Published online 2019 December 21.

Research Article

Association Study of Tumor Necrosis Factor Receptor Type II Polymorphism (196R) with Rheumatoid Arthritis in Iranian People

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Received 2018 December 22; Revised 2019 October 16; Accepted 2019 November 14.

Abstract

Background: Rheumatoid arthritis (RA) is a common autoimmune disease characterized by inflammation. It mainly affects joints and nearby tissues. Tumor necrosis factor (TNF) is the main role-player in the pathogenesis of RA. It binds to both TNF receptors namely TNF-RI and TNF-RII. Several studies have indicated a relationship between the TNF-RII 196R/R genotype with increased production of cytokines and susceptibility to inflammatory diseases and autoimmune disorders.

Objectives: Therefore, the present study aimed to investigate the association between TNF-RII polymorphism and RA. It also aimed to study the predictive value of the TNF-RII 196R allele for patients' susceptibility to RA.

Methods: A total of 100 patients and 100 controls were enrolled in the study and their peripheral blood DNA was extracted. Allelic discrimination was performed between case and control groups to investigate the association between the functional TNF-RII 196R polymorphism and RA. Screening analysis for genotyping of TNF-RII functional polymorphism was performed by the TaqMan allelic discrimination assay using real-time PCR.

Results: There was a significant difference between cases and controls in terms of the distribution of TNF-RII genotypes (TT, TG, GG). Correlation analysis showed an association between the G mutant allele and functional 196R polymorphism (CI = 95%, OR = 2.6, P = 0.035).

Conclusions: According to our results, there is a significant correlation between TNF-RII polymorphism and the risk of RA.

Keywords: TNFR II, Polymorphism, Rheumatoid Arthritis

1. Background

Rheumatoid arthritis (RA) is one of the most common autoimmune diseases. So far, no factor has been found to cause this disease. The prevalence of RA is about 1% in the human population worldwide; moreover, it is twice as prevalent in females than in males (1). A 10-year follow-up study from 2004 to 2014 in the United States showed that the prevalence of RA is increasing (2).

Rheumatoid arthritis is known as a complex disorder characterized by the inflammation of the synovium (the thin lining of a joint). Rheumatoid arthritis is a chronic disease in which genetic and environmental factors contribute to decreasing the tolerance to self-antigens (3). TNF- α is an inflammatory cytokine that plays an important role in the development of RA (4, 5). TNF- α initiates

and regulates the cytokine cascade during the pathogenesis of RA. TNF- α stimulates monocytes/macrophages, fibroblasts, and endothelial cells to produce IL-l and IL-6 or chemokines (CXCL8, CCL2), which are heavily involved in the pathogenesis of RA including leukocytes infiltration and tissue destruction. Inflammation induced by IL-1 and TNF- α can lead to the increased expression of the matrix metalloproteinase, collagenase, and elastase during the pathogenesis of RA, which is responsible for cartilage degradation and bone resorption (6, 7).

TNF- α affects the target cell via two receptors, TNF-RI (TNFRSF1A) and TNF-RII (TNFRSF1B). Although the homology of the extracellular domains of the TNF- α receptor is very close, their intracellular domains are different (8). TNF binding to TNF-RI triggers apoptosis while its binding to

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TNF-RII triggers cell survival. Both TNF-RI and TNF-RII are expressed in the synovial tissue in patients with RA (7, 9). The TNF-RI and TNF-RII genes are located on chromosomes 12p13 and 1p36, respectively (10).

Genome-wide association studies have identified that 1p36 locus is associated with RA. Interestingly, TNF-RII is located within this gene area with 10 exons (11). Reports indicate that one of the most important single nucleotide polymorphisms is located in exon 6. This missense mutation results in methionine to arginine substitution (ATG \rightarrow AGG) at position 196 (196R), which is within the fourth extracellular domain of TNF-RII (12).

Based on the biological and molecular analysis, the TNF-RII gene is considered as one of the best candidates among RA-susceptibility genes. SNP analyses at 196 (rs1061622) indicate that the 196R mutant allele has a different performance than the 196M wild-type allele (13). As reported, there appears to be an association between the exon 6 polymorphism (T676G) and susceptibility to RA(14).

