



# In Vitro Scolicidal Effect of *Urtica dioica* and *Pyrus boissieriana* Extracts Against Protoscoleces of *Echinococcus granulosus*

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## Abstract

**Background:** Cystic echinococcosis (CE) is one of the most important neglected diseases and a public health concern worldwide. Due to the poor efficacy of current drugs, novel therapeutic approaches are urgently needed.

**Objectives:** This study evaluated the scolicidal effects of the hydroalcoholic extract of *Urtica dioica* and the chloroformic extract of *Pyrus boissieriana* on protoscoleces of CE cysts.

**Methods:** Protoscoleces were aseptically aspirated from the livers of sheep naturally infected with CE cysts. To assess the scolicidal effect of these herbal extracts, various concentrations of both extracts (5, 10, 20, 40, and 80 mg/mL) were added to a suspension of  $3 \times 10^3$  washed protoscoleces. After 10, 20, 30, 60, and 120 minutes of exposure, eosin stain was added to each tube, and the viability of protoscoleces was evaluated by flame cell motility under a light microscope, as well as impermeability to 0.1% eosin solution. All experiments were performed in triplicate.

**Results:** The scolicidal effects of *U. dioica* extracts at a concentration of 80 mg/mL were 81% and 89% after 60 and 120 minutes of exposure, respectively. Almost the same results were obtained for *P. boissieriana* extracts at a concentration of 80 mg/mL (81.33% and 89% after 60 and 120 minutes of exposure, respectively), which were significantly different from the negative control group ( $P < 0.001$ ). However, the extract of *U. dioica* exhibited stronger scolicidal effects compared to the extract of *P. boissieriana* at concentrations lower than 40 mg/mL ( $P < 0.001$ ).

**Conclusions:** The present findings indicate that both *U. dioica* and *P. boissieriana* extracts possess high protoscolicidal activities and could be used as alternative natural medicines in the treatment of CE. However, further studies are required to verify these findings through assessment in animal models and clinical subjects.

**Keywords:** Cystic Echinococcosis, In Vitro, *Pyrus boissieriana*, Scolicidal Activity, *Urtica dioica*

## 1. Background

Cystic echinococcosis (CE) arises from infection by the dog tapeworm, *Echinococcus granulosus*, during its larval (metacestode) stages (1). In the life cycle of *E. granulosus*, dogs and canids serve as primary definitive hosts, while ungulates (such as sheep, goats, and pigs) and humans act as intermediate hosts. The adult cestode produces infective eggs through sexual reproduction in the small intestine of a definitive host. Intermediate hosts become infected through direct

contact with a dog or indirectly via the ingestion of egg-contaminated water, soil, or vegetables (2). Following infection, the intermediate host typically develops *E. granulosus* cysts primarily in the liver (70%), lung (20 - 30%), and less frequently in the kidney, spleen, brain, heart, and other organs (3, 4). Cystic echinococcosis is classified as one of the 17 neglected tropical diseases, prevalent in various parts of the world, particularly in rural areas, and endemic in regions including South America, North and East Africa, Asia, Australia, and China (5, 6).

Currently, surgical removal of CE cysts remains the primary and preferred treatment method (7). Besides surgery, alternative treatment options for CE include chemotherapy with benzimidazoles, PAIR (puncture-aspiration-injection-reaspiration), and a watch/wait approach (8). However, during surgery, the leakage of protoscoleces-rich fluid into adjacent tissues poses a significant risk of secondary echinococcosis and recurrence (9, 10). Hence, the use of effective scolical agents is crucial to prevent such complications (11). Various scolical agents like chlorhexidine gluconate, hypertonic saline, silver nitrate, cetrimide, ethyl alcohol, and povidone-iodine are currently employed for protoscoleces inactivation during surgery (8). However, many of these agents are associated with different complications, including sclerosant cholangitis, liver necrosis, and methemoglobinemia. Therefore, the development of new scolical agents with fewer side effects, lower cost, and higher efficiency is imperative (12, 13).

Due to the reported side effects of biochemical agents and the quest for safer and more effective treatments, current medical research has increasingly focused on traditional and herbal medicine. Plant extracts are widely used in developing countries due to their availability, safety, and cost efficiency. Investigating the scolical effects of herbal extracts like *Urtica dioica* and *Pyrus boissieriana* presents a promising avenue for developing alternative treatments for CE (14).

*Urtica dioica*, commonly known as the nettle plant, is a perennial flowering plant cultivated in temperate regions and belonging to the family *Urticaceae* (15, 16). Various studies have demonstrated its diverse medicinal properties, including antioxidant, antiulcer, anticancer, antiviral, antibacterial, antifungal, and immunomodulatory activities. *Urtica dioica* has been noted for its potential in managing hypertension and hyperglycemia, with its root commonly used in the treatment of prostatic hyperplasia. Additionally, its leaf extract has been traditionally utilized to alleviate pain and inflammation, making it a folk remedy for conditions such as arthritis (17). Carvacrol, a major compound found in *U. dioica*, exhibits significant antiparasitic and antifungal properties (18).

*Pyrus boissieriana*, known as the wild pear, belongs to the *Rosaceae* family and is predominantly found in the northern regions of Iran, particularly the Hyrcanian forests (19). Studies have highlighted its notable antioxidant activities and its effectiveness in inhibiting  $\alpha$ -glucosidase, suggesting its potential as an antidiabetic agent (20). *Pyrus boissieriana* also demonstrates antihyperlipidemic and diuretic effects,

with its leaves traditionally used to address hypertension, bladder inflammation, bacteriuria, and renal stones. Arbutin, a hydroquinone derivative abundant in *P. boissieriana*, possesses antioxidant, antibacterial, and antifungal properties, along with skin whitening and UV-protective effects (21).

## 2. Objectives

Given the beneficial properties of *U. dioica* and *P. boissieriana* and the imperative to explore novel therapeutic approaches for CE, endemic in the region, we investigated the in vitro scolical effects of their extracts on protoscoleces of CE cysts.

