

# Evaluation of gender-related differences in response to oxidative stress in *Toxoplasma gondii* positive serum

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## ABSTRACT

**Purpose:** *Toxoplasma gondii* (*T. gondii*) is the causative agent of toxoplasmosis. It infects up to one third of the human population. The aim of this study was to evaluate the effects of *T. gondii* infection on induction of oxidative stress in serum of infected men and women.

**Materials and Methods:** This case-control study was carried out on 150 individuals who had referred to our center in Tehran. Serum was obtained from venous blood samples. Immunoglobulin G (IgG) anti-*Toxoplasma* antibody enzyme linked immunosorbent assay (ELISA) test was performed on all of the samples. Those who were IgG positive were regarded as the case group (52 women and 23 men) and the others as the control group (43 women and 32 men). The data were analyzed by INSTAT software using ANOVA followed by Tukey.

**Results:** Serum superoxide dismutase activity in men of the case group was significantly higher than in the control group ( $7.81 \pm 0.38$  vs.  $6.69 \pm 0.17$ ,  $P = .045$ ). Catalase activity in men of the case group was significantly higher than the control group ( $8.64 \pm 0.55$  vs.  $6.23 \pm 0.38$ ,  $P = .006$ ). Glutathione S-transferase activity and malondialdehyde level in women of the case group were significantly higher than the control group ( $5.98 \pm 0.24$  vs.  $4.73 \pm 0.28$ ,  $P = .037$  and  $2.3 \pm 0.09$  vs.  $1.9 \pm 0.09$ ,  $P = .032$ , respectively). Catalase activity and glutathione level in women of the case group were lower than the control group ( $6.0 \pm 0.45$  vs.  $7.63 \pm 0.48$ ,  $P = .043$  and  $0.62 \pm 0.05$  vs.  $0.89 \pm 0.05$ ,  $P = .007$ , respectively).

**Conclusion:** *T. gondii* infection induces oxidative stress in women's serum because of the decreased catalase activity, glutathione depletion and increasing lipid peroxidation. The increased antioxidant enzyme activities in infected men were because of the adaptive response to the generated free radicals. Women were found to be more sensitive to the effects of *Toxoplasma* infection on oxidative stress induction compared to men.

**Keywords:** *toxoplasma gondii*; antioxidant enzymes; lipid peroxidation; human; serum.

AMHSR 2014;12:64-69  
www.journals.ajaums.ac.ir

## INTRODUCTION

Toxoplasmosis is one of the common parasitic infections in tropical and subtropical climates. Its causative agent is *Toxoplasma gondii* (*T. gondii*). It exists in a chronic asymptomatic form in 500 million to 1 billion people.<sup>1,2</sup> In Iran, at least 30% of the population in most regions are infected with it.<sup>3</sup> Toxoplasmosis

occurs in cases of congenital infection and in immune-compromised individuals, particularly malignant patients under chemotherapy, organ transplant recipients and people with AIDS.<sup>4-6</sup> Toxoplasmosis can cause serious pathologies including hepatitis, pneumonia, blindness and severe neurological disorders.<sup>7</sup>

*T. gondii* is an obligated intracellular protozoan

parasite, which infects humans and most warm-blooded animals, depending on hygiene standards, eating habits, profession and living place (urban or rural).<sup>2,6,8</sup> *Toxoplasma* has a complex life cycle consisting of a sexual cycle in its feline definitive hosts and an asexual cycle in its intermediate hosts. Intermediate hosts like humans can be infected by eating raw or uncooked meat of infected animals, ingestion of oocysts shed in cat feces or from mother to fetus.<sup>4,6,9,10</sup> Acute infections in pregnant women cause severe congenital toxoplasmosis, in which the symptoms could be mental retardation, cerebral calcifications, hydrocephaly, chorioretinitis, microcephaly, convulsions or even death of the fetus.<sup>8,11</sup> Chronic *T. gondii* infection can lead to cryptogenic epilepsy, headaches, and schizophrenia.<sup>8</sup>

