



Protection Against Radiation-Induced Duox1 and Duox2 Upregulation in Rat's Lung Tissues by a Combination of Curcumin and L-Selenomethionine

Masoud Najafi¹, Peyman Amini², Hana Saffar³, Sedighe Kolivand⁴, Elahe Motevaseli^{4,*}, Saeed Rezapoor², Mohsen Cheki⁵, Dheyauldeen Shabeeb^{6,7} and Ahmed Eleojo Musa⁸

¹Medical Technology Research Center, Institute of Health Technology, Kermanshah University of Medical Sciences, Kermanshah, Iran

²Department of Radiology, Faculty of Paramedical, Tehran University of Medical Sciences, Tehran, Iran

³Clinical and Anatomical Pathologist, Imam Khomeini Hospital Complex, Tehran University of Medical Science, Tehran, Iran

⁴Department of Molecular Medicine, School of Advanced Technologies in Medicine, Tehran University of Medical Sciences, Tehran, Iran

⁵Department of Radiologic Technology, Faculty of Paramedicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

⁶Department of Medical Physics and Biomedical Engineering, Faculty of Medicine, International Campus, Tehran University of Medical Sciences, Tehran, Iran

⁷Department of Physiology, College of Medicine, University of Misan, Misan, Iraq

⁸Research Center for Molecular and Cellular Imaging, International Campus, Tehran University of Medical Sciences, Tehran, Iran

*Corresponding author: Department of Molecular Medicine, School of Advanced Technologies in Medicine, Tehran University of Medical Sciences, Tehran, Iran. Email: e_motevaseli@tums.ac.ir

Received 2018 July 07; Revised 2018 October 09; Accepted 2019 February 27.

Abstract

Background: It has been proposed that increased levels of pro-inflammatory and pro-fibrotic cytokines play a key role in radiation-induced lung injury. Interleukin-4 (IL-4) and IL-13 are two pro-fibrotic cytokines that promote the production of free radicals through stimulation of Duox1 and Duox2. In this experimental study, we aimed to evaluate the expression of IL4Ra1, Duox1, IL13Ra2, and Duox2 genes following rat's lung irradiation.

Objectives: Also, we detected the modulatory effect of a combination of curcumin and L-selenomethionine on the expression of these genes.

Methods: Twenty male rats were divided into four groups as G1: control (no treatment or radiation); G2: treatment with a combination of curcumin and L-selenomethionine; G3: radiation; G4: radiation plus a combination of curcumin and L-selenomethionine. sixty-seven days after irradiation, rats were killed for detecting the expression of IL4Ra1, IL13Ra2, Duox1, and Duox2.

Results: The results showed no detectable expression for IL13Ra2, while the expression of IL4Ra1, Duox1, and Duox2 was increased. Treatment with a combination of curcumin and L-selenomethionine could attenuate the expression of these genes.

Conclusions: This study proposes that upregulation of Duox1 and Duox2 may be involved in radiation-induced lung injury. Treatment with a combination of curcumin and L-selenomethionine may be useful for the mitigation of lung injury through modulation of these genes.

Keywords: Radiation, Lung, Pneumonitis, Curcumin, L-selenomethionine, Dual Oxidase

1. Background

Lung tissue is one of the most sensitive organs to ionizing radiation. The main side effects of lung exposure to radiation include acute inflammation (pneumonitis) and late fibrosis, which may appear months to years following exposure (1). These side effects may pose a threat to the lives of patients who had undergone radiotherapy for chest cancer as well as for people who have been exposed to an accidental nuclear or radiological event (2, 3). In recent years, several studies have been conducted to develop effective agents for better amelioration of radiation injury

(4-6). Amifostine is an FDA-approved radioprotector for the management of xerostomia in patients with head and neck cancer (7). However, its radioprotective effect is limited to some organs (8). In addition, its high toxicity is the main limiting factor for clinical applications. Some studies reported termination of the radiotherapy procedure, resulting from high toxicity of amifostine (9). In this situation, amifostine treatment may lead to the reduction of therapeutic outcome due to the repopulation of tumor cells (10). Hence, for effective alleviation of the complications to normal tissues, it is necessary to develop low toxic agents with suitable radioprotective effects (11). It is also impor-

tant that these agents do not interfere with the eradication of cancer cells by ionizing radiation (12).