Genetic markers may differ among different populations. These differences may affect the TNF-RII association with RA disease in different populations, leading to contradictory results. Previous studies have reported an association between TNF-RII 196R allele polymorphisms and RA in other populations (15, 16). Therefore, we need to evaluate the genotype and allele frequency in 196 (rs1061622) the TNF-RII gene and investigate the genetic risk of this polymorphism in our population.

2. Objectives

In this study, we tried to evaluate the association between single nucleotide polymorphisms (SNPs) of the TNF-RII gene (rs 106 16 22) and the risk of susceptibility to RA in a sample of the Iranian population.

3. Methods

3.1. Participants

After confirming the protocol of the study by the Ethics Committee of the Semnan University of Medical Sciences (ethical code; 92/372650), the study was conducted in the Department of Immunology at the University. All patients entering the study signed informed consent forms. Blood samples were taken from RA patients and healthy controls and the genomic DNA of all individuals was extracted (Qiagen DNA mini kit, Basel) from peripheral blood cells.

Our case-control study included 100 RA patients. All patients were screened as RA patients according to the American College of Rheumatology criteria. Patients were selected from those who referred to Tehran Baqiyatallah Hospital for RA treatment. The control group consisted of 100 healthy individuals.

3.2. DNA Extraction

Using QIAamp[®] DNA Mini extracting kit, genomic DNA was extracted from 500 μ L of blood lymphocytes; the extracted DNA was stored at -20°C until use for genotyping.

3.3. SNP Genotyping

The real-time PCR method was used for the determination of M196R allele polymorphism in the TNF-RII gen. All PCR tests were performed in a volume of 50 μ L, containing TaqMan Universal PCR Master Mix, specific TaqMan SNP genotyping assays, life technologies, assay ID (C 8861232 20, (part number) 4351379, USA) and genomic DNA.

Thermal cycling conditions were 5 min at 95°C, followed by 40 cycles of 95°C for 15 s, and annealing and extension were performed at 60°C for one minute. After PCR, to measure the allele-specific fluorescence, the genotyping of rs061622 SNPs was performed by TaqMan allelic discrimination with a thermocycler 7900 ABI system (17).

In this study, two TaqMan probes labeled with different fluorescent dyes were used for each sample in an allelic discrimination assay. The probes labeled with fluorescent dye FAM had a perfect match with the wild-type allele. The probes were labeled with fluorescent dye VIC because it was perfectly matched with the mutated allele (Figure 1).

3.4. Statistical Analysis

We used SPSS version 22 software for statistical analyses. The *t*-test was used to compare the mean age of cases and controls. In addition, Hardy-Weinberg equilibrium (HWE) was investigated in case and control groups of RA. The chi-square test was used to compare genotype frequencies and allele and genotype distributions in patients and controls. The associations between genotypes of the TNF-RII gene and RA were assessed by computing odds ratios (ORs) and 95% confidence intervals (95% CIs) using logistic regression analysis. The significance level was set at P < 0.05.

4. Results

Our study aimed to investigate the association between the functional 196R polymorphism of TNF-RII and RA. The mean age (\pm SD) of the participants was 48.3 \pm 13.1 years, ranging from 23 to 77 years. The mean age (\pm SD) of patients was 48.1 \pm 12.6 years. Table 1 presents the characteristics of RA patients. The differences between normal and patient groups were not significant in terms of age and sex (P > 0.05, Table 1).