*T. gondii* infection is associated with activation of T helper 1 (Th1) cells secreting cytokines. Cytokine-activated macrophages release a great number of reactive oxygen species (ROS), which are responsible for parasite killing in macrophages.<sup>4,5,12-14</sup> Antioxidant defense system including glutathione, superoxide dismutase (SOD), catalase and glutathione S-transferase (GST) form a network protecting cells against ROS.<sup>15,16</sup> Several studies have shown that decrease of antioxidant enzyme activities in *T. gondii*-infected patients are associated with a depletion of glutathione and an increase of lipid peroxidation, all of which can lead to oxidative stress and finally cell death.<sup>7,17-21</sup> Previous studies have also stated the role of cytosolic antioxidant enzymes in *T. gondii* in defense against oxidative injury.<sup>22-24</sup>

Elucidating the molecular and cellular pathways activated in response to infection is crucial to understanding disease pathogenesis and to developing control strategies rationally.<sup>25,26</sup> Oxidative events against *T. gondii* infection are not well elucidated in animals. The response to infection and the ability to neutralize oxidant species differ between men and women.<sup>27,28</sup> There are few reports on the gender-dependent effects of *T. gondii* infection on induction of oxidative stress in various tissues of the host.<sup>27</sup> To our knowledge, this is the first study on these effects in Iran. Thus, the aim of this study was to evaluate the gender-dependent effects of *T. gondii* infection on important biomarkers of oxidative stress including malondialdehyde content as an important index of lipid peroxidation and antioxidant defense parameters such as glutathione level, activities of SOD, catalase and GST in serum of *T. gondii*-infected patients.

## MATERIALS AND METHODS

This case-control study was carried out on 150

individuals who had been referred to Baqiyatallah hospital in Tehran from January until July 2013 (age range: 20-50 years old). None of the participants of this study took medication or supplementation upon entering the study. Venous blood samples were obtained and immediately centrifuged at 1500×g for 10 minutes at 4°C. Serum was separated and stored in 0.5 mL aliquots at -70°C until biochemical analysis. Enzyme linked immunosorbent assay (ELISA) test was performed on all of the samples using immunoglobulin G (IgG) Kit (Pishtaz Teb Co., Tehran, Iran) and the final results were recorded by ELISA reader (optical absorbance, OD = 450). Using this kit, samples less than 0.9 unit/ml and more than 1.1 unit/mL were considered as negative and positive, respectively. Participants with IgG (anti-*Toxoplasma* antibody) positive samples were considered as the case group (52 women and 23 men) and the rest as the control group (43 women and 32 men). This study was approved by the Ethical Committee of Baqiyatallah University of Medical Sciences.

### SOD Activity Assay

The SOD activity was determined using the method described by Winterbourn, based on the ability of SOD to inhibit the reduction of nitro-blue tetrazolium by superoxide.<sup>29</sup> The absorbance of samples was read on a Genesys 10 UV spectrophotometer at 560 nm for 5 minutes. The amount of enzyme required to produce 50% inhibition was taken as 1 U and the results were expressed as U/mg protein.

### Catalase Activity Assay

Serum catalase activity was measured spectrophotometrically at 240 nm by calculating the rate of degradation of H<sub>2</sub>O<sub>2</sub> as the substrate of the enzyme using the Aebi method.<sup>30</sup> A molar absorption of 43.6 Mcm<sup>-1</sup> was used to determine catalase activity. Enzymatic activity was expressed as U/mg protein, one unit (U) of which was equal to 1 mole of H<sub>2</sub>O<sub>2</sub> degraded/min/mg of protein.

### GST Activity Assay

GST activity was assayed by monitoring the formation of the thioether product of the reaction between glutathione and 1-chloro-2, 4-dinitrobenzene (CDNB) at 340 nm.<sup>31</sup> The enzyme activity was calculated using extinction coefficient 9.6 mMcm<sup>-1</sup> and expressed as μmol CDNB utilized/min/mg protein.

### Determination of Glutathione Level

Glutathione level was measured using Tietz method.<sup>32</sup>

Glutathione in the supernatant was assayed at 412 nm by monitoring the absorbance of 5, 5'-dithiobis 2-nitrobenzoic acid for 5 minutes. Glutathione level was determined from a standard curve and expressed as nmol/mg protein.