## 3. Methods

### 3.1. Collection and Identification of the Plants

*Pyrus boissieriana* and *U. dioica* were collected during the summer of 2019 from Guilan Province, Northern Iran, and identified by professors of Pharmacognosy at the Faculty of Pharmacy of Shiraz University of Medical Sciences. Subsequently, the aerial parts of *P. boissieriana*, including flowers, and the leaves of *U. dioica* were separated and air-dried under shade.

### 3.2. Preparation of Hydroalcoholic Extract of *Urtica dioica* and Chloroformic Extract of *Pyrus boissieriana*

For the hydroalcoholic extract of *U. dioica*, the dried leaves underwent a soaking method. Initially, 150 g of dried leaves were combined with 300 ml of a hydroalcoholic solvent (70% methanol and 30% water) in an Erlenmeyer flask. Covered with aluminum foil, the flask was shaken overnight. Subsequently, the solution was filtered and refrigerated at 4°C. Using a rotary evaporator, the methanol extracts were completely evaporated. The remaining water portion was subjected to dry ice to form a thick extract, which was then stored in amber bottles at 4°C for future use.

To obtain the chloroformic extract of *P. boissieriana*, 250 g of dried plant material was powdered using an electric blender. The powder was extracted via the percolation method using a hydroalcoholic solution at a ratio of 4:1. The resulting solution was decanted with chloroform at a ratio of 1:10, with each hydroalcoholic extract washed three times with chloroform. After separation of the chloroform phase, the solution was evaporated using a rotary evaporator and dried to obtain the powder for further use (13).

### 3.3. Obtaining and Preparation of Protoscoleces

Cystic echinococcosis cyst samples were collected from slaughtered sheep at slaughterhouses under sterile conditions and promptly transported to the Parasitology Laboratory at the Department of Parasitology and Mycology, affiliated with Shiraz University of Medical Science. Cysts with a single cavity containing sufficient protoscoleces were selected. Upon arrival at the laboratory, the cyst surfaces were washed with 70% alcohol in a sterile environment. The CE fluid containing protoscoleces was aspirated using a sterile 50 mL syringe and transferred to Falcon tubes. After centrifugation for two minutes at 1500 rpm to sediment the protoscoleces, the supernatant was transferred to another container. The collected protoscoleces were washed three times with normal saline, and isolated protoscoleces were obtained from the cyst-infected livers (22).

#### 3.4. Effectiveness of *Urtica dioica* and *Pyrus boissieriana* Extracts on Protoscoleces

To assess the scolical activity of the hydroalcoholic extract of *U. dioica* and the chloroformic extract of *P. boissieriana* on protoscoleces of CE cysts, five concentrations (5, 10, 20, 40, and 80 mg/ml) of the extracts were applied for durations of 10, 20, 30, 60, and 120 minutes. Initially, a suspension of protoscoleces ( $3 \times 10^3$  washed protoscoleces) was placed in test tubes, and each concentration was added to the respective tube, gently mixed, and then incubated at 37°C for the specified durations. Subsequently, the upper phase of the mixture was carefully removed, and 0.1% eosin stain was added to the pellet of protoscoleces, which was then gently mixed. Protoscoleces sediment was smeared onto glass slides, and the viability of protoscoleces was determined (23). Sterile normal saline containing tween 20 served as the negative control group, while hypertonic saline (20%) served as the positive control group. All experiments were conducted in triplicate across 48 plates, and the mean and standard deviation (SD) were calculated for each sample.

#### 3.5. Viability Test

To assess protoscoleces viability, 0.1% eosin staining (1 gram of eosin powder in 1000 mL distilled water) was employed. Following staining, protoscoleces were evaluated for flame cell motility under a light microscope and impermeability to 0.1% eosin solution. Living protoscoleces remained colorless and transparent, whereas dead protoscoleces were stained red.

#### 3.6. Statistical Analysis

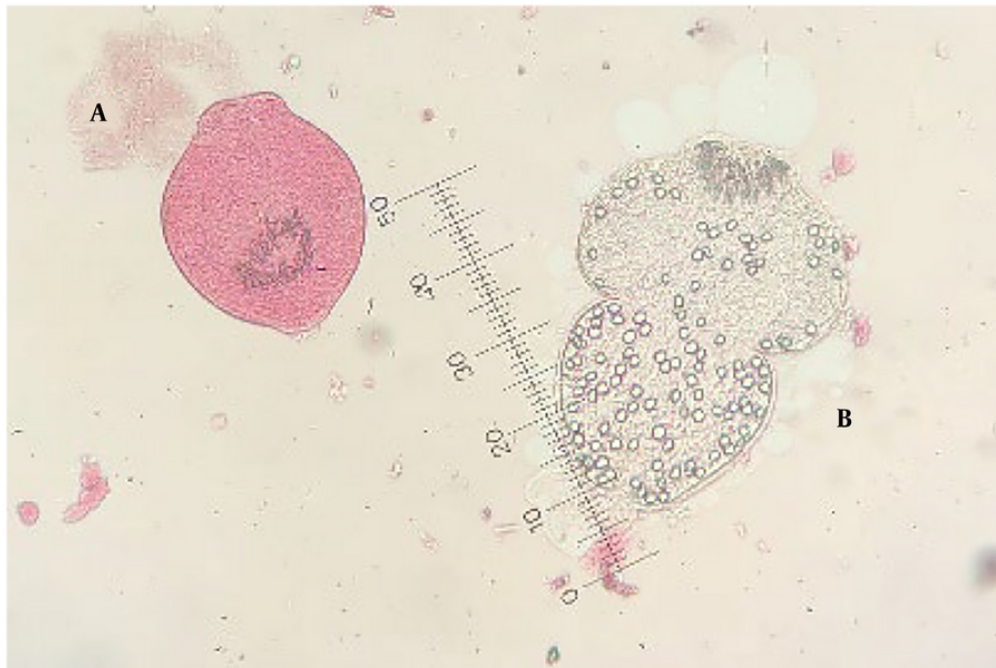
Statistical analysis was conducted using GraphPad Prism version 8.0.0 for Windows (GraphPad Software, San Diego, California, USA). Differences in scolical effects among various concentrations of *U. dioica*, *P. boissieriana*, and control groups were analyzed using one-way analysis of variance (ANOVA) followed by the Games-Howell multiple comparisons test (24). Additionally, *t*-tests were employed to assess differences in scolical effects between *U. dioica* and *P. boissieriana* at each concentration and exposure time. Results were reported as mean  $\pm$  SD, with *P*-values < 0.05 considered statistically significant.