Table 2 presents the genotype and allele frequencies of non-synonymous polymorphism rs1061622 of the TNF-RII gene in RA patients and controls. Significant differences were observed between genotype frequencies of the two groups in terms of 196R polymorphism. Moreover, the

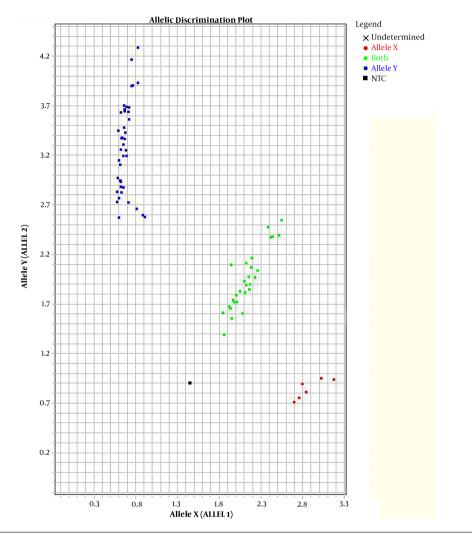


Figure 1. TaqMan[™] genotyping plot; each sample was analyzed with two probes, one specific for the wild-type and one for the mutation. The strength of fluorescence from each probe was plotted on a graph (wild-type on X-axis, mutant on Y-axis). Each sample is represented by a single point. The samples fall into three clusters representing the possible genotypes: homozygous wild-type (blue), homozygous mutant (red), heterozygous (green).

Table 1. Clinical Characteristics of the Patients ^a				
R	Values			
Male/Female (n = 100)	41/59			
Age, y	48.3 ± 13.1			
Disease duration, y	8.5 ± 5.5			
RF (IgM isotype,% of positivity)	67			
Anti-CCP, % of positivity	34			

Abbreviations: Anti-CCP, anti-cyclic citrullinated peptide antibody; RA, rheumatoid arthritis; RF, rheumatoid factor.

^aValues are expressed as mean \pm SD.

rs1061622 variant increased the risk of RA. The GG genotype was more common in RA patients than in normal subjects (OR for GG genotype: 2.6, 95% CI: 1.135 - 5.6, P = 0.035) (Table 2). Moreover, the G allele frequency in RA patients the rs1061622 was significantly higher than in the control group (OR for G phenotype: 2.30, 95% CI: 1.1 - 5.1, P = 0.04) (Table 2).

Of a total of 135 (32.5%) alleles in patients with RA, 65 cases had 196R alleles, whereas of a total of 200 (26%) alleles in healthy individuals, 52 cases had 196R alleles. In addition, the G allele increased the risk of RA.

5. Discussion

In this case-control study, we investigated the relationship between TNF-RII rs1061622 polymorphism and

Table 2. Genotype and All Genotype	lele Frequency Distribution of TN RA Patients	F-RII rs1061622 Polymorphisr Control	n in Rheumatoid Arthri OR	tis (RA) Patients and Healthy Subjects Confidence Interval	а Р
196 MM	47 (47)	53 (53)	0.9	(0.5 - 1.8)	0.74
196 MR	41 (41)	42 (42)	2.6	(1.135 - 5.6)	0.035
196RR	12 (12)	5 (5)			
Allele196M	135 (67.50)	148 (74)	2.3	(1.1 - 5.1)	0.04
196R	65	52			

Abbreviations: CI, 95% confidence interval; OR: odds ratio; RA; rheumatoid arthritis; TNF-RII, tumor necrosis factor receptor II.

^aValues are expressed as No. (%).

RA in the Iranian population. The results of the present study indicated a significant relationship between TNF-RII rs1061622 polymorphism and RA disease. There seems to be a positive correlation between TNF-RII rs1061622 polymorphism and RA.

The results showed that the TNF-RII R allele was present in 32.5% and 26% of RA patients and controls, respectively. The association between the polymorphism and the disease was confirmed by the odd ratio (CI = 95%, OR = 2.6, P = 0.035). Several studies have been conducted in different populations to investigate the relationship between TNF-RII polymorphism and RA disease. The findings of this study are in line with the results of Barton et al. studies, which demonstrated an association between the TNF-RII 196R/R genotype and familial RA in the UK and among French Caucasian populations (14, 15), and Dieude et al. study in the Japanese population (16). However, studies conducted on the Swedish population did not show any significant association between RA and the TNF-RII 196M/R polymorphism (18). It seems that heterozygous patients carrying the TNF-RII 196R allele show RA at an earlier age than earlier age than those carrying homozygous TNF-RII 196M allele. The age of the disease onset is an important factor in the study of RA. The study by Ghelani et al. on RA patients in Southeast Asia did not show any association between TNF-RII 196R/R genotype and RA disease (19).