Knowledge of the mechanisms involved in radiation-induced lung injury can aid the development of new compounds for better radioprotection of injured organs (13, 14). Studies have proposed that mechanisms of radiation injury can be various in different organs. Emerging evidence from published studies have shown that there is an important interrelationship between inflammatory responses and reduction/oxidation (redox) interactions, which mediate radiation toxicity in several organs (15, 16). However, signaling pathways for these interactions may be different. It has been confirmed that an increased level of both inflammatory and fibrotic cytokines such as IL-1, IL-2, IL-6, IL-8, IL-4, IL-13, IL-33, TNF- α , TGF- β , and IFN- γ are involved in the late effects of lung injury by ionizing radiation (17). On the other hand, it is well-known that these cytokines, through upregulation of genes involved in the redox system such as NADPH oxidase, COX-2, iNOS, lipoxygenases, and mitochondria, stimulate continuous production of free radicals, including both reactive oxygen species (ROS) and reactive nitrogen species (RNS) (18-22). So far, studies have confirmed the role of some of these genes, such as NADPH oxidase 1 (NOX1), NOX4, COX-2, iNOS, and mitochondria, in radiation lung injury (23). However, the roles of some others, such as dual oxidases (Duox1 and Duox2), remain to be elucidated.

With regards to the above-mentioned points, it is important to target both inflammatory and fibrotic processes, as well as oxidative injury, for effective protection of the lung against ionizing radiation. Curcumin is a potent modulator of immune responses that can alleviate both inflammation and fibrosis (24). On the other hand, L-selenomethionine is a potent antioxidant that has been shown to be more effective for the amelioration of radiation-induced DNA damage (25).

2. Objectives

In the present study, we aimed to detect the regulation of IL-4Ra1, Duox2, IL-13Ra2, and Duox1 gene expression following rat's lung irradiation and treatment with a combination of curcumin and L-selenomethionine.

3. Methods

3.1. Drug Treatment and Irradiation

Both curcumin and L-selenomethionine were purchased from Sigma Aldrich (USA). L-selenomethionine was dissolved in distilled water to form a concentration of 0.16 mg per each milliliter. Curcumin was dissolved in 20%

ethanol at a concentration of 30 mg per each milliliter. Treatment began a day before radiation exposure. L-selenomethionine was administered through intraperitoneal injection (IP) with a dose of 0.8 mg/kg (26). Immediately, curcumin was administered orally in a 150 mg/kg body weight (27). The protocol was continued for five consecutive days. Prior to irradiation, the rats were anesthetized using a combination of Ketamine and Xylazine for fixation under the source of gamma rays. Irradiation was done with 15 Gy from a Cobalt-60 gamma source at a dose rate of 109 cGy/min in the supine position (PA) in a field size 6 \times 6 cm. The other field size area was shield using lead block.

3.2. Experimental Design

This study involved 4 groups of 5 rats in each group, including G1: control: this group did not receive any radiation or drug treatment, including curcumin and L-selenomethionine; G2: treatment with curcumin and L-selenomethionine: this group received both curcumin and L-selenomethionine at 150 mg/kg and 0.8 mg/kg for five consecutive days; G3: irradiation: this group only received 15 Gy gamma rays to their chest; and G4: irradiation plus curcumin and L-selenomethionine: this group received curcumin and L-selenomethionine 24 hours before irradiation and five consecutive days afterward. On the day of irradiation, both curcumin and L-selenomethionine were administered 30 minutes before exposure to radiation. Sixty-seven days after irradiation, all rats were sacrificed, and their lung tissues were removed after chest opening. The Lungs were frozen at -70°C for Real-time PCR.

3.3. Real-time PCR

The lung tissues were homogenized in TRIzol solution (Takara, Japan), and then total RNA was obtained. Then, cDNA was synthesized for all samples using a thermocycler device and a cDNA Synthesis Kit (GeneAll, South Korea). The primers used in this study were first designed using the Genrunner software, followed by blasting all sequences in NCBI for confirmation. The sequence of primers is shown in Table 1. Real-time PCR was done using Applied Biosystems real-time PCR (USA). Moreover, PGM1 was chosen as the internal control gene or housekeeping.

Real-time PCR was performed in duplicate, and the amplifications were done using Master Mix Green (Ampliqon). Real-Time PCR efficiency for all genes, including Duox1, Duox2, IL4Ra1, IL13Ra2, and PGM1, was calculated using a linear regression described by Pfaffl (28).