### Determination of Malondialdehyde Level

As an indicator of lipid peroxidation, malondialdehyde serum level was determined at 532 nm using 2-thiobarbituric acid according to the method of Satoh.<sup>33</sup> Malondialdehyde concentration was determined using 1,1,3,3-tetraethoxypropane as standard and expressed as nmol/mg protein.

### Protein Level Assay

The total protein contents of the samples were measured by Bradford's method using bovine serum albumin as a standard.<sup>34</sup>

### Statistical Analysis

All calculations were performed using INSTAT statistical software. For the sex-dependent studies, the data were statistically analyzed using ANOVA followed by Tukey post hoc multiple comparison test. *P*-values less than 0.05 were considered as statistically significant. The results were expressed as mean  $\pm$  standard error of the mean (SEM), with *n* denoting the number of experiments performed.

## RESULTS

Table 1 shows the prevalence of IgG antibodies related to *T. gondii* according to the sex of control and case groups. Overall, 75 (50%) of 150 participants were

**Table 1.** The prevalence of IgG antibodies specific to *Toxoplasma gondii* according to the sex of the control and the case groups.

Sex	Seroprevalence		
	Negative <i>n</i> (%)	Positive <i>n</i> (%)	Total <i>n</i> (%)
Man	32 (21.3)	23 (15.3)	66 (36.6)
Woman	43 (28.7)	52 (34.7)	84 (63.4)
Total	75 (50.0)	75 (50.0)	150 (100.0)

**Table 2.** Effect of *Toxoplasma gondii* infection on weight, serum glutathione and malondialdehyde levels in the control and the case groups.

Parameters	Control (Mean $\pm$ SEM)		TOX (Mean $\pm$ SEM)	
	Men ( <i>n</i> = 43)	Women ( <i>n</i> = 32)	Men ( <i>n</i> = 23)	Women ( <i>n</i> = 52)
Weight (g)	36.744 $\pm$ 1.185	34.437 $\pm$ 1.382	39.565 $\pm$ 1.415	36.077 $\pm$ 0.779
GSH (nmol/mg protein)	0.838 $\pm$ 0.059	0.891 $\pm$ 0.046	0.757 $\pm$ 0.075	0.622 $\pm$ 0.053**
MDA (nmol/mg protein)	1.669 $\pm$ 0.099	1.902 $\pm$ 0.086	1.832 $\pm$ 0.107	2.305 $\pm$ 0.095*,#

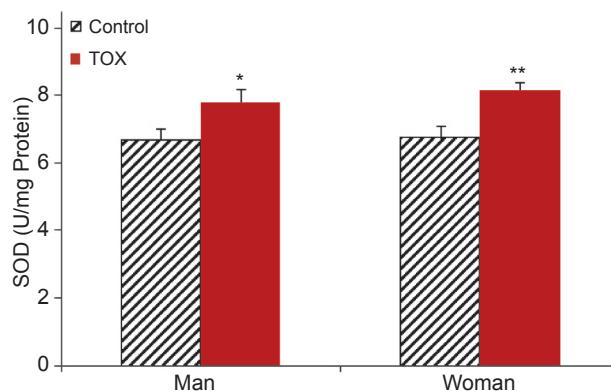
Keys: GSH, glutathione; MDA, malondialdehyde; SEM, standard error of measurement.

Values are expressed as mean  $\pm$  SEM. \**P* = .032 and \*\**P* = .007 vs. the control group; #*P* = .028 vs. infected men.

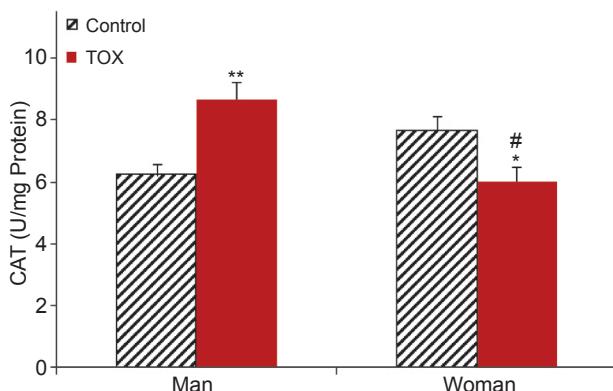
positive for anti-*T. gondii* IgG antibodies including 23 (15.33%) men and 52 (34.66%) women.

