Published online: 2024 March 2.

Research Article



Antibacterial Agent of *Echinochloa esculenta*-Mediated ZnO Nanoparticles on Urinary Infection-Causing Bacteria

Moses Stella Bharathy^{1,*}, Gnana Sekar Dayana Jeyaleela², Joseph Devaraj Rosaline Vimala¹, Aranganathan Agila¹, Arockiya raj Divya¹

¹ PG and Department of Chemistry, Holy Cross College (Autonomous), Affiliated to Bharathidasan University, Tiruchirappalli, Tamilnadu, India
² PG and Research Department of Chemistry, Joseph's College (Autonomous), Affiliated to Bharathidasan University, Tiruchirappalli, Tamilnadu, India

*Corresponding author: PG and Department of Chemistry, Holy Cross College (Autonomous), Affiliated to Bharathidasan University, Tiruchirappalli, Tamilnadu, India. Email: stellabharathy@hcctrichy.ac.in

Received 2024 February 19; Accepted 2024 March 2.

Abstract

Background:*Echinochloa esculenta* is the Indian Barnyard millet which is a grass type of species. This study focused on the evaluation of the antibacterial activity of *Echinochloa esculenta*-mediated zinc oxide nanoparticles (ZnONPs) against urinary tract infection (UTI) microbes, such as *Escherichia coli* (*E. coli*), *Enterobacter aerogenes*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*, and *Proteus vulgaris*.

Methods: The millet *Echinochloa esculenta*-mediated ZnONPs are synthesized using a green method. Synthesized ZnONPs are characterized by ultraviolet (UV), Fourier transform-infrared resonance spectroscopy (FTIR), X-ray diffraction (XRD), scanning electron microscopy (SEM), and energy dispersive X-ray analysis (EDX) to determine the functional groups, crystalline size, shape, and elemental composition of the synthesized NPs.

Results: The UV and FTIR results of *Echinochloa esculenta*-mediated ZnONPs preliminarily confirmed the formation of ZnONPs. The XRD and SEM results also confirmed that the synthesized ZnONPs are well-crystalline in nature, and the average size was observed to be 23.38 nm with a spherical shape. *Echinochloa esculenta*-mediated ZnONPs contained 73.33% of zinc and 26.77% of oxygen, which was examined by EDX. Synthesized ZnONPs were tested against the five UTI pathogens, and they exhibited a greater zone of inhibitions than zinc acetate and plant extract. Especially against *S. aureus, Echinochloa esculenta*-mediated ZnONPs performed well with an inhibitory effect.

Conclusions: Therefore, this study had the potential value of developing new eco-friendly, low-cost, and less toxic nanomaterials that can be used as a bio-control for diseases.

Keywords: Echinochloa esculenta,Zinc Acetate Dihydrate, Synthesis and Characterizations, Anti-bacterial Effect, Urinary Tract Infection Microbes

1. Background

Urinary tract infections (UTIs) are the most common and severe public health problem. They are caused by bacterial infections and affect around 150 million individuals worldwide every year. Urinary tract infections are considered a significant cause of morbidity in males and females of all ages, even in infants. These infections are caused by both grampositive bacteria and gram-negative and by certain fungi. The most common causative agent for both uncomplicated and complicated UTIs is uropathogenic *Escherichia coli* (UPEC). Other bacteria causing UTIs are Klebsiella pneumoniae, Proteus mirabilis, Enterococcus faecalis, and Staphylococcus saprophyticus.

Metal oxide nanomaterials of CuO, Fe_3O_4 , TiO_2 , and ZnO have been scrutinized for their biological activities. Among all the metal oxides, ZnO is widely used in the elimination of heavy metals (toxic elements), such as sulfur and arsenic, from the polluted water owing to their large surface area by the volume ratio than the bulk materials (1-5). Biosynthesis or green of nanoparticles (NPs) is an alternative novel approach to the synthesis of NPs using plant extracts, algae, fungi, yeast, and bacteria (6, 7). Specifically, the plant extracts mediated-NP preparation approach is a more effective,

Copyright © 2023, Stella Bharathy et al. This open-access article is available under the Creative Commons Attribution 4.0 (CC BY 4.0) International License (https://creativecommons.org/licenses/by/4.0/), which allows for unrestricted use, distribution, and reproduction in any medium, provided that the original work is properly cited.