Table 1. The Sequences of Primers for Real-time PCR

Gene	Forward Sequence	Reverse Sequence
<i>IL-13Ra2</i>	TCGTGTTAGCGGATGGGGAT	GCCTGGAAGCCTGGATCTCTA
<i>Duox1</i>	AAGAAAGGAAGCATCAACACCC	ACCAGGGCAGTCAGGAAGAT
<i>IL-4R1</i>	GAGTGAGTGGAGTCCCAGCATC	GCTGAAGTAACAGGTCAGGC
<i>Duox2</i>	AGTCTCATCTCACCCGGA	GTAACACACAGTGTGGCG
<i>PGM1</i>	CATGATTCTGGGCAAGCACG	GCCAGTTGGGGTCTCATACAAA

3.4. Statistical Analysis

In this study, we used SPSS software version 24 for all statistical analyses. A student *t*-test was performed to evaluate significant differences in gene expression. For all analyses, a *P*-value < 0.05 was considered statistically significant.

4. Results

The results of IL4Ra1 gene expression showed that when rats' lung tissues received irradiation by gamma rays, the expression of this gene increased by 5.94 ± 1.74 fold compared to the control group ($P < 0.05$). When rats were treated with a combination of curcumin and L-selenomethionine, the expression of IL4Ra1 was reduced by 2.21 ± 0.59 fold compared to the non-treated irradiated rats significantly ($P < 0.05$). Treatment with a combination of curcumin and L-selenomethionine did not cause any significant change in the expression of IL4Ra1 compared to the control group (1.48 ± 0.52). Real-time PCR results showed no detectable expression for IL13Ra2. The expression of Duox1 when rats received gamma rays to lung tissues was increased by 10.24 ± 2.61 fold compared to the control group ($P < 0.05$). When rats received a combination of curcumin and L-selenomethionine, the expression of Duox1 was attenuated 4.13 ± 1.07 fold compared to the rats irradiated without curcumin and L-selenomethionine treatment ($P < 0.05$). Administration of a combination of curcumin and L-selenomethionine alone did not change the expression of Duox1 (1.02 ± 0.19 fold).

The results of Duox2 gene expression showed a significant increase in the expression of this gene following irradiation of rat's lung tissues (11.70 ± 2.47 fold) ($P < 0.05$). When rats were treated with a combination of curcumin and L-selenomethionine before and after the irradiation, the expression of Duox2 was attenuated significantly (4.37 ± 0.54 fold) compared to the rats irradiated without treatment ($P < 0.05$). Similar to other genes, the expression of Duox2 did not change in rats treated with a combination of curcumin and L-selenomethionine alone (1.18 ± 0.32 fold) (Figure 1).

5. Discussion

Emerging evidence in recent years has confirmed that the upregulation of some genes, including pro-inflammatory and pro-fibrotic cytokines, plays a central role in the development of radiation-induced lung injury (15). Some studies proposed that modulation of some pathways, including IL-4 signaling, may help mitigate radiation-induced injury in the lung and other tissues such as the heart (29, 30). In addition, supplements with some antioxidants have confirmed that chronic oxidative damage plays a key role in the late effects of radiation on the lung (17). Ameziane-El-Hassani et al. (31) showed that IL-4 and IL-13 could stimulate the upregulation of Duox1 and Duox2, leading to the continuous production of free radicals following exposure of thyroid cells to radiation. In addition, they showed that the upregulation of these genes is associated with genomic instability, which may increase the risk of carcinogenesis. In this study, we hypothesized that irradiation of rat's lung tissues might lead to the upregulation of Duox1 and Duox2 gene expression. Also, we hypothesized that the expression of these genes in the lung may be dependent on the IL13Ra2 and IL4Ra1 expressions. Results of our study showed that irradiation of lung tissues led to a significant increase in the expression of IL4Ra1, but did not show a detectable expression for IL-13. Also, results showed an increase in the expression of both Duox1 and Duox2.

In the present study, we detected the modulatory effect of curcumin and L-selenomethionine before and after irradiation on the expression of IL4Ra1, Duox1, and Duox2. The results showed that this combination reduces the expression of all three genes. This may indicate that the combination of curcumin and L-selenomethionine may be useful for the mitigation of radiation-induced lung injury through modulation of pro-oxidant enzymes such as Duox1 and Duox2. Previous studies have shown that curcumin can suppress several inflammatory mediators, including inflammatory cytokines, transcription factors such as NF- κ B and STATs, and also pro-oxidant enzymes such as iNOS and COX-2 (32-34). On the other hand, L-selenomethionine has been shown to mitigate radiation-induced injury in some organs such as the kidney, bone marrow, and gastrointestinal system (35, 36). The combination of curcumin and L-selenomethionine may be a potent anti-inflammation and antioxidant compound for amelioration of radiation injury.