Table 2 summarizes the changes of glutathione and malondialdehyde levels in serum of control and case groups. Malondialdehyde level in women of the case group was significantly higher than in the control ( $2.3 \pm 0.09$  vs.  $1.9 \pm 0.09$ , *P* = .032), while glutathione level of the case group was lower ( $0.62 \pm 0.05$  vs.  $0.89 \pm 0.05$ , *P* = .007). In addition, malondialdehyde level was higher in women compared to men in the case group ( $2.3 \pm 0.09$  vs.  $1.8 \pm 0.01$ , *P* = .028). Serum SOD activity was significantly higher in men and women of the case group compared to the control group ( $7.81 \pm 0.38$  vs.  $6.69 \pm 0.17$ , *P* = .045 and  $8.14 \pm 0.23$  vs.  $6.76 \pm 0.36$ , *P* = .004, respectively). (Figure 1).

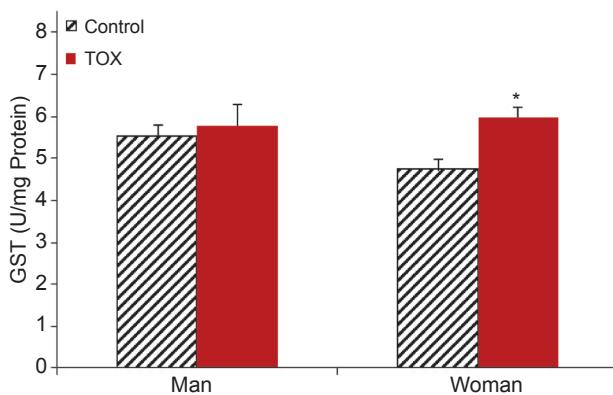
Serum catalase activity had increased in men of the case group (*P* = .006), while it had decreased in women of the case group, as compared to the control group (*P* = .043). Catalase activity in women of the case group was significantly lower than that of men (*P* = .003) (Figure 2). Moreover, serum GST activity in women of the case group was significantly higher compared to the control group ( $5.98 \pm 0.24$  vs.  $4.73 \pm 0.28$ , *P* = .043) (Figure 3).



**Figure 1.** Effect of *Toxoplasma gondii* infection on superoxide dismutase (SOD) activity in control and case groups. Values are expressed as mean  $\pm$  SEM. \**P* = .045 and \*\**P* = .004 vs. the control group.



**Figure 2.** Effect of *Toxoplasma gondii* infection on catalase (CAT) activity in control and case groups. Values are expressed as mean  $\pm$  SEM. \* $P = .043$  and \*\* $P = .006$  vs. the control group; # $P = .003$  vs. infected men.



**Figure 3.** Effect of *Toxoplasma gondii* infection on glutathione S-transferase (GST) activity in control and case groups. Values are expressed as mean  $\pm$  SEM. \* $P = .037$  vs. the control group.

## DISCUSSION

About 30-60% of the population in different countries are infected with *T. gondii*.<sup>35,36</sup> Our results showed that seropositive percentual is somewhat higher in women than in men. Several studies have reported no significant difference regarding the incidence of toxoplasmosis between the two sexes.<sup>27,37,38</sup> However, there are those which point to higher incidence either for men<sup>39,40</sup> or women.<sup>41,42</sup>

Neutrophils and macrophages release ROS as part of the oxidative burst during *T. gondii* infection.<sup>13,22</sup> ROS generation is controlled by the cellular antioxidant enzymes such as SOD and catalase. SOD detoxifies superoxide to hydrogen peroxide ( $H_2O_2$ ) and catalase converts  $H_2O_2$  to  $H_2O$ .<sup>15,43</sup> In this study, SOD activity in men and women and catalase activity in men of the case group were higher, suggesting that elevation of these antioxidant enzymes provides mainly protection against ROS-induced tissue injury. The increased SOD activity was associated with a significant decrease in

catalase activity in women of the case group leading to the accumulation of  $H_2O_2$ , which may be the cause of the induction of oxidative stress.<sup>43</sup> In this regard, Al-Kennany reported a significant decrease in SOD activity in placentae of ewes infected with *T. gondii*.<sup>18</sup> Also, Al-Khshab showed that there were no changes in serum SOD activity in infected women with *T. gondii*.<sup>14</sup>