Several studies investigating the relationship between the TNF-RII 196M/R polymorphism and RA severity have reported controversial results (19-21). van der Helm-van Mil et al. (22) and Glossop et al. (20) studies did not show any significant association between the severity of RA disease and 196R allele (19, 20). However, Goeb et al. study revealed a significant correlation between TNF-RII 196R allele and radiographic severity and diagnosis of RA (21, 23).

Previous studies have shown that the TNF-RII 196 M/R gene tends to characterize increased cytokine production and apoptosis after TNF- α stimulation (13, 24). Studies by Horiuchi et al. showed that a polymorphism in codon 196 exon 6 TNF-RII changes methionine to arginine which increases IL-6 production.in the cells carrying the 196R allele (13). Till et al. showed that the change of methionine to arginine changes the pathway for TNF-RII apoptosis that is

performed by NF-kb signaling (24).

A recent meta-analysis study performed by Song et al. (25) showed an association between the functional TNF-RII 196M/R polymorphism and RA in the European population and East Asian population. In addition, in previous studies, a significant association was found between TNF-RII 196R polymorphism and lupus disease in the Japanese population (26). As noted, this association between RA and TNF-RII polymorphism was studied in the Japanese population. A similar study in France showed the same results (15).

In conclusion, we investigated the association between *TNF-RII* gene (rs1061622) polymorphisms and RA in a sample of the Iranian population for the first time. Our results supported a significant association between the missense mutation, which involves a single base substitution at codon 196 (ATG \rightarrow AGG) in exon 6 of the *TNF-RII* gene, and susceptibility to RA.

Association studies may be limited by the heterogeneity of the population, small sample sizes, and the statistically significant differences between experimental and control groups. The difference in results could be due to different patient population genetic background in different studies. To confirm our findings, it is necessary to conduct further association studies in different ethnicities.

Footnotes

Authors' Contribution: Concept and design: Fateme Pak, Parviz Kokhaei, Mohammad Hassan Aminikhoo, and Mehdi Fasihi. Acquisition of data: Mehdi Barati and Mohammad Hassan Aminikhoo. Primary analysis and interpretation: Kazem Ahmadi. Drafting manuscript: Mohammad Hassan Aminikhoo, Mehdi Barati, and Zahra Rasouli Nejad. Supervision and critical revision: Parviz Kokhaei and Fateme Pak.

Conflict of Interests: The authors do not have any direct or indirect financial payment for the research or manuscript production by the sponsor of a product or service evaluated in this article. The authors do not own any shares in any the company or competing company sponsoring a product or service evaluated in this article. The authors hereby declare that they do not play any role as per-

sonal consultants or promoters for companies or other organizations with financial interests for the promotion of particular health care products and services.

Ethical Approval: All human interventions performed in this study were conducted following the standards of the Ethics Committee of the Semnan University of Medical Sciences and the Helsinki Declaration and its later amendments. After obtaining the consent form, we enrolled individuals in this study.

Funding/Support: This study was funded by grant no. 569 from the Semnan University of Medical Sciences.

Informed Consent: Informed consent was obtained from all individual participants. The ethical approval number was 92/372650 at the Semnan University of Medical Sciences.