5.1. Conclusions

This study showed that exposing rat's lung tissues to a high dose of ionizing radiation leads to upregulation of IL4ra1, Duox1, and Duox2 gene expression. However, we

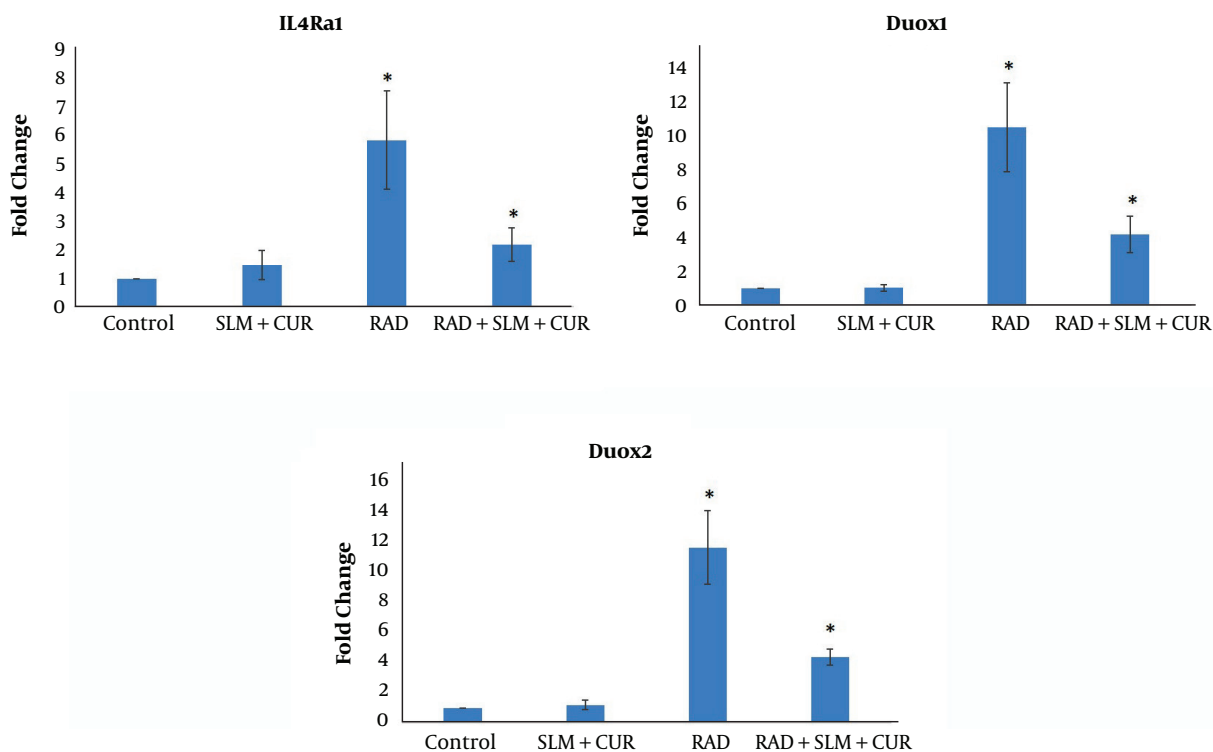


Figure 1. The expression of IL4Ra1, Duox1, and Duox2 following rat's lung irradiation and administration of a combination of curcumin and L-selenomethionine. A, The radiation group was compared to the control group; and B, the radiation plus treatment group was compared to the radiation group, *, $P < 0.05$.

did not detect the regulation of IL13Ra2. Treatment of rats with a combination of curcumin and L-selenomethionine could attenuate the expression of these genes. These results indicate that upregulation of Duox1 and Duox2 may be involved in the late effect of radiation on the lung tissue. Eventually, our results indicated that a combination of curcumin and L-selenomethionine may be useful for the mitigation of lung injury through modulation of these genes.

Footnotes

Authors' Contribution: All authors were involved in this project.

Conflict of Interests: The authors declare that they have no competing interests.

Ethical Approval: This study was approved by the Ethics Committee of the Tehran University of Medical Sciences on Animal Care.