### 4. Results

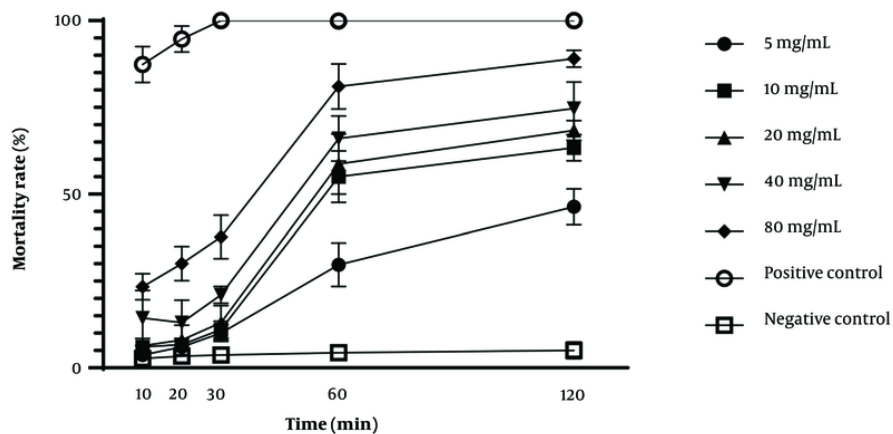
The protoscolical activity of *U. dioica* and *P. boissieriana* extracts at various concentrations following different exposure times is demonstrated in Table 1. Additionally, the scolical effectiveness of the studied extracts is illustrated in Figures 2, and 3, considering time and concentration. The scolical effects of *U. dioica* extract at a concentration of 80 mg/mL were 81% and 89% after 60 and 120 minutes of exposure, respectively. Similarly, almost identical results were obtained for *P. boissieriana* extract at a concentration of 80 mg/mL (81.33% and 89% after 60 and 120 minutes of exposure, respectively), which significantly differed from the negative control group (*P* < 0.001). However, *U. dioica* extract exhibited stronger scolical effects compared to *P. boissieriana* extract. Specifically, the scolical activity of *U. dioica* was significantly higher than that of *P. boissieriana* at concentrations lower than 40 mg/mL after 60 and 120 minutes of exposure, at a concentration of 80 mg/mL after 10 and 20 minutes of exposure, and at a concentration of 40 mg/mL after 30 minutes of exposure (*P* < 0.001). ANOVA analysis revealed that the protoscolical activity of *P. boissieriana* extract was significantly higher at concentrations of 40 and 80 mg/mL compared to lower concentrations after 30, 60, and 120 minutes of exposure (*P* < 0.001). The lowest protoscolical activity was observed in *U. dioica* at a concentration of 5 mg/mL after 10 minutes of exposure (3.67%). Our study indicates that both extracts exhibit dose-dependent and time-dependent protoscolical activity.

### 5. Discussion

The results of our study demonstrate that the extracts of *U. dioica* and *P. boissieriana* exhibit strong scolical activity, particularly at higher concentrations.



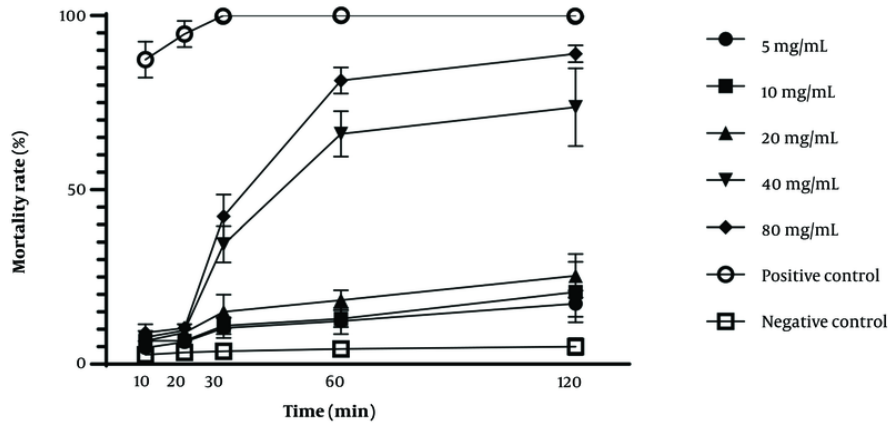
**Figure 1.** A, Viability test showing dead red-colored; and B, alive colorless protoscoleces of *Echinococcus granulosus* after staining with 0.1% eosin.



**Figure 2.** The relationship between the in vitro mortality rate of protoscoleces of *Echinococcus granulosus* and different concentrations of the hydroalcoholic extract of *Urtica dioica* following various exposure times, compared to negative and positive controls. Each point represents the mean percentage of dead protoscoleces from three different experiments.

While various studies have explored the protoscolicidal effects of hypertonic saline (25), cetrimide, ethyl alcohol (95%) (26), silver nitrate (27), 10% povidone-iodine (28),

albendazole (29), and mannitol (27), their usage is limited due to multiple dangerous complications (3). Consequently, there is an urgent need for novel



**Figure 3.** The relationship between the in vitro mortality rate of protoscoleces of *Echinococcus granulosus* and different concentrations of the chloroform extract of *Pyrus boissieriana* following various exposure times, compared to negative and positive controls. Each point represents the mean percentage of dead protoscoleces from three different experiments.

therapeutic approaches due to the inadequate therapeutic effects of existing drugs. Natural remedies and herbal products have long been utilized in treating microbial and fungal infections (30), owing to their biodegradability and safe nature for host organs. In recent decades (31), traditional and natural medicines have undergone detailed study for drug discovery purposes. Additionally, natural and herbal products serve as inspiration for developing new medications to treat various diseases (32). Although the extract of *P. boissieriana* exhibited lower potency compared to *U. dioica* at lower concentrations, both extracts at a concentration of 80 mg/mL showed significant scolicidal effects after 60 and 120 minutes. Therefore, these natural extracts have the potential to be utilized as safe medications in the treatment of CE.

