

Vol. 7, No. 2, April 2006

# The Relationship between High Dose Lead Exposure and Serum

## Lipids and Lipoprotein Levels.

Hami j\*, Dashti Gh R\*\*, Nemat-bakhsh M‡, Afshar M±, Ghaffari HR∏.

 \* Instructor, Department of Anatomical Sciences, School of Medicine, Birjand University of Medical Sciences, Birjand, Iran., \*\* Assistant Professor, Department of Anatomical Sciences, School of Medicine, Isfahan University of Medical Sciences, Isfahan, Iran., ‡ Professor, Department of Physiology, School of Medicine, Isfahan University of Medical Sciences, Isfahan, Iran. and ± Associate Professor, ∏ Instructor, Department of Anatomical Sciences, School of Medicine, Birjand University of Medical Sciences, Birjand, Iran.

Correspondence: Dr. J. Hami, Ghaffari Ave., Birjand, Iran., Tel: +98 (561)444-3041, Fax: +98 (561)444-0447, Email: hami\_javad@yahoo.com

#### Abstract:

This study sought to clarify the possible associations between serum lead level and serum cholestrol and lipoprotein levels in high dose lead exposed rabbits.

Levels of serum lead, total cholesterol, high density lipoprotein (HDL) cholesterol, low density lipoprotein cholesterol(LDL), very low density lipoprotein (VLDL) cholesterol and triglyceride in 11 male rabbits who were exposed to high dose lead (547ppm lead acetate in drinking water) were compared with those in 9 unexposed subjects by SPSS package and student t test. Statistical significance was defined as P<0.05.

Mean serum lead levels were significantly elevated from  $36.24\pm10.1 \ \mu$ g/L in nonexposed group to  $60.55 \pm 8.09 \ \mu$ g/L in the exposed group(P<0.001). The exposed subjects had significantly higher mean levels of total cholesterol, HDL cholesterol, LDL cholesterol (P<0.001). In contrast, VLDL cholesterol and Triglyceride level was decreased( P=0.012). Serum lead level is positively associated with total cholesterol, HDL cholesterol, LDL cholesterol.

Key Words: Serum lead level, Serum cholesterol level, Serum triglyceride level, Rabbit.

### Introduction:

Lead is a ubiquitous environmental and industrial pollutant that has been detected in almost all phases of environmental and biological systems. This heavy use has caused local and global contamination of air, dust, and soil<sup>(1)</sup>.

Whereas, cardiovascular disease is a leading cause of disability and premature death. Extensive evidence of an association between serum lipid and lipoprotein levels and coronary artery disease has been well documented<sup>(2-5)</sup>. The positive association is continuous, with no single level of cholesterol separating those who are at risk from those who are not.

Several reports have shown that both acute and chronic lead poisoning cause impairment of heart and vessel function<sup>(6-7)</sup> and that rates of death from cerebrovascular disease are significantly increased in lead-exposed workers compared with the general population<sup>(8-10)</sup>. However, no clear data are available demonstrating a higher mortality rate from heart disease in subjects exposed to lead <sup>(11)</sup>.

An association between atherosclerosis and lead exposure is biologically plausible. Microscopic analysis of lead-intoxicated animals has indicated fatty degeneration of the myocardium and sclerotic changes in the aorta and walls of the small arteries, especially the renal, cerebral and coronary arteries<sup>(12-15)</sup>, and atrophy of elastic fibers in the aorta<sup>(16)</sup>. Thus, it has been suggested that one of the underlying mechanisms in the association between cardiovascular damage and lead exposure is the induction or acceleration of atherosclerosis<sup>(7)</sup>.

According to Wojtczak-Jaroszowa and Kubow<sup>(7)</sup>, there are at least 3

pathophysiological mechanisms whereby lead could induce atherosclerosis:

(I) inhibition of superoxide dismutase, resulting in the elevation of serum lipid peroxide<sup>(17,18)</sup>; (II) formation of atherosclerotic plagues from a single mutated cell proliferating (monoc1onal hypothesis); and (III) inhibition of the activity of cytochrome P-450 <sup>(19)</sup>, leading to an increase in serum lipids and their accumulation in vessel walls. There have been few controlled studies of the effects of lead exposure with different dose on serum lipids and lipoprotein levels in either animals (14-16,20-30) or humans (31-36). (14,20,22,24,26,28,29,32,33,36) Increases decreases<sup>(16,21,23,27,30,31)</sup> and no changes<sup>(34)</sup> in serum cholesterol levels have been reported. The aim of the present study was to elucidate the effect of lead exposure on serum lipid and lipoprotein levels.