biocompatible, safe, and low-cost method than other chemical and physical methods (8). The literature evidence shows that plant extracts were successfully used as reducing and stabilizing agents in the synthesis of NPs, such as nickel, iron, copper, gold, silver, cobalt, lead, titanium, palladium, magnetite, platinum, and zinc (9, 10). Plant extract-mediated nanomaterials revealed a potential for pharmacological effects on microbes, cancer, malaria, wound healing, hepatitis, inflammation, and other acute diseases. Zinc oxide nanoparticles (ZnONPs) also have the potential application in the pharmacological (medicine) field, including drug delivery, diagnosis of diseases, and effects, such as antioxidant and biological antimicrobial, which can also be applied in UTI treatments. The high ionic nanosized metal oxides, such as ZnONPs, were unique because they were synthesized with an unusual crystal structure, along with the large surface areas. Moreover, the mineral element zinc is very essential to human health, and the oxide form of zinc, such as ZnO, is the daily supplement for zinc.

Zinc oxide nanoparticles exhibit good biocompatibility to the human cells. In addition, such NPs reveal cell bactericidal effects and internalization effects against the numerous microbial strains that are involved in UTIs. Due to these advantages, several researchers were interested in using zinc-based NPs for UTI therapy.

Echinochloa esculenta (Japanese Barnyard millet) and Echinochloa frumentacea (Indian Barnyard millet) are the two main species of Barnvard millet belonging to the family Poaceae (11-15). Tamil Nadu, Bihar, Punjab, Gujarat, Orissa, Maharashtra, and Madhva Pradesh are the main cultivators of Barnyard millet in India. Barnyard millet has micronutrients, such as iron and zinc, and contains an enormous amount of macronutrients, alkaloids, steroids, glycosides, and tannins, which contribute to the different medicinal properties, such as antianti-inflammatory, carcinogenic, antioxidant, antimicrobial, and wound healing capacity (16, 17). The present study aimed to biosynthesize and characterize the Echinochloa esculenta-mediated ZnONPs and their antibacterial potential against UTI-causing microbes.

2. Methods

2.1. Preparation of Precursor and Echinochloa esculenta Extract

Zinc acetate dihydrate (Zn ($C_2H_3O_2$)₂.2H₂O) and ethanol (C_2H_5OH) were of analytical grades purchased from Merck. The fresh *Echinochloa esculenta* millets were purchased from the local herbal store Aarogya Herbals, Trichy, Tamilnadu, India. Collected materials were washed properly and shade-dried. Moreover, 500 g of dried *Echinochloa esculenta* millets were taken in a 500 ml beaker, and 70% ethanol solvent was added, mixed well, and kept aside for at least 3 to 5 days. After the extraction process was completed, the extract was filtered, and the filtrate was stored in a refrigerator for future study (18-20).

2.2. Synthesis of Echinochloa esculenta-Mediated ZnONPs

In this study, 50 mL of 0.1 M zinc acetate solution was taken in a 250 ml beaker and placed on a magnetic stirrer at 60°C. Additionally, 10 ml of *Echinochloa esculenta* extract was slowly added at every 30-minute interval. After the reaction time was over, the final reaction mixture (colloidal liquid) was stood at room temperature for 12 hours. Zinc oxide nanoparticles were obtained by centrifuging at 6 000 rpm for 15 minutes. The precipitate was dried in an oven for the complete conversion of zinc hydroxide (Zn (OH)₂) into ZnO. Finally, the purified ZnONPs were further taken for characterization and microbial activity.

2.3. Characterization of Echinochloa esculenta-Mediated ZnONPs

PerkinElmer makes an ultraviolet-visible (UV-visible) spectrometer in the model of lambda 35 with a resolution and wavelength of 1 cm and 200-800 nm, respectively. The dried sample's Fourier transforminfrared resonance spectroscopy (FTIR) spectrum was documented in the range between 450 and 4000 cm⁻¹ with a resolution of 4 cm^{-1.} The crystallographic structure of a material was determined by X-ray diffraction (XRD) (Shimadzu), and the morphology of the ZnONPs was examined by scanning electron microscopy (SEM) (JEOL JSM- 6480 LV SEM). The elemental analysis of bio-synthesized ZnONPs was analyzed by EDX (Bruker) (21-24).