References

- Hochberg MC, Spector TD. Epidemiology of rheumatoid arthritis: Update. *Epidemiol Rev.* 1990;12:247-52. doi: 10.1093/oxfordjournals.epirev.a036058. [PubMed: 2286222].
- Hunter TM, Boytsov NN, Zhang X, Schroeder K, Michaud K, Araujo AB. Prevalence of rheumatoid arthritis in the United States adult population in healthcare claims databases, 2004-2014. *Rheumatol Int.* 2017;37(9):1551-7. doi: 10.1007/s00296-017-3726-1. [PubMed: 28455559].
- 3. Turesson C, Matteson EL. Genetics of rheumatoid arthritis. *Mayo Clin Proc.* 2006;**81**(1):94–101. doi: 10.4065/81.1.94. [PubMed: 16438485].
- McInnes IB, Schett G. Cytokines in the pathogenesis of rheumatoid arthritis. Nat Rev Immunol. 2007;7(6):429–42. doi: 10.1038/nri2094. [PubMed: 17525752].
- McInnes IB, Buckley CD, Isaacs JD. Cytokines in rheumatoid arthritis - shaping the immunological landscape. Nat Rev Rheumatol. 2016;12(1):63-8. doi: 10.1038/nrrheum.2015.171. [PubMed: 26656659].
- Cornelis F, Faure S, Martinez M, Prud'homme JF, Fritz P, Dib C, et al. New susceptibility locus for rheumatoid arthritis suggested by a genome-wide linkage study. *Proc Natl Acad Sci U S A*. 1998;**95**(18):10746– 50. doi: 10.1073/pnas.95.18.10746. [PubMed: 9724775]. [PubMed Central: PMC27966].
- Tseng WY, Huang YS, Lin HH, Luo SF, McCann F, McNamee K, et al. TNFR signalling and its clinical implications. *Cytokine*. 2018;101:19–25. doi: 10.1016/j.cyto.2016.08.027.
- Brockhaus M, Schoenfeld HJ, Schlaeger EJ, Hunziker W, Lesslauer W, Loetscher H. Identification of two types of tumor necrosis factor receptors on human cell lines by monoclonal antibodies. *Proc Natl Acad Sci U S A*. 1990;87(8):3127–31. doi: 10.1073/pnas.87.8.3127. [PubMed: 2158104]. [PubMed Central: PMC53847].
- Tartaglia LA, Ayres TM, Wong GH, Goeddel DV. A novel domain within the 55 kd TNF receptor signals cell death. *Cell*. 1993;74(5):845–53. doi: 10.1016/0092-8674(93)90464-2. [PubMed: 8397073].
- Baker E, Chen LZ, Smith CA, Callen DF, Goodwin R, Sutherland GR. Chromosomal location of the human tumor necrosis factor receptor genes. *Cytogenet Cell Genet*. 1991;57(2-3):117–8. doi: 10.1159/000133127. [PubMed: 1655358].
- Shiozawa S, Hayashi S, Tsukamoto Y, Goko H, Kawasaki H, Wada T, et al. Identification of the gene loci that predispose to rheumatoid arthritis. *Int Immunol.* 1998;10(12):1891–5. doi: 10.1093/intimm/10.12.1891. [PubMed: 9885910].
- Kemper O, Derre J, Cherif D, Engelmann H, Wallach D, Berger R. The gene for the type II (p75) tumor necrosis factor receptor (TNF-RII) is localized on band 1p36.2-p36.3. *Hum Genet*. 1991;87(5):623–4. doi: 10.1007/bf00209026. [PubMed: 1655619].