Numerous studies have explored the effects of various herbal extracts on the protoscoleces of CE cysts. In 2008, Sadjjadi et al. conducted a study assessing the in vitro effects of different *Allium sativum* extracts on protoscoleces. They found that the chloroform extract of garlic at a concentration of 200 mg/mL exhibited the highest scolicidal activity, with a mortality rate of 99.97% and 100% after 60 and 120 minutes of exposure, respectively (33). Conversely, the aqueous extract of garlic at a concentration of 500 mg/mL showed the lowest scolicidal activity, with mortality rates of 29.51% and 40.6% after 60 and 120 minutes of exposure, respectively. A study conducted in 2019 investigated the protoscolicidal effects of the methanolic extract of *Allium hirtifolium* on protoscoleces of *E. granulosus*. It revealed that at a concentration of 50 mg/mL of *A.*

*hirtifolium* extract, only a 16.96% scolicidal effect was observed after 50 minutes of exposure. Despite belonging to the allium family, *A. hirtifolium*'s methanolic extract demonstrated less scolicidal effect (23). Another study by Zibaei et al. in 2012 evaluated the scolicidal effects of *Satureja khuzestanica* and *Olea europaea* against CE, showing *S. khuzestanica* to be a more effective scolicidal agent. At 0.01% concentrations, *S. khuzestanica* exhibited 83.3% and 68.6% scolicidal effects after 60 and 120 minutes of exposure, respectively (34), which aligns closely with our results. *U. dioica* and *P. boissieriana*, at a concentration of 80 mg/mL, demonstrated approximately 81% and 89% mortality rates, respectively, after 60 and 120 minutes of exposure, consistent with these findings. Moazeni and Roozitalab investigated the scolicidal effects of *Zataria multiflora* on protoscoleces and found that the extract of *Z. multiflora* is a potent protoscolicidal agent (35).

*Urtica dioica* has been utilized for centuries in treating various ailments. Previous studies have confirmed its antioxidant, antiviral, antibacterial, antifungal, anthelmintic, and analgesic properties, alongside its richness in minerals and vitamins (20, 36). Hadizadeh et al. explored the fungicidal potential of *U. dioica* against plant pathogenic fungi, establishing its efficacy as a strong fungicide (37). Mikaeili et al. investigated its antifungal effects against *Microsporum canis* both in vitro and in a guinea pig model, suggesting its topical application for treating tinea corporis (38). Sarma Katarki demonstrated the dose-dependent anthelmintic activity of the methanolic extract of *U. dioica* against adult Indian earthworms

**Table 1.** Protoscolicidal Effects of Different Concentrations of *Urtica dioica* and *Pyrus boissieriana* on  $3 \times 10^3$  Protoscoleces of Cystic Echinococcus Cyst after 10, 20, 30, 60, and 120 Minutes<sup>a, b</sup>

Exposure Time, min and Concentrations of the Extracts, mg/mL	<i>Urtica dioica</i> Extract		<i>Pyrus boissieriana</i> Extract		P-Values <sup>d</sup> of the Comparison Between <i>U. dioica</i> & <i>P. boissieriana</i> Extracts <sup>e</sup>
	Means of Mortality %	P-Value <sup>c</sup>	Means of Mortality %	P-Value <sup>c</sup>	
<b>10</b>					
5	3.67 ± 0.58 <sup>a</sup>	< 0.001	4.67 ± 0.58 <sup>a, b</sup>	< 0.001	0.1
10	6.00 ± 1.00 <sup>a, b</sup>		6.67 ± 1.16 <sup>a, c</sup>		0.49
20	6.33 ± 0.58 <sup>a, b</sup>		6.67 ± 0.58 <sup>b, c</sup>		0.51
40	14.33 ± 3.22 <sup>b, c</sup>		7.67 ± 0.58 <sup>c</sup>		0.06
80	23.33 ± 1.53 <sup>c</sup>		9.00 ± 1.00 <sup>c</sup>		< 0.001
Positive control	87.33 ± 2.08		87.33 ± 2.08		-
Negative control	2.67 ± 0.58 <sup>d</sup>		2.67 ± 0.58 <sup>a</sup>		-
<b>20</b>					
5	6.00 ± 1.00 <sup>a, e</sup>	< 0.001	6.33 ± 0.58 <sup>a</sup>	< 0.001	0.64
10	6.67 ± 0.58 <sup>a, b</sup>		6.67 ± 0.58 <sup>a</sup>		0.99
20	8.00 ± 1.73 <sup>a, b, c</sup>		9.00 ± 1.00 <sup>a, b</sup>		0.43
40	13.00 ± 2.65 <sup>a, b, c</sup>		9.67 ± 0.58 <sup>b</sup>		0.1
80	30.00 ± 2.00		10.33 ± 0.58 <sup>b</sup>		< 0.001
Positive control	94.67 ± 1.53		94.67 ± 1.53		-
Negative control	3.33 ± 0.58 <sup>a, c</sup>		3.33 ± 0.58		-
<b>30</b>					
5	10.00 ± 1.00 <sup>a</sup>	< 0.001	10.33 ± 1.16 <sup>a</sup>	< 0.001	0.72
10	11.00 ± 1.00 <sup>a</sup>		11.00 ± 1.00 <sup>a</sup>		0.99
20	13.00 ± 2.00 <sup>a</sup>		15.00 ± 2.00 <sup>a</sup>		0.28
40	21.00 ± 1.00		34.33 ± 2.08 <sup>b</sup>		0.001
80	37.67 ± 2.52		42.33 ± 2.52 <sup>b</sup>		0.08
Positive control	100.00 ± 0.00		100.00 ± 0.00		-
Negative control	3.67 ± 0.58		3.67 ± 0.58		-
<b>60</b>					
5	29.67 ± 2.52	< 0.001	12.33 ± 1.53 <sup>a</sup>	< 0.001	0.001
10	55.00 ± 3.00 <sup>a</sup>		13.00 ± 1.00 <sup>a</sup>		< 0.001
20	58.67 ± 3.51 <sup>a</sup>		18.33 ± 1.16		< 0.001
40	66.00 ± 2.65 <sup>a</sup>		66.00 ± 2.65		0.99
80	81.00 ± 2.65		81.33 ± 1.53		0.85
Positive control	100.00 ± 0.00		100.00 ± 0.00		-
Negative control	4.33 ± 0.58		4.33 ± 0.58		-
<b>120</b>					
5	46.33 ± 2.08	< 0.001	17.33 ± 1.53 <sup>a</sup>	< 0.001	< 0.001
10	63.33 ± 1.53 <sup>a</sup>		20.67 ± 3.51 <sup>a, b, c</sup>		< 0.001
20	68.33 ± 1.16 <sup>a</sup>		25.33 ± 2.52 <sup>c</sup>		< 0.001
40	74.67 ± 3.06 <sup>a</sup>		73.67 ± 4.51 <sup>d</sup>		0.76
80	89.00 ± 1.00		89.00 ± 1.00 <sup>d</sup>		0.99
Positive control	100.00 ± 0.00		100.00 ± 0.00		-
Negative control	5.00 ± 1.00		5.00 ± 1.00 <sup>b</sup>		-