#### Materials and Methods:

Animals: Twenty (20) male white rabbits, purchased from the animal house of Pasteur Institute of Iran (Tehran, Iran). Weighing about  $1.91\pm0.21$  Kg were used as experimental animals. Throughout the present investigation, animals were housed ingroups in cages at  $22 \pm 2$  °C with free access to pellet food and water and on a 12 h light/dark cycle. They were fed a regular rabbit chow.

The animals were daily weighted. Rabbit's weight was measured with a laboratory electronic scale, which is accurate to within 1 0 g.

**Experimental Design**: Animals were randomly assigned into two groups:

1. Normal controls (n=9) were provided with regular drinking water.

2. Lead-exposed group (n=11) were received with a drinking water containing 547ppm(547  $\mu$ g/l) lead.

**Collection of Blood Sample**: Blood samples were collected from marginal ear vein of overnight fasted rabbits before starting experiments and after 40 days of treatment. The blood samples were collected in glass centrifuge tubes, then centrifuged for 15 min at 1000 × g. Sera were separated and stored at -30 °C in deep freezer till further biochemical measurements.

**Biochemical Analysis**: For measuring lead, a diphasic ammonium phosphate matrix modifier in the Ultrex and Triton X-100 grade was added to an aliquot of blood serum. A Perkin-Elmer Model 373 atomic absorption spectrophotometer with a Perkin-Elmer HGA 2200 graphite furnace was used together with a hollow cathode lamp and a Perkin-Elmer deuterium corrector.

Laboratory Analysis: Serum triglyceride and total cholesterol were analyzed using kits obtained from Sigma Diagnostics, and serum HDL, LDL, and VLDL cholesterol were measured using the Cholestech Lipid LDX system (Hayward, CA).

**Statistical Analysis:** SPSS package and student t test was used to compare the weight, serum lead, total cholesterol, HDL cholesterol, LDL cholesterol, and VLDL

cholesterol and triglyceride betweens case and control groups. Statistical significance was defined as P<0.05.

#### **Results:**

In the recent years, many researchers have studied relationship between environmental pollutions and cardiovascular diseases<sup>(37-39)</sup>.

The animals weight in two groups is shown in Table 1. A student t test showed a nonsignificant decrease in body weight in lead exposed rabbits; in comparison to controls (p > 0.05). The differences of serum lead concentrations between two groups were significant (P < 0.001)(Table 2).

Mean cholesterol, lipoproteins and triglyceride values for both groups, before and after experiment are given in Table 2. Total cholesterol , HDL cholesterol and LDL cholesterol values were significantly higher in the high dose lead-exposed subjects (P<0.001); and VLDL and triglyceride significantly decreased (P=0.012).

Table 1. Changes of body weight in rabbits before and aft	er of experiment.
---	-------------------

Group	Mean weight before experiment	Mean weight after experiment		
Group	(Kg)	(Kg)		
Case ( lead exposed)	1.89 ± 0.19	1.82 ± 0.3		
Control (Non exposed)	1.94 ± 0.24	1.96 ± 0.31		

Average ± standard deviation

Table 2. Changes in lead, total cholesterol, LDL-cholesterol, HDL-cholesterol, VLDL and

Group	Lead (µg/l)	Total cholesterol (µg/dl)	LDL- cholesterol (µg/dl)	HDL- cholesterol (µg/dl)	VLDL cholesterol (µg/dl)	Triglyceride (µg/dl)
Case (Lead exposed)	60.55± 8.09	116.81 ± 41.62	87.74 ± 36.93	17.0 ± 4.87	12.07 ± 3.24	60.36 ± 16.22
Control (Nonexposed)	36.24±10.11	60.77± 5.73	29.84 ± 2.26	10.33 ± 0.7	20.6 ± 7.67	103.0± 38.36
Ρ	0.000	0.002	0.001	0.003	0.012	0.012

triglyceride following exposed with lead.

Average ± standard déviation

### Discussion

Lead is dispersed throughout the environment, in ambient air, in many foods, in drinking water and in dust <sup>(40)</sup>. The major environmental sources of metallic lead and its salts are paint, auto exhaust, and contaminated food and water <sup>(41-43)</sup>.

The main finding of this study is that serum cholesterol and lipoprotein levels were higher in subjects who were exposed to high dose lead than in those who were not exposed (Table 2). This relationship between serum lead and serum lipid levels in the exposed subjects, suggesting an altered lipid metabolism related to lead exposure.