To examine the antibacterial activity of *Echinochloa esculenta*-mediated ZnONPs, five strains (*Escherichia coli* [MTCC 25922], *Enterobacter aerogenes* [MTCC 29212]. *Pseudomonas aeruginosa* [MTCC 27853], *Staphylococcus aureus* [MTCC 25923], and *Proteus vulgaris* (MTCC 7299]) were prepared as test organisms. The strains were procured from the microbial type culture and collection (MTCC) at Chandigarh, India. Bacterial strains were cultivated at 37°C and maintained on nutrient agar (Difco, USA) slant at 4°C. The antibacterial potential of synthesized ZnONPs was investigated against the UTI-

causing above-mentioned bacteria by the disc diffusion method (25-28). The anti-bacterial data of the zone of inhibition of extract, zinc acetate, standard, and ZnONPs were analyzed using MS Excel 2010 and presented as mean \pm standard deviation (SD) of three diameters of each zone.

3. Results and Discussion

The color changes after adding the plant extract in the reaction mixture preliminarily confirmed the formation of ZnONPs. The color change from yellow to white indicated the formation of ZnONPs. The UV spectra of *Echinochloa esculenta* extracts and qualitative analysis revealed that flavonoid aromatics (nucleus) are mostly present in it, which is shifted to 324 nm in the UV spectrum of synthesized ZnONPs. Flavonoid biomolecules in the extract played a role in the conversion of Zn²⁺ to Zn⁰ (Figure 1) (29-31).



Figure 1. UV-visible and FTIR spectrum of Echinochloa esculenta extract and Echinochloa esculenta-mediated ZnONPs

The FTIR spectrum of *Echinochloa esculenta* extracts and synthesized ZnONPs were shown in Figure 1, and inplant extract vibrational peaks were observed at 3399, 2948, 1651, 1049, 1032, 1021, 879, and 678 cm⁻¹ which denoted the existence of O-H stretching of alcohol, C-H stretching of alkane, C=C stretching of alkene, C-O stretching of alcohol, CO-O-CO stretching of anhydride, C-N stretching of amine, C=C bending of alkene, and C-X stretching of halo compound, respectively. Similarly, in the FTIR spectrum of ZnONPs, the peaks were observed at 3431, 2925,1592,1405,1341, 1119, 1049,679, and 497 cm⁻¹ which was associated at O-H Stretching of alcohol, C-H Stretching of alkane, N-H bending of amine, O-H bending of alcohol, O-H bending of phenol, C-O stretching of aliphatic ether or secondary alcohol, CO-O-CO stretching of anhydride, C=C bending of alkene, and M-O Stretching of Zn-O. The *Echinochloa esculenta* extract showed a broad peak at 3431 cm⁻¹, which indicates the presence of O-H groups, and after the synthesis of ZnONPs, there is a shift in the broad peak to the 3399 cm⁻¹, which indicates that the flavonoids or phenolic compounds O-H group involved the reducing process. The peak at 1119 cm⁻¹ is shifted to 1019 cm⁻¹ in synthesized ZnONPs, indicating that the C=O functional group might get involved in the reduction process (32).

The resulting XRD pattern of Echinochloa esculentamediated ZnONPs showed main peaks corresponding to 2θ values of 31.86⁰, 34.48⁰, 36.35⁰, 47.61⁰, 56.68⁰, 62.92⁰, and 69.09⁰ as shown in Figure 2. The obtained XRD patterns of Echinochloa esculenta-mediated ZnONPs are well correlated with the standard patterns of ZnO, which means that all the peaks match the standard JCPDS card No. 36-145. Furthermore, it confirmed that formed ZnONPs are in hexagonal wurtzite pattern/structure. The average crystallite size of the observed ZnONPs calculated using the Debye Scherrer formula was about 23.38 nm (33).





The SEM image of ZnONPs synthesized by *Echinochloa esculenta* showed relatively polydispersed shape (mostly spherical) NPs (Figure 3). The shapes and sizes of NPs play a crucial role in the effectiveness of microbial activity against any microbes/pathogens. Because spherical nanoparticles tend to be very potent during antibacterial activity owing to their ability to easily penetrate into the cell wall of pathogens, ZnONPs synthesized from the *Echinochloa esculenta* millet can be of great importance in treating clinical pathogens.

