 compared to TGF- β group (*P < 0.05; **P < 0.01).

5. Discussion

The mechanism of anti-inflammatory/antioxidant activity of curcumin is not known clearly to date. Results of the current study showed that curcumin could downregulate the expression of ET-1 mRNA in BAECs when stimulated with TGF- β . TGF- β is among the factors that can induce ET-1 expression (14, 15). In agreement with this fact, the current study also demonstrated that TGF- β can significantly increase the level of ET-1 mRNA (Figure 1). ET-1 upregulation by TGF- β results in stimulate the synthesis of proteoglycans, increased free radicals and oxidative stress aggravation, the result of which is LDL oxidation, followed by oxidized low-density lipoproteins (LDLs) binding to proteoglycans, and finally resulting in atherosclerotic plaques formation (16, 19). The presence of inhibitors in each part of this pathway could be an approach to improve this disease through inhibitory impact on atherosclerotic plaques formation.

SB431542 is known as a chemical inhibitor of TGF- β receptor that significantly reduces ET-1 expression, but is not considered as a therapeutic compound, while curcumin could decrease ET-1 mRNA expression (Figure 2). Gingerol is a natural compound that inhibits proteoglycans synthesis (13). Curcumin is highly similar to gingerol regarding the chemical structure; therefore, curcumin could affect atherosclerotic plaques formation negatively both by reducing ET-1 expression through TGF- β inhibition, and proteoglycans synthesis inhibition.

Curcumin is a natural therapeutic compound mostly known for its antioxidant feature, and little research is conducted on its anti-sclerotic properties. The current study results indicated that this compound could prevent increased ET-1 stimulated by TGF- β . So, curcumin through inhibition of ET-1 expression may lead to decreased formation and development of atherosclerotic plaque.

Nicholson et al. showed that dietary polyphenols (resveratrol and quercetin) decreased the expression of Et-1 in HUVEC (human umbilical vein endothelial cells) (20). In another study, Virdis et al. found that ET-1 expression was higher in small artery of the subjects with obesity than those of the control; they also indicated that tumor necrosis factor-alpha (TNF- α) associated with imbalance of ET-1/nitric oxide in small arteries of the subjects with obesity (21).

In contrast with the current study results, Farhangkhoee et al. showed that curcumin upregulated ET-1 in diabetic rats (22). Jane Chiu et al. investigated the role of curcumin to prevent abnormality due to diabetes; they showed TGF- β and ET-1 induced in diabetic rats and then curcumin significantly inhibited the expression of TGF- β and ET-1 (23).

How curcumin inhibits TGF- β or how it prevents proteoglycans synthesis are among the questions, which require more studies to respond, and the current study was an introduction to more studies in order to determine the antiatherosclerotic mechanisms of curcumin.

In spite of the current study limitation to use bovine cells that differ from those of human, other studies and also the experiments in authors' lab showed that these cells were closely correlated with human cells.

5.1. Conclusion

Curcumin is known for its antioxidant properties. In previous studies the anti-inflammatory and antiatherosclerotic mechanisms of curcumin were not clearly described. The current study revealed that curcumin could prevent enhancements of ET-1 expression stimulated by TGF- β and showed anti-atherosclerotic properties of curcumin.

Footnotes

Conflict of Interests: The authors declared no conflict of interest.

Funding/Support: The current study was supported by Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran (grant number: CMRC 9419).

References

- Chen WJ, Yin K, Zhao GJ, Fu YC, Tang CK. The magic and mystery of microRNA-27 in atherosclerosis. *Atherosclerosis*. 2012;222(2):314–23. doi: 10.1016/j.atherosclerosis.2012.01.020. [PubMed: 22307089].
- Stoger JL, Gijbels MJ, van der Velden S, Manca M, van der Loos CM, Biessen EA, et al. Distribution of macrophage polarization markers in human atherosclerosis. *Atherosclerosis*. 2012;**225**(2):461-8. doi: 10.1016/j.atherosclerosis.2012.09.013. [PubMed: 23078881].
- Stoll G, Bendszus M. Inflammation and atherosclerosis: Novel insights into plaque formation and destabilization. *Stroke*. 2006;**37**(7):1923-32. doi: 10.1161/01.STR.0000226901.34927.10. [PubMed: 16741184].
- Husain K, Hernandez W, Ansari RA, Ferder L. Inflammation, oxidative stress and renin angiotensin system in atherosclerosis. *World J Biol Chem.* 2015;6(3):209–17. doi: 10.4331/wjbc.v6.i3.209. [PubMed: 26322175]. [PubMed Central: PMC4549761].