The assessment of a possible relationship between serum lead level and lipids is an important step in elucidating the mechanisms underlying the excess cardiovascular morbidity among lead-exposed subjects<sup>(6)</sup>. The associations between serum lead level and serum total cholesterol , HDL cholesterol and LDL cholesterol reached statistical significance (p<0.001), but VLDL cholesterol and triglyceride levels significantly decreased in lead exposed animals (P=0.012);(Table.2.).

Our finding of elevated total cholesterol levels supports 3 previous reports<sup>(32,33,36)</sup> and disagrees with 2 others<sup>(31,34)</sup>

In the study by El Gazzar et al.<sup>(33)</sup> lipoproteins were not examined and in the others the results were inconsistent. An increase in HDL cholesterol was reported by Cocco et al.<sup>(34)</sup>, but this increase was accompanied by reduced total cholesterol. Differences between our results and those of the studies cited may be due to differences in sample subjects or to high dose of lead acetate used in this study or an absence of control for confounding variables in the other studies.

Our results are consistent with the hypothesis that a lead induced accumulation of serum lipids is one of the underlying mechanisms in the association between lead exposure and cardiovascular damage. Because of the positive and continuous association between serum cholesterol and coronary artery disease<sup>(5)</sup>, it seems that subjects who are occupationally exposed to lead are at higher risk of coronary artery disease than those who are not exposed.

The association between lead exposure, and high serum lipid levels is biologically plausible and could be due to either increased synthesis or decreased removal of lipoproteins.

Decreased removal may occur as a result of the alteration of cell surface receptors for lipoproteins<sup>(20)</sup> or as a result of the inhibition of hepatic lipoprotein lipase activity<sup>(44)</sup>. Furthermore, lead has been shown to depress the activity of cytochrome P-450<sup>(19,45-46)</sup>. this can limit the biosynthesis of bile acids, which is the only significant route for e1imination of cholesterol from the body. Increased synthesis may be due to a lead induced increase in hepatic enzymes at important control points for de novo cholesterol synthesis, as has been found in Wistar rats<sup>(47)</sup>, or it may be due to impaired feedback inhibition.

We conclude that serum lead level is positively associated with levels of serum total cholesterol, HDL cholesterol and LDL cholesterol . The positive association between serum lead level and serum cholesterol among exposed subjects may have important clinical implications.

Researchers have demonstrated the benefit of antioxidants including, vitamin E, vitamin C, vitamin B6,  $\beta$ -carotene, zinc, and selenium in preventing lead toxicity<sup>(48-52).</sup>

Further studies should focus on exploring the benefits of these antioxidants in prevention of lead Toxicity.

#### Acknowledgements:

The authors thank Mohammad Mardani, Ph.D., and Ebrahim Esfandiari Ph.D. for their excellent technical assistances.

#### **References:**

1. Centers for Disease Control and Prevention. Developmental assessment and interventions. In: Managing Elevated Blood Lead Levels Among Young Children: Recommendations from the Advisory Committee on Childhood Lead Poisoning Prevention. CDC. Atlanta; 2002: 79–95 (Chapter 5).

2. Haust MD, More RH. Development of modern theories on the pathogenesis of atherosclerosis. In: Wissler RW, Geer JC. Editors the pathogenesis of atherosclerosis. Baltimore : Williams and Wilkins; 1972: 1–19.

3. Dawber TR, Kannel WB, Revotskie N, Kagall A. The epidemiology of coronary heart disease. The Framingham inquiry. Proc R Soc Med 1962;58:551-565.

4. Stamler J. Diet, serum lipids, and coronary heart disease: the epidemiologic evidence. In: Levy RI, Ritkind BM, Dennis BH, Emst N, editors Nutrition, Lipids and Coronary Heart Disease: A Global View. New York: Raven Press; 1979.

5. Kannel WB, Castelli WP, Gordon T. Cholesterol in the prediction of atherosclerotic disease. New perspectives based on the Framingham Study. Ann Intern Med 1979;90:85-91.

6. Kopp SJ, Barron JT, Tow JP. Cardiovascular action of lead and relationship to hypertension: a review. Environ Health Perspect 1988;78: 91-99.

7. Wojtczak-Jaroszowa J, Kubow S. Carbon monoxide, carbon disulfide, lead and cadmium-four examples of

occupational toxic agents linked to cardiovascular disease. Med Hypotheses 1989;30: 141-150.