Energy dispersive X-ray spectroscopy (EDX) was used to find the elemental composition in the formed ZnONPs. The EDX of ZnONPs revealed the presence of pure zinc at 73.23% and oxygen at 26.77%, as shown in Figure 4, which evidenced that formed nanoparticles were obtained with purity (less impurity).



Figure 3. SEM image of Echinochloa esculenta-mediated ZnONPs



The antibacterial activity of synthesized ZnONPs was investigated against UTI microbes, such as *E. coli*, *E.* aerogenes, P. aeruginosa, S. aureus, and P. vulgaris, with respect to amoxicillin. Synthesized ZnONPs were tested against the five UTI-causing pathogens, and their results revealed a noticeable zone of inhibition against the pathogens compared to zinc acetate (precursor) and the *Echinochloa esculenta* extract. The maximum inhibition effect of ZnONPs was observed against the pathogen S. *aureus* (gram-positive); however, no zero inhibitory effect was observed against *P. vulgaris* (gram-negative), which is shown in Table 1. Similarly, Souza et al. (34), Azizi-Lalabadi et al. (35), Demissie et al. (36), and Farzana R et al. (37) synthesized the ZnONPs by using plant extracts and reported its anti-microbial activities against the gram-positive and negative pathogens.

Microorganisms	Zone of Inhibition (mm) (10 μ L)			
	A (Amoxicillin)	B(Zinc Acetate)	C (Plant Extract)	D (ZnONPs
Escherichia coli	8.0 ± 0.33	0	1.0 ± 0.13	2.0 ± 0.16
Enterobacter aerogenes	8.0 ± 0.62	0	3.0 ± 0.20	4.0±0.20
Pseudomonas aeruginosa	7.0 ± 0.23	0	4.0 ± 0.15	3.0 ± 0.13
Staphylococcus aureus	8.0 ± 0.30	0	0	5.0 ± 0.17
Proteus vulgaris	8.0 ± 0.66	0	2.0 ± 0.26	0

Abbreviation: ZnONPs, zinc oxide nanoparticles.

According to the results of the above-mentioned studies, it is observed that zinc oxide nanomaterial reported noticeable inhibitory actions with the zone of inhibition of 12 mm to 25 mm. A different mechanism was observed in the action of NPs against the bacteria, and their reports state that the differences in the structural/composition of NPs can affect their inhibitory potential. The zone of inhibition of *Echinochloa esculenta*-mediated ZnONPs was much lower than that of amoxicillin (standard antibiotics), which indicates the need for further modification of NPs to obtain the desired biological effects (Figure 5). The *Echinochloa esculenta*-mediated ZnONPs inhibit the microbial growth of five bacteria in the in-vitro antimicrobial assay (38, 39).



Figure 5. Antibacterial activity of Echinochloa esculenta extract, zinc acetate, ZnONPs, and standard amoxicillin on urinary tract infection pathogens

3.1. Conclusions

The synthesis of ZnONPs from *Echinochloa esculenta* through an eco-friendly approach was observed to be successful, and the UV-vis peak at 324 nm confirmed the synthesized particles were ZnONPs. The FTIR spectral data confirmed the presence of functionalities of phytochemicals, such as flavonoids and phenolics, which played a role in the reduction of zinc. The antibacterial activity of ZnONPs against urinary infection pathogens has proved that it can be used as a potent antibacterial agent. Therefore, this study had the potential value of developing new eco-friendly, low-cost, and less toxic nanomaterials that can be used as a biocontrol for diseases.

Acknowledgements

The authors would like to express their gratitude to CECRI Karaikudi for technical assistance. The authors would like to express their gratitude to the Keba Scientific Research Center and Holy Cross College (Autonomous) Trichy for permitting them to perform this research work in their laboratory.

Footnotes

Authors' Contribution: MSB and GDJ were involved in the conceptualization, methodology, writing of the original draft, and supervision. JRV, AA, and AD were involved in formal analysis, writing, review, and editing. All the authors read and approved the final manuscript. **Data Availability:** The dataset analyzed during this study is available from the corresponding author upon reasonable request.