<sup>a</sup> Groups with the same letters denote no statistically significant differences according to the Games-Howell multiple comparisons test. Other comparisons that lack letters or have different letters denote statistically significant differences. Statistical significance was accepted at P-value < 0.05.

<sup>b</sup> Values are expressed as mean ± SD.

<sup>c</sup> One-way analysis of variance (ANOVA)

<sup>d</sup> Independent sample t-test

<sup>e</sup> Using t-test, statistical differences between the scolical effects of *U. dioica* and *P. boissieriana* at various concentrations and exposure times were analyzed, with P-value < 0.05 considered statistically significant.

(*Pheretima posthuma*) (39). Similarly, Turel et al. examined its anthelmintic potency against mice naturally infected with *Aspicularis tetraptera*, confirming its effectiveness (40). A 2020 study validated the dose-dependent anti-leishmania effects of *U. dioica* in vitro

and in vivo (41). Additionally, Yongabi et al. evaluated its activity against multiple fungi, affirming its potent antifungal properties without significant side effects, attributed to the plant lectin agglutinin and its effective component (42), carvacrol (18). Furthermore, tannin,

another compound found in *U. dioica*, has been shown to inhibit enzymes and prevent microbial adhesion, rendering the plant effective against bacteria and fungi (43). Given the favorable outcomes of our study and other research, *U. dioica* could serve as a safe protoscolicidal agent.

Several studies have highlighted the antibacterial, antifungal, antioxidant, and antidiabetic activities of *P. boissieriana* (21). Azadbakht et al. demonstrated its antilarval, antibacterial, and antifungal properties against *Candida albicans* and *Cladosporium*. Arbutin, a significant component of *P. boissieriana*, exhibited antifungal and antioxidant effects (44). Additionally, arbutin was found to possess significant antibacterial activity, particularly in treating urinary tract infections (45). Jin and Sato investigated the antibacterial effects of aqueous extracts from young shoots of *Pyrus* spp., identifying benzoquinone, derived from arbutin, as a potent antibacterial agent (46). Güven et al. confirmed significant antifungal effects of *Pyrus serikensis* (47), while a recent study in 2019 established the antifungal effects of *Pyrus communis* against *Aspergillus flavus* and *Candida albicans*, along with antibacterial activities against *Salmonella typhimurium* (48). Considering this evidence alongside our study results, *P. boissieriana* shows promise in treating CE.

Natural remedies are widely used for their safety and minimal side effects in treating infections globally (31, 49-51). However, this study did not evaluate the cytotoxic effects of the extracts, which is a limitation. Nonetheless, other studies have assessed and confirmed the safety of these extracts. Asadi and Abbasi Maleki demonstrated the LD<sub>50</sub> of hydroalcoholic extract *U. dioica* in mice as 2900 mg/kg during acute toxicity testing, much higher than the doses used in our experiment (52). Similarly, *P. boissieriana* extract given to mice models at doses of 500 and 1000 mg/kg showed no reported side effects or mortalities (21, 53). Other *Pyrus* species, such as *Pyrus pashia* and *P. communis*, exhibited LD<sub>50</sub> of 1500 and 2000 mg/kg, respectively, in rat models (54, 55), further supporting the safety of these herbal extracts in in vivo models.

### 5.1. Conclusions

In conclusion, this in vitro study suggests that both *U. dioica* and *P. boissieriana* extracts possess significant protoscolicidal activities and could serve as alternative natural medicines for treating CE. Furthermore, these plants and their properties could inspire the development of new scolicidal agents. However, further investigations are necessary to identify and isolate the

active components of these extracts and to explore their therapeutic effects in in vivo models. Despite the absence of significant side effects in animals and humans thus far, additional studies are warranted in this regard.

### Footnotes

**Authors' Contribution:** AT and KG, conceived and designed the study; MS and EK, prepared reagents and materials; AT, KG, TZ, MG and MSB, carried out the experiments; AT and RA, analyzed and interpreted the data; AT and KG, prepared the original draft paper; AT and KG, reviewed and edited the final version of the manuscript. All authors read and approved the final manuscript.

**Conflict of Interests Statement:** The authors declared no potential conflicts of interest.

**Data Availability:** The dataset presented in the study is available on request from the corresponding author during submission or after publication.

**Ethical Approval:** This study was approved by the research ethics committee of Shiraz University of Medical Sciences with the ethical code: IR.SUMS.MED.REC.1401.030 .