- Morita C, Horiuchi T, Tsukamoto H, Hatta N, Kikuchi Y, Arinobu Y, et al. Association of tumor necrosis factor receptor type II polymorphism 196R with systemic lupus erythematosus in the Japanese: Molecular and functional analysis. *Arthritis Rheum*. 2001;44(12):2819–27. doi: 10.1002/1529-0131(200112)44:12<2819::aid-art469>3.0.co;2-2. [PubMed: 11762942].
- Barton A, John S, Ollier WE, Silman A, Worthington J. Association between rheumatoid arthritis and polymorphism of tumor necrosis factor receptor II, but not tumor necrosis factor receptor I, in Caucasians. Arthritis Rheum. 2001;44(1):61–5. doi: 10.1002/1529-0131(200101)44:1<61::AID-ANR9>3.0.CO;2-Q. [PubMed: 11212177].
- Dieude P, Petit E, Cailleau-Moindrault S, Osorio J, Pierlot C, Martinez M, et al. Association between tumor necrosis factor receptor II and familial, but not sporadic, rheumatoid arthritis: Evidence for genetic heterogeneity. *Arthritis Rheum*. 2002;**46**(8):2039–44. doi: 10.1002/art.10101. [PubMed: 12209506].
- Kyogoku C, Tsuchiya N, Shibue T, Tokunaga K, Matsuta K. TNFR2 position 196 polymorphism in Japanese patients with rheumatoid arthritis: Comment on the article by Dieude et al. *Arthritis Rheum*. 2003;48(1):273-4. doi: 10.1002/art.10600. [PubMed: 12528135].
- Pak F, Mwakigonja AR, Kokhaei P, Hosseinzadeh N, Pyakurel P, Kaaya E, et al. Kaposi's sarcoma herpesvirus load in biopsies of cutaneous and oral Kaposi's sarcoma lesions. *Eur J Cancer*. 2007;**43**(12):1877-82. doi:10.1016/j.ejca.2007.05.023. [PubMed: 17627810].
- Dahlqvist SR, Arlestig L, Sikstrom C, Linghult S. Tumor necrosis factor receptor type II (exon 6) and interleukin-6 (-174) gene polymorphisms are not associated with family history but tumor necrosis factor receptor type II is associated with hypertension in patients with rheumatoid arthritis from northern Sweden. *Arthritis Rheum.* 2002;46(11):3096–8. doi: 10.1002/art.10592. [PubMed: 12428254].
- Ghelani AM, Samanta A, Jones AC, Mastana SS. Association analysis of TNFR2, VDR, A2M, GSTT1, GSTM1, and ACE genes with rheumatoid arthritis in South Asians and Caucasians of East Midlands in the United Kingdom. *Rheumatol Int.* 2011;31(10):1355–61. doi: 10.1007/s00296-010-1478-2. [PubMed: 20401725].
- Glossop JR, Nixon NB, Dawes PT, Hassell AB, Mattey DL. No association of polymorphisms in the tumor necrosis factor receptor I and receptor II genes with disease severity in rheumatoid arthritis. *J Rheumatol.* 2003;**30**(7):1406–9. [PubMed: 12858434].
- Goeb V, Dieude P, Vittecoq O, Mejjad O, Menard JF, Thomas M, et al. Association between the TNFRII 196R allele and diagnosis of rheumatoid arthritis. *Arthritis Res Ther*. 2005;7(5):R1056-62. doi: 10.1186/ar1777. [PubMed: 16207322]. [PubMed Central: PMC1257430].
- van der Helm-van Mil AH, Dieude P, Schonkeren JJ, Cornelis F, Huizinga TW. No association between tumour necrosis factor receptor type 2 gene polymorphism and rheumatoid arthritis severity: A comparison of the extremes of phenotypes. *Rheumatology (Oxford)*. 2004;**43**(10):1232–4. doi: 10.1093/rheumatology/keh314. [PubMed: 15252214].
- Constantin A, Dieude P, Lauwers-Cances V, Jamard B, Mazieres B, Cambon-Thomsen A, et al. Tumor necrosis factor receptor II gene polymorphism and severity of rheumatoid arthritis. *Arthritis Rheum*. 2004;**50**(3):742–7. doi: 10.1002/art.20113. [PubMed: 15022314].
- 24. Till A, Rosenstiel P, Krippner-Heidenreich A, Mascheretti-Croucher S, Croucher PJ, Schafer H, et al. The Met-196 -> Arg variation of human tumor necrosis factor receptor 2 (TNFR2) affects TNF-alpha-induced apoptosis by impaired NF-kappaB signaling and target gene expression. J Biol Chem. 2005;280(7):5994–6004. doi: 10.1074/jbc.M411541200. [PubMed: 15572357].
- Song GG, Bae SC, Lee YH. Associations between functional TNFR2 196 M/R polymorphisms and susceptibility to rheumatoid arthritis: A meta-analysis. *Rheumatol Int*. 2014;**34**(11):1529–37. doi: 10.1007/s00296-014-3027-x. [PubMed: 24777778].
- Komata T, Tsuchiya N, Matsushita M, Hagiwara K, Tokunaga K. Association of tumor necrosis factor receptor 2 (TNFR2) polymorphism with susceptibility to systemic lupus erythematosus. *Tissue Antigens*. 1999;**53**(6):527–33. doi: 10.1034/j.1399-0039.1999.530602.x. [PubMed: 10395102].