Funding/Support: The authors declared no funding/support.

References

- 1. Kaur H, Sharma S. An overview of Barnyard millet (Echinochloa frumentacea). J Pharmacogn Phytochem. 2020;9(4):819-22.
- KUMAR PRAMOD, LAKSHMI NJ, Dube SD, Mani VP. Genotypic differences in photosynthesis and its associated parameters in relation to yield among barnyard millet (Echinochloa frumentacea), genotypes under rainfed condition in hills. *The Indian Journal of Agricultural Sciences*. 2000;**70**(4).
- Renganathan VG, Vanniarajan C, Karthikeyan A, Ramalingam J. Barnyard Millet for Food and Nutritional Security: Current Status and Future Research Direction. *Front Genet*. 2020;**11**:500. [PubMed ID: 32655612]. [PubMed Central ID: PMC7325689]. https://doi.org/10.3389/fgene.2020.00500.
- Ajaib M, Khan KM. Antimicrobial and antioxidant activities of Echinochloa colona (Linn.) link and Sporobolus coromandelianus (Retz.) kunth. *Journal of the Chemical Society of Pakistan*. 2013;**35**(3):961-6.
- Borkar VS, Kumaran KS, Kumar KL, Gangurde HH, Chordiya MA. Ethno medicinal properties of Echinochloa colona and Hydrolea zeylanica: A review. *World J Pharm Pharmc Sci.* 2016;5:354-60.
- Fakhari S, Jamzad M, Kabiri Fard H. Green synthesis of zinc oxide nanoparticles: a comparison. *Green Chemistry Letters and Reviews*. 2019;**12**(1):19-24. https://doi.org/10.1080/17518253.2018.1547925.
- 7. Vidya C, Hiremath S, Chandraprabha MN, Antonyraj ML, Gopal IV, Jain A, et al. Green synthesis of ZnO nanoparticles by Calotropis gigantea. *Int J Curr Eng Technol*. 2013;1(1):118-20.
- Naveed Ul Haq A, Nadhman A, Ullah I, Mustafa G, Yasinzai M, Khan I. Synthesis Approaches of Zinc Oxide Nanoparticles: The Dilemma of Ecotoxicity. *Journal of Nanomaterials*. 2017;2017:1-14. https://doi.org/10.1155/2017/8510342.
- Agarwal H, Venkat Kumar S, Rajeshkumar S. A review on green synthesis of zinc oxide nanoparticles – An eco-friendly approach. *Resource-Efficient Technologies*. 2017;3(4):406-13. https://doi.org/10.1016/j.reffit.2017.03.002.
- Selim YA, Azb MA, Ragab I, H. M. Abd El-Azim M. Green Synthesis of Zinc Oxide Nanoparticles Using Aqueous Extract of Deverra tortuosa and their Cytotoxic Activities. *Sci Rep.* 2020;**10**(1):3445. [PubMed ID: 32103090]. [PubMed Central ID: PMC7044426]. https://doi.org/10.1038/s41598-020-60541-1.
- Kim JY, Jang KC, Park B, Han S, Choi K, Kim S, et al. Physicochemical and antioxidative properties of selected barnyard millet (Echinochloa utilis) species in Korea. *Food Science and Biotechnology*. 2011;20(2):461-9. https://doi.org/10.1007/s10068-011-0064-z.
- 12. Moreno Amador MDL, Comino Montilla IM, Sousa Martín C. Alternative grains as potential raw material for gluten-free food development in the diet of celiac and gluten-sensitive patients. *Austin Journal of Nutrition and Metabolism*, 2 (3), 1-9. 2014.
- Nguyen DH, Zhao BT, Le DD, Yoon YH, Ko JY, Woo KS, et al. Isolation of Lignan and Fatty Acid Derivatives from the Grains of Echinochloa utilis and Their Inhibition of Lipopolysaccharide-Induced Nitric Oxide Production in RAW 264.7 Cells. J Agric Food Chem.

2016;**64**(2):425-32. [PubMed ID: 26725284]. https://doi.org/10.1021/acs.jafc.5b05638.