Glutathione is the most abundant non-protein thiol source in the cell, which acts as a substrate for several enzymes, including glutathione peroxidase and GST.<sup>14,15,43</sup> GST can remove ROS and its levels can reflect the antioxidant capacity of the body.<sup>31</sup> A significant depletion of glutathione and increased GST activity were noted in the present study in serum of women infected with *T. gondii* which was the result of high oxidative stress and glutathione over-use by the cells. Our finding is in agreement with the results of the previous reports in which the infection with *T. gondii* depleted glutathione in host's different tissues.<sup>18,44-46</sup> In addition, the decreased glutathione level in serum of toxoplasmosis patients has been demonstrated.<sup>7,14</sup> Evidences indicate that pre-treatment with N-acetyl cysteine as glutathione precursor can reduce *T. gondii* infection-induced oxidative stress in mice.<sup>46</sup>

Lipid peroxidation is the process of oxidative degeneration of polyunsaturated fatty acids membranes of tissues because of free radical generation. A common marker of lipid peroxidation is malondialdehyde, which has been frequently used as markers of oxidative stress in response to different agents such as infection.<sup>43</sup> In this study, malondialdehyde serum level had increased in women of the case group. The increased lipid peroxidation shows that *T. gondii* infection-induced ROS are not totally scavenged by the antioxidant enzymes in tissues. Numerous studies have shown that malondialdehyde level in various tissues had significantly increased among *T. gondii* infected cats, ewes, chickens and mice.<sup>18,20,44-46</sup> Malondialdehyde level had also increased in serum of toxoplasmosis patients.<sup>7,17,19,21,27</sup> A study has shown that there was no change in serum malondialdehyde level in infected mice with *T. gondii*.<sup>20</sup> Yazar and colleagues have shown that no significant correlation could be found in malondialdehyde levels of either women and men in *T. gondii* infected and control groups.<sup>27</sup>

## CONCLUSION

Seropositive percentual of *T. gondii* was higher for women (34.7%) than for men (15.3%). *T. gondii* infection induces the production of ROS and oxidative stress in serum of women because of the decreased catalase

activity, glutathione depletion, and increasing lipid peroxidation. Increased ROS not only kills the parasites but also damages the host cells. Women are more sensitive to the effects of *Toxoplasma* infection on oxidative stress induction compared to men. Depletion of glutathione leads to oxidized glutathione production, decreasing the reduced glutathione /oxidized glutathione ratio, which may shift cells through apoptosis and necrosis.

## ACKNOWLEDGMENTS

The authors would like to thank Dr. M. Soltanpour for his assistance. This work was supported by a grant from Faculty of Medicine of Baqiyatallah University of Medical Sciences (Grant No. 31).

## CONFLICT OF INTEREST

None declared.