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### References

- Eckert J, Deplazes P. Biological, epidemiological, and clinical aspects of echinococcosis, a zoonosis of increasing concern. *Clin Microbiol Rev.* 2004;17(1):107-35. [PubMed ID: 14726458]. [PubMed Central ID: PMC321468]. <https://doi.org/10.1128/CMR.17.1.107-135.2004>.
- Moro P, Schantz PM. Echinococcosis: a review. *Int J Infect Dis.* 2009;13(2):125-33. [PubMed ID: 18938096]. <https://doi.org/10.1016/j.ijid.2008.03.037>.
- McManus DP, Zhang W, Li J, Bartley PB. Echinococcosis. *Lancet.* 2003;362(9392):1295-304. [PubMed ID: 14575976]. [https://doi.org/10.1016/S0140-6736\(03\)14573-4](https://doi.org/10.1016/S0140-6736(03)14573-4).
- Eckert J, Gemmell MA, Meslin F, Pawlowski ZS; World Health Organization. *WHO/OIE manual on echinococcosis in humans and animals : a public health problem of global concern / edited by J. Eckert ... [et al.]*. Paris: World Organisation for Animal Health; 2001, [updated 2001]. Available from: <https://iris.who.int/handle/10665/42427>.
- Mandal S, Mandal MD. Human cystic echinococcosis: epidemiologic, zoonotic, clinical, diagnostic and therapeutic aspects. *Asian Pac J Trop Med.* 2012;5(4):253-60. [PubMed ID: 22449514]. [https://doi.org/10.1016/S1995-7645\(12\)60035-2](https://doi.org/10.1016/S1995-7645(12)60035-2).
- World Health Organization. *Working to overcome the global impact of neglected tropical diseases: first WHO report on neglected tropical diseases*. Geneva, Switzerland: World Health Organization; 2010.