8. Dingwall-Fordyce I, Lane RE. A follow-up study of lead workers. Br J Indust Med 1963; 20:313-315.

9. Malkolm D. Prevention of long-term sequelae following the absorption of lead. Arch Environ Health 1971;23:292-298.

10. Gerhardsson M, Rozenqvist U, Ahlbom A, Carlson LA. Serum cholesterol and cancer-a retrospective case-control study. Int J Epidemiol 1986; 15: 155-159.

11. Cooper WC, Galley WR. Mortality of lead workers. J Occup Med 1975; 17:100-107.

12. Kuzminskaya GN. Effect of lead poisoning on experimental atherosclerosis. Arkh Parologii 1964;26:833-835.

13. Kuzminskaya GN. Effect of lead poisoning on experimental atherosclerosis. Fed Proc 1965;24:833.

14. Revis NW, Horton Y, Majors T. The effects of calcium, magnesium, lead or cadmium on lipoprotein metabolism and atherosclerosis in the pigeon. J Environ Pathol Toxicol1980;4: 293-303.

15. Hass GM, Brown DVL, Eisenstein R, Hemmens A. Relations between lead poisoning in rabbit and man. Am J Pathol 1964;45:691-693.

16. Skoczynska A, Smolik R, jelen M. Lipid abnormalities in rats given small doses of lead. Arch Toxicol 1993;67:200-204.

17. Ding Y, Gonick HC, Vaziri ND: Lead promotes hydroxyl radical generation and lipid peroxidation in cultured aortic endothelial cells. Am J Hypertens 2000;13: 552-555.

18. Qinlan GJ, Halliwell B, Moorhouse CP, Gutteridge JM. Action of lead (II) and aluminum (III) ions on iron stimulated lipid peroxidation in liposomes, erythrocytes and rat liver microsomal fractions. Biochim Biophys Acta 1988; 962: 196-200.

19. Alvares AP, Fischbein A, Sassa S, Anderson KE, Kappas A. Lead intoxication: effects on cytochrome P-450-mediated hepatic oxidations. Clin Pharmacol Ther 1976; 19: 183-190.

20. Tarugi P, Calandra S, Borella P. Vivoli GF. Effect oflead intoxication on rabbit plasma lipoproteins. Atherosclerosis 1982;45:221-234.

21. Schroeder HA, Balassa JJ. Innuence of chromium, cadmium and lead on rat aortic lipids and circulating cholesterol. Am J 1965;209:433-437. Physiol

22. Stofen D. Environmental lead and the heart. J Mol Cell Cardiol 1974;6:285-290.

23. Ruparelia SG, Yogendra V, Metha NS, Salyed SR. Lead-induced biochemical changes in freshwater fish Oreochromis mossombicus. Bull Environ Contam Toxicol 1989;43:310-314.

24. Ledda-Columbano GM, Columbano A, Dessi S. Hexose monophosphate shunt and cholesterogenesis in lead-induced kidney hyperplasia. Chem Biol Interact 1987;62:209-215

25. Columbano A, Ledda GM, Sirigu P, Perra T, Pani P. Liver cell proliferation induced by a single dose of lead nitrate. Am J Pathol 1983: 110: 83-88.

26. Xiao GH, Wu JL, Liu YG. The effect of cadmium, mercury and lead in vitro on hepatic microsomal mixed function oxidase and lipid peroxidation. J Tongji Med Univ 1989;9:81-85.

27. Tulasi SJ, Reddy PUM, Ramana Rao JS. Accumulation of lead and effects on total lipids and lipid derivatives in the freshwater fish Anabas testudineus. Ecotoxicol Environ Safety 1992; 23:33-38.

28. Yagminas AP, Franklin CA, Villeneuve DC, Gilman AP, Little PB, Valli VEO. Subchronic oral toxicity of triethyl lead in the male weanling rat. Clinical, biochemical, hematological and histopathological effects. Fundam Appl Toxicol 1990; 15:580-596.

29. Dessi S, Batetta B, Carrucciu A, Pulisci D,Laconi S, Fadda AM, Anchisi C, Pani P. Variations of serum lipoproteins during cell proliferation induced by lead nitrate. Exp Mol Pathol 1989;51: 97-102.

30. Khan MZ, Szarek J, Krasnodebska-Oepta A, Koncicki A. Effects of concurrent administration of lead and selenium on some haematological and biochemical parameters of broiler chickens. Acta vet Hung 1993;41:123-137.