Funding/Support: Tehran University of Medical Sciences and health service, grant number 36668.

References

- Rezaeyan A, Fardid R, Haddadi GH, Takhshid MA, Hosseinzadeh M, Najafi M, et al. Evaluating Radioprotective Effect of Hesperidin on Acute Radiation Damage in the Lung Tissue of Rats. *J Biomed Phys Eng.* 2016;6(3):165-74. [PubMed: 27853724]. [PubMed Central: PMC5106549].
- Bahig H, Filion E, Vu T, Chalaoui J, Lambert L, Roberge D, et al. Severe radiation pneumonitis after lung stereotactic ablative radiation therapy in patients with interstitial lung disease. *Pract Radiat Oncol.* 2016;6(5):367-74. doi: 10.1016/j.prro.2016.01.009. [PubMed: 27068780].
- Yahyapour R, Amini P, Rezapour S, Cheki M, Rezaeyan A, Farhood B, et al. Radiation-induced inflammation and autoimmune diseases. *Mil Med Res.* 2018;5(1):9. doi: 10.1186/s40779-018-0156-7. [PubMed: 29554942]. [PubMed Central: PMC5859747].
- Yahyapour R, Shabeeb D, Cheki M, Musa AE, Farhood B, Rezaeyan A, et al. Radiation Protection and Mitigation by Natural Antioxidants and Flavonoids: Implications to Radiotherapy and Radiation Disasters. *Curr Mol Pharmacol.* 2018;11(4):285-304. doi: 10.2174/1874467211666180619125653. [PubMed: 29921213].
- Medhora M, Gao F, Jacobs ER, Moulder JE. Radiation damage to the lung: mitigation by angiotensin-converting enzyme (ACE) inhibitors. *Respirology.* 2012;17(1):66-71. doi: 10.1111/j.1440-1843.2011.02092.x. [PubMed: 22023053]. [PubMed Central: PMC3245332].
- Yahyapour R, Amini P, Rezapour S, Rezaeyan A, Farhood B, Cheki M, et al. Targeting of Inflammation for Radiation Protection and Mitigation. *Curr Mol Pharmacol.* 2018;11(3):203-10. doi: 10.2174/1874467211666171108165641. [PubMed: 29119941].