- Sayani R, Chatterjee A. Nutritional and biological importance of the weed Echinochloa colona: a review. Int. J. of Food Sci. Biotechnol. 2017;2:31-7.
- Gunalan S, Sivaraj R, Rajendran V. Green synthesized ZnO nanoparticles against bacterial and fungal pathogens. *Progress in Natural Science: Materials International.* 2012;22(6):693-700. https://doi.org/10.1016/j.pnsc.2012.11.015.
- Ingle A, Gade A, Pierrat S, Sonnichsen C, Rai M. Mycosynthesis of Silver Nanoparticles Using the Fungus Fusarium acuminatum and its Activity Against Some Human Pathogenic Bacteria. *Current Nanoscience.* 2008;4(2):141-4. https://doi.org/10.2174/157341308784340804.
- Anand K, Varghese S, Kurian T. Synthesis of ZnO nano rods through mechano-chemical route: a solvent free approach. *Int J Theor Appl Sci.* 2014;6(1):87.
- Colon G, Ward BC, Webster TJ. Increased osteoblast and decreased Staphylococcus epidermidis functions on nanophase ZnO and TiO2. J Biomed Mater Res A. 2006;78(3):595-604. [PubMed ID: 16752397]. https://doi.org/10.1002/jbm.a.30789.
- Arora A, Devi S, Jaswal V, Singh J, Kinger M, Gupta V. Synthesis and characterization of ZnO nanoparticles. *Oriental Journal of Chemistry*. 2014;**30**(4):1671-9. https://doi.org/10.13005/ojc/300427.
- Padmavathy N, Vijayaraghavan R. Enhanced bioactivity of ZnO nanoparticles-an antimicrobial study. Sci Technol Adv Mater. 2008;9(3):35004. [PubMed ID: 27878001]. [PubMed Central ID: PMC5099658]. https://doi.org/10.1088/1468-6996/9/3/035004.
- Sierra-Ávila R, Pérez-Alvarez M, Cadenas-Pliego G, Ávila-Orta CA, Betancourt-Galindo R, Jiménez-Regalado E, et al. Synthesis of Copper Nanoparticles Coated with Nitrogen Ligands. *Journal of* Nanomaterials. 2014;**2014**:1-8. https://doi.org/10.1155/2014/361791.
- Rajiv P, Rajeshwari S, Venckatesh R. Bio-fabrication of zinc oxide nanoparticles using leaf extract of Parthenium hysterophorus L. and its size-dependent antifungal activity against plant fungal pathogens. Spectrochim Acta A Mol Biomol Spectrosc. 2013;112:384-7. [PubMed ID: 23686093]. https://doi.org/10.1016/j.saa.2013.04.072.
- 23. Venkataraju JL, Sharath R, Chandraprabha MN, Neelufar E, Hazra A, Patra M. Synthesis, characterization and evaluation of antimicrobial activity of zinc oxide nanoparticles. *Journal of Biochemical Technology*. 2014;**3**(5):151-4.
- Zhang DH, Xue ZY, Wang QP. The mechanisms of blue emission from ZnO films deposited on glass substrate by r.f. magnetron sputtering. *Journal of Physics D: Applied Physics*. 2002;**35**(21):2837-40. https://doi.org/10.1088/0022-3727/35/21/321.
- Geetha A, Sakthivel R, Mallika J, Kannusamy R, Rajendran R. Green Synthesis of Antibacterial Zinc Oxide Nanoparticles Using Biopolymer Azadirachta indica Gum. Oriental Journal of Chemistry. 2016;32(2):955-63. https://doi.org/10.13005/0jc/320222.
- Zhou J, Zhao F, Wang Y, Zhang Y, Yang L. Size-controlled synthesis of ZnO nanoparticles and their photoluminescence properties. *Journal* of *Luminescence*. 2007;**122-123**:195-7. https://doi.org/10.1016/j.jlumin.2006.01.089.
- 27. Khoshhesab ZM, Sarfaraz M, Asadabad MA. Preparation of ZnO Nanostructures by Chemical Precipitation Method. Synthesis and

Reactivity in Inorganic, Metal-Organic, and Nano-Metal Chemistry. 2011;41(7):814-9. https://doi.org/10.1080/15533174.2011.591308.