## REFERENCES

1. Abdi J, Shojaee S, Mirzaee A, et al. Seroprevalence of toxoplasmosis in pregnant women in Ilam province, Iran. *Iran J Parasitol.* 2008;3:34-7.
2. Jalalou N, Bandepour M, Khazan H, et al. Recombinant SAG1 Antigen to detect *Toxoplasma gondii* specific immunoglobulin G in human sera by ELISA Test. *Iran J Parasitol.* 2010;5:1-9.
3. Hajsoleimani F, Ataeian A, Nourian AA, et al. Seroprevalence of *Toxoplasma gondii* in pregnant women and bioassay of IgM positive cases in Zanjan, Northwest of Iran. *Iran J Parasitol.* 2012;7:82-6.
4. Hill RD. *Investigation of host responses upon infection of distinct Toxoplasma strains* [PhD dissertation]. USA: University of Tennessee; 2011.
5. Araujo EC, Barbosa BF, Coutinho LB, et al. Heme oxygenase-1 activity is involved in the control of *Toxoplasma gondii* infection in the lung of BALB/c and C57BL/6 and in the small intestine of C57BL/6 mice. *Vet Res.* 2013;44:89.
6. Ghasemian M, Maraghi Sh, Saki J, et al. Determination of antibodies (IgG, IgM) against *Toxoplasma gondii* in patients with cancer. *Iran J Parasitol.* 2007;2:1-6.
7. Karaman U, Celik T, Kiran TR, et al. Malondialdehyde, glutathione, and nitric oxide levels in *Toxoplasma gondii* Seropositive Patients. *Korean J Parasitol.* 2008;46:293-5.
8. Prandota J. Autism spectrum disorders may be due to cerebral toxoplasmosis associated with chronic neuroinflammation causing persistent hypercytokinemia that resulted in an increased lipid peroxidation, oxidative stress, and depressed metabolism of endogenous and exogenous substances. *Res Autism Spect Dis.* 2010;4:119-55.
9. Blader IJ, Saeij JP. Communication between *Toxoplasma gondii* and its host: impact on parasite growth, development, immune evasion, and virulence. *APMIS.* 2009;117:458-76.
10. Garedaghi Y, Bahavarnia SR. Repairing effect of Allium Cepa on Testis degeneration caused by *Toxoplasma gondii* in the rat. *Int J Women Health Reprod Sci.* 2014;2:80-9.
11. Kvitha N, Noordin R, Kit-Lam C, et al. Real time anti-*Toxoplasma gondii* activity of an active fraction of *Eurycoma longifolia* root studied by in situ scanning and transmission electron microscopy. *Molecules.* 2012;17:9207-19.
12. Shrestha SP, Tomita T, Weiss LM, et al. Proliferation of *Toxoplasma gondii* in inflammatory macrophages in vivo is associated with diminished oxygen radical production in the host cell. *Int J Parasitol.* 2006;36:433-41.
13. Correa G, Marques da Silva C, de Abreu Moreira-Souza AC, et al. Activation of the P2X (7) receptor triggers the elimination of *Toxoplasma gondii* tachyzoites from infected macrophages. *Microbes Infect.* 2010;12:497-504.
14. Al-Khshab EM. Some antioxidants level in seropositive toxoplasmosis woman in Mosul. *Tikrit J Pre Sci.* 2010;15:17-22.
15. Halliwell B. Free radicals and antioxidants: Updating a personal view. *Nutr Rev.* 2012;70:257-65.
16. Rafieian-Kopaei M, Baradaran A, Rafieian M. Oxidative stress and the paradoxical effects of antioxidants. *J Res Med Sci.* 2013;18:629.
17. Elsheikha HM, El-Motayam MH, Abouel-Nour MF, et al. Oxidative stress and immune-suppression in *Toxoplasma gondii* positive blood donors: implications for safe blood transfusion. *J Egypt Soc Parasitol.* 2009;39:421-8.
18. Al-Kennany ER. Oxygen free radicals released in placentae of ewes naturally infected with *Toxoplasma gondii*. *Al-Anbar J Vet Sci.* 2009;2:1-6.
19. Azab MS, Abousamra NK, Rahbar MH, et al. Prevalence of, risk factors for, and oxidative stress associated with *Toxoplasma gondii* antibodies among asymptomatic blood donors in Egypt. *Retrovirology.* 2012;9(Suppl 1):P27.
20. Engin AB, Dogruman-Al F, Ercin U, et al. Oxidative stress and tryptophan degradation pattern of acute *Toxoplasma gondii* infection in mice. *Parasitol Res.* 2012;111:1725-30.
21. Abousamra NK. *Toxoplasma gondii* antibodies and associated oxidative stress among asymptomatic blood donors. *Ann Emerg Med.* 2008;52:S138.
22. Ding M, Kwok LY, Schluter D, et al. The antioxidant systems in *Toxoplasma gondii* and the role of cytosolic catalase in defence against oxidative injury. *Mol Microbiol.* 2004;51:47-61.
23. Ding M, Clayton C, Soldati D. *Toxoplasma gondii* catalase: are there peroxisomes in toxoplasma? *J Cell Sci.* 2000;113(Pt 13):2409-19.
24. Pino P, Foth BJ, Kwok LY, et al. Dual targeting of antioxidant and metabolic enzymes to the mitochondrion and the apicoplast of toxoplasma gondii. *PLoS Pathog.* 2007;3:e115.
25. Turrens JF. Oxidative stress and antioxidant defenses: a target for the treatment of diseases caused by parasitic protozoa. *Mol Aspects Med.* 2004;25:211-20.
26. Jafari M, Shirbazou S, Norozi M. Induction of oxidative stress in skin and lung of infected BALB/C mice with

Iranian strain of *Leishmania major* (MRHO/IR/75/ER). *Iran J Parasitol.* 2014;9:60-9.