7. Brizel DM, Wasserman TH, Henke M, Strnad V, Rudat V, Monnier A, et al. Phase III randomized trial of amifostine as a radioprotector in head and neck cancer. *J Clin Oncol*. 2000;**18**(19):3339-45. doi: [10.1200/JCO.2000.18.19.3339](https://doi.org/10.1200/JCO.2000.18.19.3339). [PubMed: [11013273](https://pubmed.ncbi.nlm.nih.gov/11013273/)].
8. Brizel DM, Overgaard J. Does amifostine have a role in chemoradiation treatment? *Lancet Oncol*. 2003;**4**(6):378-81. doi: [10.1016/S1470-2045\(03\)01132-X](https://doi.org/10.1016/S1470-2045(03)01132-X). [PubMed: [12788413](https://pubmed.ncbi.nlm.nih.gov/12788413/)].
9. Bourhis J, De Crevoisier R, Abdulkarim B, Deutsch E, Lusinchi A, Luboinski B, et al. A randomized study of very accelerated radiotherapy with and without amifostine in head and neck squamous cell carcinoma. *Int J Radiat Oncol Biol Phys*. 2000;**46**(5):1105-8. doi: [10.1016/S0360-3016\(99\)00532-5](https://doi.org/10.1016/S0360-3016(99)00532-5). [PubMed: [10725619](https://pubmed.ncbi.nlm.nih.gov/10725619/)].
10. Rades D, Fehlauer F, Bajrovic A, Mahlmann B, Richter E, Alberti W. Serious adverse effects of amifostine during radiotherapy in head and neck cancer patients. *Radiother Oncol*. 2004;**70**(3):261-4. doi: [10.1016/j.radonc.2003.10.005](https://doi.org/10.1016/j.radonc.2003.10.005). [PubMed: [15064010](https://pubmed.ncbi.nlm.nih.gov/15064010/)].
11. Verma V. Relationship and interactions of curcumin with radiation therapy. *World J Clin Oncol*. 2016;**7**(3):275-83. doi: [10.5306/wjco.v7.i3.275](https://doi.org/10.5306/wjco.v7.i3.275). [PubMed: [27298767](https://pubmed.ncbi.nlm.nih.gov/27298767/)]. [PubMed Central: [PMC4896895](https://pubmed.ncbi.nlm.nih.gov/PMC4896895/)].
12. Najafi M, Motevaseli E, Shirazi A, Geraily G, Rezaeyan A, Norouzi F, et al. Mechanisms of inflammatory responses to radiation and normal tissues toxicity: clinical implications. *Int J Radiat Biol*. 2018;**94**(4):335-56. doi: [10.1080/09553002.2018.1440092](https://doi.org/10.1080/09553002.2018.1440092). [PubMed: [29504497](https://pubmed.ncbi.nlm.nih.gov/29504497/)].
13. Ding NH, Li JJ, Sun LQ. Molecular mechanisms and treatment of radiation-induced lung fibrosis. *Curr Drug Targets*. 2013;**14**(11):1347-56. doi: [10.2174/1389450113149990198](https://doi.org/10.2174/1389450113149990198). [PubMed: [23909719](https://pubmed.ncbi.nlm.nih.gov/23909719/)]. [PubMed Central: [PMC4156316](https://pubmed.ncbi.nlm.nih.gov/PMC4156316/)].
14. Zhao W, Robbins ME. Inflammation and chronic oxidative stress in radiation-induced late normal tissue injury: therapeutic implications. *Curr Med Chem*. 2009;**16**(2):130-43. doi: [10.2174/092986709787002790](https://doi.org/10.2174/092986709787002790). [PubMed: [19149566](https://pubmed.ncbi.nlm.nih.gov/19149566/)].
15. Farhood B, Goradel NH, Mortezaei K, Khanlarkhani N, Salehi E, Nash-taei MS, et al. Inter-cellular communications-redox interactions in radiation toxicity; potential targets for radiation mitigation. *J Cell Commun Signal*. 2019;**13**(1):3-16. doi: [10.1007/s12079-018-0473-3](https://doi.org/10.1007/s12079-018-0473-3). [PubMed: [29911259](https://pubmed.ncbi.nlm.nih.gov/29911259/)]. [PubMed Central: [PMC6381372](https://pubmed.ncbi.nlm.nih.gov/PMC6381372/)].
16. Yahyapour R, Motevaseli E, Rezaeyan A, Abdollahi H, Farhood B, Cheki M, et al. Reduction-oxidation (redox) system in radiation-induced normal tissue injury: molecular mechanisms and implications in radiation therapeutics. *Clin Transl Oncol*. 2018;**20**(8):975-88. doi: [10.1007/s12094-017-1828-6](https://doi.org/10.1007/s12094-017-1828-6). [PubMed: [29318449](https://pubmed.ncbi.nlm.nih.gov/29318449/)].