- Wiley B, Sun Y, Mayers B, Xia Y. Shape-controlled synthesis of metal nanostructures: the case of silver. *Chemistry*. 2005;11(2):454-63. [PubMed ID:15565727]. https://doi.org/10.1002/chem.200400927.
- S G, Belay A, Reddy Ar C, Z B. Synthesis and Characterizations of Zinc Oxide Nanoparticles for Antibacterial Applications. *Journal of Nanomedicine & Nanotechnology*. 2017;**s8**. https://doi.org/10.4172/2157-7439.S8-004.
- Reddy KM, Feris K, Bell J, Wingett DG, Hanley C, Punnoose A. Selective toxicity of zinc oxide nanoparticles to prokaryotic and eukaryotic systems. *Appl Phys Lett.* 2007;**90**(213902):2139021-3. [PubMed ID: 18160973]. [PubMed Central ID: PMC2153488]. https://doi.org/10.1063/1.2742324.
- Tayel AA, El-Tras WF, Moussa S, El-Baz AF, Mahrous H, Salem MF, et al. Antibacterial Action of Zinc Oxide Nanoparticles against Foodborne Pathogens. *Journal of Food Safety*. 2011;31(2):211-8. https://doi.org/10.1111/j.1745-4565.2010.00287.x.
- Dutta RK, Nenavathu BP, Gangishetty MK, Reddy AV. Studies on antibacterial activity of ZnO nanoparticles by ROS induced lipid peroxidation. *Colloids Surf B Biointerfaces*. 2012;94:143-50. [PubMed ID: 22348987]. https://doi.org/10.1016/j.colsurfb.2012.01.046.
- Akhtar MJ, Ahamed M, Kumar S, Khan MM, Ahmad J, Alrokayan SA. Zinc oxide nanoparticles selectively induce apoptosis in human cancer cells through reactive oxygen species. *Int J Nanomedicine*. 2012;7:845-57. [PubMed ID: 22393286]. [PubMed Central ID: PMC3289443]. https://doi.org/10.2147/IJN.S29129.
- Souza RC, Haberbeck LU, Riella HG, Ribeiro DH, Carciofi BA. Antibacterial Activity of Zinc Oxide Nanoparticles Synthesized by Solochemical Process. *Brazilian Journal of Chemical Engineering*. 2019;**36**(2):885-93. https://doi.org/10.1590/0104-6632.20190362s20180027.
- Azizi-Lalabadi M, Ehsani A, Divband B, Alizadeh-Sani M. Antimicrobial activity of Titanium dioxide and Zinc oxide nanoparticles supported in 4A zeolite and evaluation the morphological characteristic. *Sci Rep.* 2019;9(1):17439. [PubMed ID: 31767932]. [PubMed Central ID: PMC6877518]. https://doi.org/10.1038/s41598-019-54025-0.
- 36. Demissie MG, Sabir FK, Edossa GD, Gonfa BA, Gawdzik B. Synthesis of Zinc Oxide Nanoparticles Using Leaf Extract of Lippia adoensis (Koseret) and Evaluation of Its Antibacterial Activity. *Journal of Chemistry*. 2020;**2020**:1-9. https://doi.org/10.1155/2020/7459042.
- 37. Farzana R, Iqra P, Shafaq F, Sumaira S, Zakia K, Hunaiza T, et al. Antimicrobial behavior of zinc oxide nanoparticles and β -lactam antibiotics against pathogenic bacteria. *Arch. Clin. Microbiol.* 2017;**8**:57.
- Chandra H, Patel D, Kumari P, Jangwan JS, Yadav S. Phyto-mediated synthesis of zinc oxide nanoparticles of Berberis aristata: Characterization, antioxidant activity and antibacterial activity with special reference to urinary tract pathogens. *Mater Sci Eng C Mater Biol Appl.* 2019;102:212-20. [PubMed ID: 31146992]. https://doi.org/10.1016/j.msec.2019.04.035.
- Chandra H, Kumari P, Bontempi E, Yadav S. Medicinal plants: Treasure trove for green synthesis of metallic nanoparticles and their biomedical applications. *Biocatalysis and Agricultural Biotechnology*. 2020;24. https://doi.org/10.1016/j.bcab.2020.101518.