27. Yazar SL, Kilic E, Saraymen R, et al. Serum malondialdehyde levels in toxoplasma seropositive patients. *Ann Saudi Med.* 2003;23:413-5.

28. Kamper EF, Chatzigeorgiou A, Tsimpoukidi O, et al. Sex differences in oxidant/antioxidant balance under a chronic mild stress regime. *Physiol Behav.* 2009;98:215-22.

29. Winterbourn C, Hawkins R, Brian M, et al. The estimation of red cell superoxide dismutase activity. *Lab Clin Med.* 1975;85:337-41.

30. Aebi H. Catalase in vitro. *Methods Enzymol.* 1984;105:121-6.

31. Habig WT, Jakoby WB. Glutathion S-transferases (rat and human). *Methods Enzymol.* 1981;77:218-31.

32. Tietz F. Enzymic method for quantitative determination of nanogram amount of total and oxidized glutathione: applications to mammalian blood and other tissues. *Anal Biochem.* 1969;27:502-22.

33. Satoh K. Serum lipid peroxidation in cerebrovascular disorders determined by a new colorimetric method. *Clin Chim Acta.* 1978;90:37-43.

34. Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem.* 1976;72:248-54.

35. Alipour A, Shojaee S, Mohebali M, et al. Toxoplasma infection in schizophrenia patients: A Comparative study with control group. *Iran J Parasitol.* 2011;6:31-7.

36. Shirbazou S, Abasian L, Talebi Meymand F. Effects of Toxoplasma gondii infection on plasma testosterone and cortisol level and stress index on patients referred to Sina hospital, Tehran. *Jundishapur J Microbiol.* 2011;4:167-73.

37. Garcia JL, Navarro IT, Ogawa L, et al. Soroprévalencia, epidemiología e avaliação ocular da toxoplasmose humana na zona rural de Jaguapitá (Parana), Brasil. *Rev Panam Salud Publica.* 1999;6:157-63.

38. Jones JL, Kruszon-Moran D, Wilson M, et al. Toxoplasma gondii infection in the United States: seroprevalence and risk factors. *Am J Epidemiol.* 2001;154:357-65.

39. Coelho RA, Kobayashi M, Carvalho LB. Prevalence of IgG antibodies specific to *Toxoplasma gondii* among blood donors in Recife, northeast Brazil. *Rev Inst Med Trop São Paulo.* 2003;45:229-31.

40. Excler JL, Pretat E, Pozzetto B, et al. Sero-epidemiological survey for toxoplasmosis in Burundi. *Trop Med Parasit.* 1988;39:139-41.

41. Abdel-Hameed AA. Sero-epidemiology of toxoplasmosis in Gezira, Sudan. *J Trop Med Hyg.* 1991;94:329-32.

42. Goldsmith RS, Kagan IG, Zarate R, et al. Low Toxoplasma antibody prevalence in serologic surveys of humans in southern Mexico. *Arch Invest Med (Mex).* 1991;22:63-73.

43. Jafari M, Salehi M, Zardooz H, et al. Response of liver antioxidant defense system to acute and chronic physical and psychological stresses in male rats. *EXCLI J.* 2014;13:161-71.

44. Al-Kennany ER. Pathological study on the capability of *Toxoplasma gondii* to induce oxidative stress and initiation a primary lesion of atherosclerosis experimentally in Broiler chickens. *J Anim Vet Adv.* 2007;6:938-42.

45. Al-Kennany ER. Capability of Toxoplasma gondii, to induce an oxidative stress and initiation of atherosclerotic lesions in cats experimentally infected. *Iraqi J Vet Sci.* 2006;20:165-76.

46. Xu X, Liu T, Zhang A, et al. Reactive oxygen species-triggered trophoblast apoptosis is initiated by endoplasmic reticulum stress via activation of caspase-12, CHOP, and the JNK pathway in *Toxoplasma gondii* infection in Mice. *Infect Immun.* 2012;80:2121-32.

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Received February 2014

Accepted April 2014