17. Pietrofesa R, Turowski J, Tyagi S, Dukes F, Arguiri E, Busch TM, et al. Radiation mitigating properties of the lignan component in flaxseed. *BMC Cancer*. 2013;**13**:179. doi: [10.1186/1471-2407-13-179](https://doi.org/10.1186/1471-2407-13-179). [PubMed: [23557217](https://pubmed.ncbi.nlm.nih.gov/23557217/)]. [PubMed Central: [PMC3636021](https://pubmed.ncbi.nlm.nih.gov/PMC3636021/)].
18. Yahyapour R, Motevaseli E, Rezaeyan A, Abdollahi H, Farhood B, Cheki M, et al. Mechanisms of Radiation Bystander and Non-Targeted Effects: Implications to Radiation Carcinogenesis and Radiotherapy. *Curr Radiopharm*. 2018;**11**(1):34-45. doi: [10.2174/18744710116666171229123130](https://doi.org/10.2174/18744710116666171229123130). [PubMed: [29284398](https://pubmed.ncbi.nlm.nih.gov/29284398/)].
19. Bedard K, Krause KH. The NOX family of ROS-generating NADPH oxidases: physiology and pathophysiology. *Physiol Rev*. 2007;**87**(1):245-313. doi: [10.1152/physrev.00044.2005](https://doi.org/10.1152/physrev.00044.2005). [PubMed: [17237347](https://pubmed.ncbi.nlm.nih.gov/17237347/)].
20. Cho HJ, Lee WH, Hwang OMH, Sonntag WE, Lee YW. Role of NADPH oxidase in radiation-induced pro-oxidative and pro-inflammatory pathways in mouse brain. *Int J Radiat Biol*. 2017;**93**(11):1257-66. doi: [10.1080/09553002.2017.1377360](https://doi.org/10.1080/09553002.2017.1377360). [PubMed: [28880721](https://pubmed.ncbi.nlm.nih.gov/28880721/)]. [PubMed Central: [PMC6080279](https://pubmed.ncbi.nlm.nih.gov/PMC6080279/)].
21. Choi SH, Kim M, Lee HJ, Kim EH, Kim CH, Lee YJ. Effects of NOX1 on fibroblastic changes of endothelial cells in radiation-induced pulmonary fibrosis. *Mol Med Rep*. 2016;**13**(5):4135-42. doi: [10.3892/mmr.2016.5090](https://doi.org/10.3892/mmr.2016.5090). [PubMed: [27053172](https://pubmed.ncbi.nlm.nih.gov/27053172/)]. [PubMed Central: [PMC4838118](https://pubmed.ncbi.nlm.nih.gov/PMC4838118/)].
22. Yahyapour R, Salajegheh A, Safari A, Amini P, Rezaeyan A, Amraee A, et al. Radiation-induced Non-targeted Effect and Carcinogenesis; Implications in Clinical Radiotherapy. *J Biomed Phys Eng*. 2018;**8**(4):435-46. [PubMed: [30568933](https://pubmed.ncbi.nlm.nih.gov/30568933/)]. [PubMed Central: [PMC6280111](https://pubmed.ncbi.nlm.nih.gov/PMC6280111/)].
23. Chai Y, Calaf GM, Zhou H, Ghandhi SA, Elliston CD, Wen G, et al. Radiation induced COX-2 expression and mutagenesis at non-targeted lung tissues of gpt delta transgenic mice. *Br J Cancer*. 2013;**108**(1):91-8. doi: [10.1038/bjc.2012.498](https://doi.org/10.1038/bjc.2012.498). [PubMed: [23321513](https://pubmed.ncbi.nlm.nih.gov/23321513/)]. [PubMed Central: [PMC3553512](https://pubmed.ncbi.nlm.nih.gov/PMC3553512/)].
24. Basnet P, Skalko-Basnet N. Curcumin: an anti-inflammatory molecule from a curry spice on the path to cancer treatment. *Molecules*. 2011;**16**(6):4567-98. doi: [10.3390/molecules16064567](https://doi.org/10.3390/molecules16064567). [PubMed: [21642934](https://pubmed.ncbi.nlm.nih.gov/21642934/)]. [PubMed Central: [PMC6264403](https://pubmed.ncbi.nlm.nih.gov/PMC6264403/)].
25. Sieber F, Muir SA, Cohen EP, Fish BL, Mader M, Schock AM, et al. Dietary selenium for the mitigation of radiation injury: effects of selenium dose escalation and timing of supplementation. *Radiat Res*. 2011;**176**(3):366-74. doi: [10.1667/rr2456.1](https://doi.org/10.1667/rr2456.1). [PubMed: [21867430](https://pubmed.ncbi.nlm.nih.gov/21867430/)]. [PubMed Central: [PMC3237945](https://pubmed.ncbi.nlm.nih.gov/PMC3237945/)].
26. Patchen ML, Macvittie TJ, Weiss JF. Combined modality radioprotection: The use of glucan and selenium with WR-2721. *Int J Radiat Oncol Biol Phys*. 1990;**18**(5):1069-75. doi: [10.1016/0360-3016\(90\)90442-m](https://doi.org/10.1016/0360-3016(90)90442-m).