- [updated 2010; cited WHO/HTM/NTD/2010.1]. Available from: <https://iris.who.int/handle/10665/44440>.
7. Tuxun T, Zhang JH, Zhao JM, Tai QW, Abudurexti M, Ma HZ, et al. World review of laparoscopic treatment of liver cystic echinococcosis-914 patients. *Int J Infect Dis*. 2014;**24**:43-50. [PubMed ID: 24747089]. <https://doi.org/10.1016/j.ijid.2014.01.012>.
  8. Velasco-Tirado V, Alonso-Sardon M, Lopez-Bernus A, Romero-Alegria A, Burguillo FJ, Muro A, et al. Medical treatment of cystic echinococcosis: systematic review and meta-analysis. *BMC Infect Dis*. 2018;**18**(1):306. [PubMed ID: 29976137]. [PubMed Central ID: PMC6034244]. <https://doi.org/10.1186/s12879-018-3201-y>.
  9. Birnbaum DJ, Hardwigsen J, Barbier L, Bouchiba N, Le Treut YP. Is hepatic resection the best treatment for hydatid cyst? *J Gastrointest Surg*. 2012;**16**(11):2086-93. [PubMed ID: 22903365]. <https://doi.org/10.1007/s11605-012-1993-4>.
  10. Jerraya H, Khalfallah M, Osman SB, Nouira R, Dziri C. Predictive factors of recurrence after surgical treatment for liver hydatid cyst. *Surg Endosc*. 2015;**29**(1):86-93. [PubMed ID: 24962861]. <https://doi.org/10.1007/s00464-014-3637-0>.
  11. Rasheed K, Zargar SA, Telwani AA. Hydatid cyst of spleen: a diagnostic challenge. *N Am J Med Sci*. 2013;**5**(1):10-20. [PubMed ID: 23378949]. [PubMed Central ID: PMC3560132]. <https://doi.org/10.4103/1947-2714.106184>.
  12. Barzin Z, Sadjjadi SM, Panjehshahin MR. Protoscolicidal Effects of the Garlic Chloroformic Extract on the Protoscolices of Hydatid Cyst at a Short Exposure Time, up to Five Minutes. *Iran J Med Sci*. 2019;**44**(1):28-34. [PubMed ID: 30666073]. [PubMed Central ID: PMC6330523].
  13. Mahmoudvand H, Fasihi Harandi M, Shakibaie M, Afatoonian MR, ZiaAli N, Makki MS, et al. Scolicidal effects of biogenic selenium nanoparticles against protoscolices of hydatid cysts. *Int J Surg*. 2014;**12**(5):399-403. [PubMed ID: 24686032]. <https://doi.org/10.1016/j.ijssu.2014.03.017>.
  14. Voon HC, Bhat R, Rusul G. Flower Extracts and Their Essential Oils as Potential Antimicrobial Agents for Food Uses and Pharmaceutical Applications. *Comprehensive Rev Food Sci Food Safety*. 2011;**11**(1):34-55. <https://doi.org/10.1111/j.1541-4337.2011.00169.x>.
  15. Fattahi S, Ardekani AM, Zabih E, Abedian Z, Mostafazadeh A, Pourbagher R, et al. Antioxidant and apoptotic effects of an aqueous extract of *Urtica dioica* on the MCF-7 human breast cancer cell line. *Asian Pac J Cancer Prev*. 2013;**14**(9):5317-23. [PubMed ID: 24175819]. <https://doi.org/10.7314/apjcp.2013.14.9.5317>.
  16. Gutowska I, Jakubczyk K, Dec K, Baranowska-Bosiacka I, Drozd A, Janda-Milczarek K, et al. Effect of the extract from nettle (*Urtica dioica* L.) fruit cluster on the synthesis of pro-inflammatory agents in hepatocytes treated with fluoride. *Fluoride*. 2014;**47**:109-18.
  17. Dhouiabi R, Affes H, Ben Salem M, Hammami S, Sahnoun Z, Zeghal KM, et al. Screening of pharmacological uses of *Urtica dioica* and others benefits. *Prog Biophys Mol Biol*. 2020;**150**:67-77. [PubMed ID: 31163183]. <https://doi.org/10.1016/j.pbiomolbio.2019.05.008>.
  18. Gul S, Demirci B, Baser KH, Akpulat HA, Aksu P. Chemical composition and in vitro cytotoxic, genotoxic effects of essential oil from *Urtica dioica* L. *Bull Environ Contam Toxicol*. 2012;**88**(5):666-71. [PubMed ID: 22310841]. <https://doi.org/10.1007/s00128-012-0535-9>.
  19. Heidari P, Rezaei M, Sahebi M, Khadivi A. Phenotypic variability of *Pyrus boissieriana* Buhse: Implications for conservation and breeding. *Scientia Horticulturae*. 2019;**247**:1-8. <https://doi.org/10.1016/j.scienta.2018.11.075>.
  20. Dehghan H, Sarrafi Y, Salehi P. Antioxidant and antidiabetic activities of 11 herbal plants from Hyrcania region, Iran. *J Food Drug Anal*. 2016;**24**(1):179-88. [PubMed ID: 28911402]. [PubMed Central ID: PMC9345419]. <https://doi.org/10.1016/j.jfda.2015.06.010>.
  21. Shahaboddin ME, Pouramir M, Moghadamnia AA, Parsian H, Lakzaei M, Mir H. *Pyrus boissieriana* Buhse leaf extract: An antioxidant, antihyperglycaemic and antihyperlipidemic agent. *Food Chem*. 2011;**126**(4):1730-3. [PubMed ID: 25213951]. <https://doi.org/10.1016/j.foodchem.2010.12.069>.
  22. Hosseini SV, Ghanbarzadeh K, Barzin J, Sadjjadi SM, Tanideh N, Mehrabani D. In vitro protoscolicidal effects of hypertonic glucose on protoscolices of hydatid cyst. *Korean J Parasitol*. 2006;**44**(3):239-42. [PubMed ID: 16969062]. [PubMed Central ID: PMC2532656]. <https://doi.org/10.3347/kjpp.2006.44.3.239>.
  23. Tabatabaei ZS, Dehshahri S, Taghi MM, Esfandiari F, Sadjjadi FS, Ebrahimipour M, et al. In Vitro Study on Protoscolicidal Effect of Methanolic Extract of *Allium hirtifolium* on Protoscolices of Cystic Echinococcosis. *Infect Disord Drug Targets*. 2019;**19**(3):264-8. [PubMed ID: 29741144]. <https://doi.org/10.2174/187526518666180509130838>.
  24. Fletcher RH, Fletcher SW, Wagner EH. *Clinical epidemiology: the essentials*. Philadelphia: Williams & Wilkins; 1996. 276 p.
  25. Kayaalp C, Balkan M, Aydin C, Ozgurtas T, Tanyuksel M, Kirimlioglu V, et al. Hypertonic saline in hydatid disease. *World J Surg*. 2001;**25**(8):975-9. [PubMed ID: 11571978]. <https://doi.org/10.1007/s00268-001-0065-9>.
  26. Besim H, Karayalcin K, Hamamci O, Gungor C, Korkmaz A. Scolicidal agents in hydatid cyst surgery. *HPB Surg*. 1998;**10**(6):347-51. [PubMed ID: 9515230]. [PubMed Central ID: PMC2423906]. <https://doi.org/10.1155/1998/78170>.
  27. Caglar R, Yuzbasioglu MF, Bulbuloglu E, Gul M, Ezberci F, Kale IT. In vitro effectiveness of different chemical agents on scolices of hydatid cyst. *J Invest Surg*. 2008;**21**(2):71-5. [PubMed ID: 18340623]. <https://doi.org/10.1080/08941930701883640>.
  28. Durgun Yetim T, Basoglu A, Taslak Sengul A, Yetim I, Serdar Bekdemir O, Hokelek M. Comparison of the protoscolicidal effectiveness of hypertonic saline, povidone-iodine and albendazole solutions in an experimental lung hydatid cyst model. *J Int Med Res*. 2011;**39**(4):1230-8. [PubMed ID: 21986125]. <https://doi.org/10.1177/147323001103900411>.
  29. Paksoy Y, Odev K, Sahin M, Arslan A, Koc O. Percutaneous treatment of liver hydatid cysts: comparison of direct injection of albendazole and hypertonic saline solution. *AJR Am J Roentgenol*. 2005;**185**(3):727-34. [PubMed ID: 16120926]. <https://doi.org/10.2214/ajr.185.3.01850727>.
  30. Harvey AL, Edrada-Ebel R, Quinn RJ. The re-emergence of natural products for drug discovery in the genomics era. *Nat Rev Drug Discov*. 2015;**14**(2):111-29. [PubMed ID: 25614221]. <https://doi.org/10.1038/nrd4510>.
  31. Lahlou M. The Success of Natural Products in Drug Discovery. *Pharmacol Amp; Pharm*. 2013;**4**(3):17-31. <https://doi.org/10.4236/pp.2013.43A003>.
  32. Yuan H, Ma Q, Ye L, Piao G. The Traditional Medicine and Modern Medicine from Natural Products. *Mol*. 2016;**21**(5). [PubMed ID: 27136524]. [PubMed Central ID: PMC6273146]. <https://doi.org/10.3390/molecules21050559>.
  33. Sadjjadi SM, Zoharizadeh MR, Panjeshahin MR. In vitro screening of different *Allium sativum* extracts on hydatid cysts protoscolices. *J Invest Surg*. 2008;**21**(6):318-22. [PubMed ID: 19160141]. <https://doi.org/10.1080/08941930802348261>.
  34. Zibaei M, Sarlak A, Delfan B, Ezatpour B, Azargoon A. Scolicidal effects of *Olea europaea* and *Satureja khuzestanica* extracts on protoscolices of hydatid cysts. *Korean J Parasitol*. 2012;**50**(1):53-6. [PubMed ID: 22451734]. [PubMed Central ID: PMC3309051]. <https://doi.org/10.3347/kjpp.2012.50.1.53>.
  35. Moazeni M, Roozitalab A. High scolicidal effect of *Zataria multiflora* on protoscolices of hydatid cyst: an in vitro study. *Comparative Clin Pathol*. 2010;**21**(1):99-104. <https://doi.org/10.1007/s00580-010-1069-3>.
  36. Kalia A, Joshi B, Mukhija M. Pharmacognostical review of *Urtica dioica* L. *Int J Green Pharm*. 2014;**8**(4):201. <https://doi.org/10.4103/0973-8258.142669>.