31. Cocco PL, Cocco E, Anni MS, Flore C, Salis S. Occupational exposure to lead and blood cholesterol in glucose-6-phosphate dehydrogenase deficient and normal subjects. Res Commun Chem Pathol Pharmacol 1991;72:81-95.

32. Nomiyama K, Nomiyama H, Liu SJ, Tao YX, Nomiyama T, Omae K. Lead induced increase of blood pressure in female lead workers. Occup Environ Med 2002;59: 734–738. 33. EI-Gazzar RM, El-Hefny SA, Noweir KH, Shamy MY. Study of the lipoprotein pattern among workers exposed to lead. J Egypt Public Health Assoc 1989;64:571-585.

34. Cocco P, Salis S, Anni M, Cocco ME, Flore C, Ibba A. Effects of short-term occupational exposure to lead on erythrocyte glucose-6phosphate dehydrogenase activity and serum cholesterol. J Appl Toxicol 1995; 15:373-378.

35. Morisi G, Menditto A, Spagnolo A, Patriarca M, Menotti A. Association of selected social, environmental and constitutional factors to blood lead levels in men aged 55-75 years. Sci Total Environ 1992; 126:209-229.

36. Gatagonova TM. Characteristics of the serum lipids in workers of lead industry. Med Tr Prom Ekol 1994;12:17-21.

37. Hosseinpoor AR, Forouzanfar MH, Yunesian M, Asghari F, Holakouie Naieni K, Farhood D. Air pollution and hospitalization due to angina pectoris in Tehran, Iran. Environ Res 2005;99(1):126-131.

38. Brindley P, Wise SM, Maheswaran R, Haining RP. The effect of alternative representations of population location on the areal interpolation of air pollution exposure. Comput Environ Urb Sys 2005; 29(4): 455-469.

39. Chang CC, Tsai SS, Ho SC, Yang CY. Air pollution and hospital admissions for cardiovascular disease in Taipei, Taiwan. Environ Res 2005; 98(1): 114-119.

40. Michael MI. Effect of lead on some physiological parameters in heat stressed rats. M.Sc. Thesis, Institute of Studies and Environmental Research, Ain Shams University: 1997.

41. Shy MC. Lead in petrol: the mistake of the XX century, Rapptrimest Statist Mond 1990; 43: 168–170.

42. Royce S, Herdert L, Needleman E. Case studies in environmental medicine. Lead toxicity. ATSDR 1990: 2–8.

43. Bornschein RL, Succop P, Dietrich KN, Clark CS, Que HS, Hammond PB. The influence of social and environmental factors on dust lead, hand lead and blood lead levels in young children. Environ Res 1985; 38: 108–118.

44. Chajet-Shaul T, Friedman G, Stein O, Shiloni F, Etienne J, Stein Y. Mechanism of the hyper triglyceridemia induced by tumor necrosis factor admimstration to rats. Biochim Biophys Acta 1989;1001:316-324.

45. Alvares AP, Kapelner S, Sassa S, Kappas A. Drug metabolism in normal children, lead poisoned children and normal adults. Clin Pharmacol Ther 1975; 17: 179-183.

46. Meredith PA, Campbell BC, Goldberg A. The effect of industrial lead poisoning on cytochrome P-450 mediated phenazone hydroxylation. Eur J Clin Pharmacol 1977; 12; 235-239.

47. Dessi S, Batcna B, Laconi E, Enms C, Pani P. Hepatic cholesterol in lead nitrate induced liver hyperplasia. Chem Biol interact 1984;48: 271-279.

48. Simon JA, Hudes ES. Relationship of ascorbic acid to blood lead levels. JAMA 1999; 281 (24): 2289–2293.

49. Vij AG, Satija NK, Flora SJ. Lead induced disorders in hematopoietic and drug metabolizing enzyme system and their protection by ascorbic acid supplementation. Biomed Environ Sci 1998; 11 (1):7–14.

50. Matte TD. Reducing blood lead levels: benefits and strategies, JAMA 1999; 281 (24): 2340–2342.

51. Houston DK, Johnson MA. Does Vitamin C intake protect against lead toxicity?. Nutr Rev 2000; 58 (3 Pt 1): 73–75.

52. Hsu PC, GuoYL.Antioxidant nutrients and lead toxicity. Toxicol 2002; 180(1):33-44.

Copyright © 2005 by Department of Internal Medicine, Shiraz University of Medical Sciences, Shiraz, Iran. All rights reserved.