27. Wang BF, Cui ZW, Zhong ZH, Sun YH, Sun QF, Yang GY, et al. Curcumin attenuates brain edema in mice with intracerebral hemorrhage through inhibition of AQP4 and AQP9 expression. *Acta Pharmacol Sin*. 2015;**36**(8):939-48. doi: [10.1038/aps.2015.47](https://doi.org/10.1038/aps.2015.47). [PubMed: [26119880](https://pubmed.ncbi.nlm.nih.gov/26119880/)]. [PubMed Central: [PMC4564884](https://pubmed.ncbi.nlm.nih.gov/PMC4564884/)].
28. Pfaffl MW. A new mathematical model for relative quantification in real-time RT-PCR. *Nucleic Acids Res*. 2001;**29**(9). e45. doi: [10.1093/nar/29.9.e45](https://doi.org/10.1093/nar/29.9.e45). [PubMed: [11328886](https://pubmed.ncbi.nlm.nih.gov/11328886/)]. [PubMed Central: [PMC55695](https://pubmed.ncbi.nlm.nih.gov/PMC55695/)].
29. Groves AM, Johnston CJ, Misra RS, Williams JP, Finkelstein JN. Effects of IL-4 on pulmonary fibrosis and the accumulation and phenotype of macrophage subpopulations following thoracic irradiation. *Int J Radiat Biol*. 2016;**92**(12):754-65. doi: [10.1080/09553002.2016.1222094](https://doi.org/10.1080/09553002.2016.1222094). [PubMed: [27539247](https://pubmed.ncbi.nlm.nih.gov/27539247/)]. [PubMed Central: [PMC5247271](https://pubmed.ncbi.nlm.nih.gov/PMC5247271/)].
30. Kolivand S, Amini P, Saffar H, Rezapour S, Motevaseli E, Najafi M, et al. Evaluating the Radioprotective Effect of Curcumin on Rat's Heart Tissues. *Curr Radiopharm*. 2019;**12**(1):23-8. doi: [10.2174/18744710116666180831101459](https://doi.org/10.2174/18744710116666180831101459). [PubMed: [30173659](https://pubmed.ncbi.nlm.nih.gov/30173659/)].
31. Ameziane-El-Hassani R, Talbot M, de Souza Dos Santos MC, Al Ghuzlan A, Hartl D, Bidart JM, et al. NADPH oxidase DUOX1 promotes long-term persistence of oxidative stress after an exposure to irradiation. *Proc Natl Acad Sci U S A*. 2015;**112**(16):5051-6. doi: [10.1073/pnas.1420707112](https://doi.org/10.1073/pnas.1420707112). [PubMed: [25848056](https://pubmed.ncbi.nlm.nih.gov/25848056/)]. [PubMed Central: [PMC4413347](https://pubmed.ncbi.nlm.nih.gov/PMC4413347/)].
32. Cho YJ, Yi CO, Jeon BT, Jeong YY, Kang GM, Lee JE, et al. Curcumin attenuates radiation-induced inflammation and fibrosis in rat lungs. *Korean J Physiol Pharmacol*. 2013;**17**(4):267-74. doi: [10.4196/kjpp.2013.17.4.267](https://doi.org/10.4196/kjpp.2013.17.4.267). [PubMed: [23946685](https://pubmed.ncbi.nlm.nih.gov/23946685/)]. [PubMed Central: [PMC3741482](https://pubmed.ncbi.nlm.nih.gov/PMC3741482/)].
33. Deguchi A. Curcumin targets in inflammation and cancer. *Endocr Metab Immune Disord Drug Targets*. 2015;**15**(2):88-96. doi: [10.2174/1871530315666150316120458](https://doi.org/10.2174/1871530315666150316120458). [PubMed: [25772169](https://pubmed.ncbi.nlm.nih.gov/25772169/)].
34. Menon VP, Sudheer AR. Antioxidant and anti-inflammatory properties of curcumin. *Adv Exp Med Biol*. 2007;**595**:105-25. doi: [10.1007/978-0-387-46401-5_3](https://doi.org/10.1007/978-0-387-46401-5_3). [PubMed: [17569207](https://pubmed.ncbi.nlm.nih.gov/17569207/)].
35. Brown SL, Kolozsvary A, Liu J, Jenrow KA, Ryu S, Kim JH. Antioxidant diet supplementation starting 24 hours after exposure reduces radiation lethality. *Radiat Res*. 2010;**173**(4):462-8. doi: [10.1667/RR1716.1](https://doi.org/10.1667/RR1716.1). [PubMed: [20334518](https://pubmed.ncbi.nlm.nih.gov/20334518/)]. [PubMed Central: [PMC2874934](https://pubmed.ncbi.nlm.nih.gov/PMC2874934/)].
36. Bagheri H, Rezapour S, Najafi M, Motevaseli E, Shekarchi B, Cheki M, et al. Protection Against Radiation-Induced Micronuclei in Rat Bone Marrow Erythrocytes by Curcumin and Selenium L-Methionine. *Iran J Med Sci*. 2018;**43**(6):645-52. [PubMed: [30510341](https://pubmed.ncbi.nlm.nih.gov/30510341/)]. [PubMed Central: [PMC6230935](https://pubmed.ncbi.nlm.nih.gov/PMC6230935/)].