- Little PJ, Burch ML, Getachew R, Al-aryahi S, Osman N. Endothelin-1 stimulation of proteoglycan synthesis in vascular smooth muscle is mediated by endothelin receptor transactivation of the transforming growth factor-[beta] type I receptor. J Cardiovasc Pharmacol. 2010;56(4):360–8. doi: 10.1097/FJC.0b013e3181ee6811. [PubMed: 20625315].
- Little PJ, Osman N, O'Brien KD. Hyperelongated biglycan: The surreptitious initiator of atherosclerosis. *Curr Opin Lipidol*. 2008;**19**(5):448– 54. doi: 10.1097/MOL.0b013e32830dd7c4. [PubMed: 18769225].
- Grainger DJ. TGF-beta and atherosclerosis in man. *Cardiovasc Res.* 2007;**74**(2):213–22. doi: 10.1016/j.cardiores.2007.02.022. [PubMed: 17382916].
- Yang SN, Burch ML, Tannock LR, Evanko S, Osman N, Little PJ. Transforming growth factor-beta regulation of proteoglycan synthesis in vascular smooth muscle: Contribution to lipid binding and accelerated atherosclerosis in diabetes. J Diabetes. 2010;2(4):233–42. doi: 10.1111/j.1753-0407.2010.00089.x. [PubMed: 20923499].
- Weng CM, Yu CC, Kuo ML, Chen BC, Lin CH. Endothelin-1 induces connective tissue growth factor expression in human lung fibroblasts by ETAR-dependent JNK/AP-1 pathway. *Biochem Pharmacol.* 2014;88(3):402-11. doi: 10.1016/j.bcp.2014.01.030. [PubMed: 24486572].
- Pernow J, Shemyakin A, Bohm F. New perspectives on endothelin-1 in atherosclerosis and diabetes mellitus. *Life Sci.* 2012;91(13-14):507–16. doi: 10.1016/j.lfs.2012.03.029. [PubMed: 22483688].
- Little PJ, Ivey ME, Osman N. Endothelin-1 actions on vascular smooth muscle cell functions as a target for the prevention of atherosclerosis. *Curr Vasc Pharmacol.* 2008;6(3):195–203. doi: 10.2174/157016108784911966. [PubMed: 18673159].
- Todirita A, Manea A, Manea SA. Endothelin type A receptor mediates endothelin-1-induced upregulation of NADPH oxidase activity in human aortic smooth muscle cells. Ann Rom Soc Cell Biol. 2013;18(1):44.
- Kamato D, Babaahmadi Rezaei H, Getachew R, Thach L, Guidone D, Osman N, et al. (S)-[6]-Gingerol inhibits TGF-β-stimulated biglycan synthesis but not glycosaminoglycan hyperelongation in human vascular smooth muscle cells. *J Pharm Pharmacol.* 2013;65(7):1026–36. doi: 10.1111/jphp.12060.
- Shimada H, Staten NR, Rajagopalan LE. TGF-beta1 mediated activation of Rho kinase induces TGF-beta2 and endothelin-1 expression in human hepatic stellate cells. *J Hepatol.* 2011;54(3):521–8. doi: 10.1016/j.jhep.2010.07.026. [PubMed: 21087804].
- Sharifat N, Mohammad Zadeh G, Ghaffari MA, Dayati P, Kamato D, Little PJ, et al. Endothelin-1 (ET-1) stimulates carboxy terminal Smad2 phosphorylation in vascular endothelial cells by a mechanism dependent on ET receptors and de novo protein synthesis. *J Pharm Pharma*col. 2017;69(1):66–72. doi: 10.1111/jphp.12654. [PubMed: 27905105].
- Carmona-Cuenca I, Herrera B, Ventura JJ, Roncero C, Fernandez M, Fabregat I. EGF blocks NADPH oxidase activation by TGF-beta in fetal rat hepatocytes, impairing oxidative stress, and cell death. *J Cell Physiol.* 2006;**207**(2):322–30. doi: 10.1002/jcp.20568. [PubMed: 16331683].
- 17. Murakami M, Nemoto T, Niwa H, Kushikata T, Ono K, Watanabe H, et al. Involvement of endothelin-1 in adrenal catecholamine regulation. *Hirosaki Med.* 2014;**65**:218–26.
- Li SW, Wang CY, Jou YJ, Yang TC, Huang SH, Wan L, et al. SARS coronavirus papain-like protease induces Egr-1-dependent up-regulation of TGF-beta1 via ROS/p38 MAPK/STAT3 pathway. *Sci Rep.* 2016;6:25754. doi: 10.1038/srep25754. [PubMed: 27173006]. [PubMed Central: PMC4865725].
- Das R, Roy A, Dutta N, Majumder HK. Reactive oxygen species and imbalance of calcium homeostasis contributes to curcumin induced programmed cell death in Leishmania donovani. *Apoptosis.* 2008;**13**(7):867–82. doi: 10.1007/s10495-008-0224-7. [PubMed: 18506627].
- Nicholson SK, Tucker GA, Brameld JM. Effects of dietary polyphenols on gene expression in human vascular endothelial cells. *Proc Nutr Soc.* 2008;67(1):42-7. doi: 10.1017/S0029665108006009. [PubMed: 18234130].

- 21. Virdis A, Duranti E, Rossi C, Dell'Agnello U, Santini E, Anselmino M, et al. Tumour necrosis factor-alpha participates on the endothelin-1/nitric oxide imbalance in small arteries from obese patients: Role of perivascular adipose tissue. *Eur Heart J.* 2015;**36**(13):784–94. doi: 10.1093/eurheartj/ehu072. [PubMed: 24578389].
- 22. Farhangkhoee H, Khan ZA, Chen S, Chakrabarti S. Differential effects of curcumin on vasoactive factors in the diabetic rat heart. *Nutr Metab*

(Lond). 2006;**3**:27. doi: 10.1186/1743-7075-3-27. [PubMed: 16848894]. [PubMed Central: PMC1543622].

Chiu J, Khan ZA, Farhangkhoee H, Chakrabarti S. Curcumin prevents diabetes-associated abnormalities in the kidneys by inhibiting p300 and nuclear factor-kappaB. *Nutrition*. 2009;25(9):964–72. doi: 10.1016/j.nut.2008.12.007. [PubMed: 19268536].