37. Hadizadeh I, Peivastegan B, Kolahi M. Antifungal activity of nettle (*Urtica dioica* L.), colocynth (*Citrullus colocynthis* L. Schrad), oleander (*Nerium oleander* L.) and konar (*Ziziphus spina-christi* L.) extracts on plants pathogenic fungi. *Pak J Biol Sci.* 2009;**12**(1):58-63. [PubMed ID: 19579919]. <https://doi.org/10.3923/pjbs.2009.58.63>.
38. Mikaeili A, Karimi I, Modaresi M, Bagherinasab Z. Assessment of Antidermatophytic Activities of *Urtica dioica* L against *Microsporum canis* in a Guinea Pig Model. *Tropical J Pharmaceutical Res.* 2014;**12**(6):997. <https://doi.org/10.4314/tjpr.v12i6.19>.
39. Sarma Katak M. Antioxidant, Hepatoprotective, and Anthelmintic Activities of Methanol Extract of *Urtica dioica* L. Leaves. *Pharmaceutical Crops.* 2012;**3**(1):38-46. <https://doi.org/10.2174/2210290601203010038>.
40. Turel I, Oto G, Ayaz E, Yılmaz O, Mercan U. Anthelmintic activity of *Urtica dioica* L. in mice naturally infected with *Aspicularis tetraptera*. *J Anim Vet Adv.* 2008;**7**(12).
41. Badirzadeh A, Heidari-Kharaji M, Fallah-Omrani V, Dabiri H, Araghi A, Salimi Chirani A. Antileishmanial activity of *Urtica dioica* extract against zoonotic cutaneous leishmaniasis. *PLoS Negl Trop Dis.* 2020;**14**(1). e0007843. [PubMed ID: 31929528]. [PubMed Central ID: PMC6957141]. <https://doi.org/10.1371/journal.pntd.0007843>.
42. Yongabi KA, Dukku UH, Agho MO, Chindo IY. Studies on the Antifungal properties of *Urtica dioica* urticaceae (Stinging Nettle). *J phytomedicine Therapeutics.* 2000;**5**(1):39-43.
43. Ghaedi M, Naghiha R, Jannesar R, dehghanian N, Mirtamizdoust B, pezeshkpour V. Antibacterial and antifungal activity of flower extracts of *Urtica dioica*, *Chamaemelum nobile* and *Salvia officinalis*: Effects of Zn[OH]<sub>2</sub> nanoparticles and Hp-2-minh on their property. *J Ind Eng Chem.* 2015;**32**:353-9. <https://doi.org/10.1016/j.jiec.2015.09.007>.
44. Azadbakht M, Marston A, Hostettmann K, Ramezani M, Jahromi Moghaddam M. Biological activity of leaf extract and phenolglycoside arbutin of *Pyrus boissieriana* Buhse. *J Med Plants.* 2004;**3**(10):9-14.
45. Migas P, Krauze-Baranowska M. The significance of arbutin and its derivatives in therapy and cosmetics. *Phytochem Lett.* 2015;**13**:35-40. <https://doi.org/10.1016/j.phytol.2015.05.015>.
46. Jin S, Sato N. Benzoquinone, the substance essential for antibacterial activity in aqueous extracts from succulent young shoots of the pear *Pyrus* spp. *Phytochemistr.* 2003;**62**(1):101-7. [PubMed ID: 12475625]. [https://doi.org/10.1016/S0031-9422\(02\)00444-2](https://doi.org/10.1016/S0031-9422(02)00444-2).
47. Güven K, Yücel E, Cetintaş F. Antimicrobial Activities of Fruits of *Crataegus* and *Pyrus*. Species. *Pharmaceutical Bio.* 2008;**44**(2):79-83. <https://doi.org/10.1080/13880200600591253>.
48. El-Hawary SS, El-Tantawi ME, Kirolos FN, Hammam WE. Chemical Composition, in Vitro Cytotoxic and Antimicrobial Activities of Volatile Constituents from *Pyrus communis* L. and *Malus domestica* Borkh. Fruits cultivated in Egypt. *J Essential Oil Bearing Plants.* 2019;**21**(6):1642-51. <https://doi.org/10.1080/0972060x.2018.1553637>.
49. Teimouri A, Azami SJ, Keshavarz H, Esmaeili F, Alimi R, Mavi SA, et al. Anti-Toxoplasma activity of various molecular weights and concentrations of chitosan nanoparticles on tachyzoites of RH strain. *Int J Nanomedicine.* 2018;**13**:1341-51. [PubMed ID: 29563791]. [PubMed Central ID: PMC5849388]. <https://doi.org/10.2147/IJN.S158736>.
50. Teimouri A, Haghi AM, Nateghpour M, Farivar L, Hanifian H, Mavi SA, et al. Antimalarial efficacy of low molecular weight chitosan against *Plasmodium berghei* infection in mice. *J Vector Borne Dis.* 2016;**53**(4):312-6. [PubMed ID: 28035107].
51. Teimouri A, Jafarpour Azami S, Hashemi Hafshejani S, Ghanimatdan M, Bahreini MS, Alimi R, et al. Protoscolicidal effects of curcumin nanoemulsion against protoscoleces of *Echinococcus granulosus*. *BMC Complement Med Ther.* 2023;**23**(1):124. [PubMed ID: 37072845]. [PubMed Central ID: PMC10111725]. <https://doi.org/10.1186/s12906-023-03927-8>.
52. Asadi A, Abbasi Maleki S. The effect of hydroalcoholic extract of *Urtica dioica* on morphine withdrawal signs in male mice. *J Herbmed Pharmacol.* 2018;**7**(4):220-4. <https://doi.org/10.15171/jhp.2018.34>.
53. Gholizade N, Khanbabapoor Z, Habibnejad F, Lakzaei M, Pouramir M. [Effects of *pyrus boissieriana* buhse leaves extract on antihyperglycemic, antioxidant and antilipidoxidative in rats]. *Journal of Babol University of Medical Sciences.* 2009;**11**(4):7-12. Persian.
54. Velmurugan C, Bhargava A. Anti-diabetic and hypolipidemic activity of fruits of *Pyrus communis* L. in hyperglycemic rats. *Asian J Pharmaceutical Clin Res.* 2013;**6**(SUPPL 5):108-11.
55. Ain Q, Khan H. Pharmacological basis for sedative and hypnotic like effects of *Pyrus pashia* using in vivo experimental models. *Int J Geriatr Psychiatry.* 2019;**34**(9):1345-50. [PubMed ID: 30609127]. <https://doi.org/10.1002/gps.5059>.