



# Epigenetic Peculiarity of *Alphapapillomavirus* Genus Based on the CpG Dinucleotides' Island Variability

Embolo Enyegue Elisee Libert<sup>1, 2, 3, \*</sup>, Ndipho Tatou Christian Kitchener<sup>1, 4</sup>, Awalou Halidou<sup>1, 2, 3</sup>, Ananga Noa Sidonie Agnès<sup>5, 6</sup>, Halmata Mohamadou<sup>1, 2, 3</sup>, Godwe Celestin<sup>7, 8</sup>, Eyebe Honore Richard<sup>1, 2</sup>, Evina Evina Basile Junior<sup>9</sup>, Bell Eric Michel<sup>10</sup> and Essame Oyono Jean Louis<sup>1, 2, 3</sup>

<sup>1</sup>Institute of Medical Research and Medicinal Plant Studies (IMPM), Douala, Cameroon

<sup>2</sup>Centre for Research on Health and Priority Diseases (CRSP), Yaoundé, Cameroon

<sup>3</sup>Anatomy and cytology pathology laboratory, Cameroon

<sup>4</sup>Centre for Research on Medicinal Plants and Traditional Medicine, Yaoundé, Cameroon

<sup>5</sup>Faculty of Medicine and Pharmaceutical Sciences, University of Douala, Douala, Cameroon

<sup>6</sup>Douala General Hospital, Douala, Cameroon

<sup>7</sup>Centre for Research on Emerging and Re-emerging Diseases, Yaoundé, Cameroon

<sup>8</sup>Faculty of Sciences, University of Douala, Douala, Cameroon

<sup>9</sup>Catholic University of Louvain, Louvain, Belgium

<sup>10</sup>Makenene Hospital, Makenene, Cameroon

\*Corresponding author: Institute of Medical Research and Medicinal Plant Studies (IMPM), Douala, Cameroon. Email: eliseembolo@yahoo.fr

Received 2023 November 13; Revised 2023 November 28; Accepted 2023 November 29.

## Abstract

**Background:** Methylation plays a crucial role in genome regulation, serving as an essential epigenetic mechanism. CpG islands, which are regions of DNA rich in CpG dinucleotides, are ubiquitous epigenetic regulatory features that can modulate the functional dynamics of genes, thereby influencing the pathophysiology of pathogenic organisms within a host.

**Objectives:** In this study, we aimed to investigate the distribution of CpG islands on *Papillomavirus* genomes and assess their potential impact on the neoplastic progression of different *Alphapapillomavirus* genera within host cells.

**Methods:** We conducted an analysis of dinucleotide frequencies in DNA sequences from various *Alphapapillomavirus* genera. The sequences for our analysis were sourced from the specialized HPV (human papillomavirus) database (pave.niaid.nih.gov). Following a comprehensive comparative examination of entire genomes from different HPV species, we specifically focused on 63 genomes belonging to the *Alphapapillomavirus* genus to identify internal CpG island profiles. These investigations were conducted using online bioinformatics software (DBCAT), and statistical analysis was performed using GraphPad Prism v. 8.0.1.

**Results:** Our preliminary findings revealed that 76.1% of the viruses in our study had fewer than 3 CpG islands but multiple CpG sites, while fewer than 25% of the viruses possessed at least 3 CpG islands. High-risk viruses were identified in 68.42% of genotypes with fewer than 3 CpG islands, whereas low-risk genotypes were observed in 15.7% of cases. Notably, some oncogenic viruses lacked CpG islands entirely, and this pattern was also observed in viruses with a single CpG island, occurring in 50% of cases. Based on the absence of CpG islands, genotypes such as HPV 67, HPV 97, and HPV 34 could potentially be classified as carcinogenic. Additionally, the distribution of CpG islands across HPV genes did not appear to be random. Genes like L2 and E2 contained a CpG island in 50% of cases, whereas oncogenes E6 and E7 consistently had a CpG island in 100% of cases in high-risk genotypes.

**Conclusions:** Our findings suggest that the loss of 1 or more CpG islands could potentially transform an HPV genotype into a high-risk genotype. This process may be accelerated in the presence of host-related risk factors. Further research is required to validate whether CpG island distribution can aid in identifying oncogenic viruses.

**Keywords:** Epigenetic, *Alphapapillomavirus*, CpG Islands, Methylation, Oncogenic Viruses

## 1. Background

Papillomaviruses (HPVs) are a persistent and highly adaptable group of viruses that have evolved in tandem with their hosts to replicate within specific stratified

epithelial niches. To date, nearly 450 distinct HPV types have been isolated and sequenced (1). The viral replication cycle of these viruses is intricately linked to the differentiation of the infected epithelium (2). The epidemiology of HPV infections reflects the genetic

diversity of HPVs, with specific geographical distributions of different variants of an HPV type (3). This genetic diversity may have implications for the pathophysiology of HPV infection. It is well-established that co-infection with multiple high-risk HPV types is a risk factor for cancer progression, and polymorphisms are associated with an elevated risk of cancer advancement (4). However, understanding the potential methylation profile of human *Papillomaviruses* (HPVs) as a means to elucidate the purpose of this diversity remains a challenge. Studies have indicated that the methylation of the virus's DNA is linked to its responsiveness to treatment for the associated pathology (5). An investigation detected increased methylation at 13 CpG sites as the disease progressed, and a high level of methylation was correlated with the risk of CIN2+ in a study examining the relationship between CpG methylation in HPV-16 L1 and persistent infections, as well as the development of cervical carcinoma in Uyghur women (6). These findings suggest that the methylation of CpG sites in HPV could be a valuable predictor of HPV infection persistence and the development of cervical pre-cancer.

## 2. Objectives

This study aimed to assess the potential utility of the presence or absence of CpG islands in determining the likely oncogenicity of a virus.

## 3. Methods

### 3.1. Study Type

This was a retrospective descriptive study.

### 3.2. Sample Collection Strategy

The sequences for analysis were obtained from [pave.niaid.nih.gov](http://pave.niaid.nih.gov) (*Alphapapillomavirus*). A total of 63 HPV genotype sequences were extracted from [pave.niaid.nih.gov](http://pave.niaid.nih.gov), a specialized HPV database.

### 3.3. Selection Criteria for CpG Islands

During the study, any region with a minimum length of 200 base pairs, a GC content of at least 50%, and a CpG ratio of at least 60% were considered as CpG islands.

### 3.4. Bioinformatics Analysis

Sequence analysis was conducted using [dbcat.cgm.ntu.edu.tw](http://dbcat.cgm.ntu.edu.tw), an online methylation analysis tool consisting of 3 components: A CpG Island Finder, a genome query browser, and an analytical tool for methylation microarray data. This analytical tool can process raw data from scanners and identify genes with methylated regions that may impact gene expression regulation.

Statistical Analysis: Statistical analyses were performed using GraphPad Prism v. 8.0.1 for percentages.

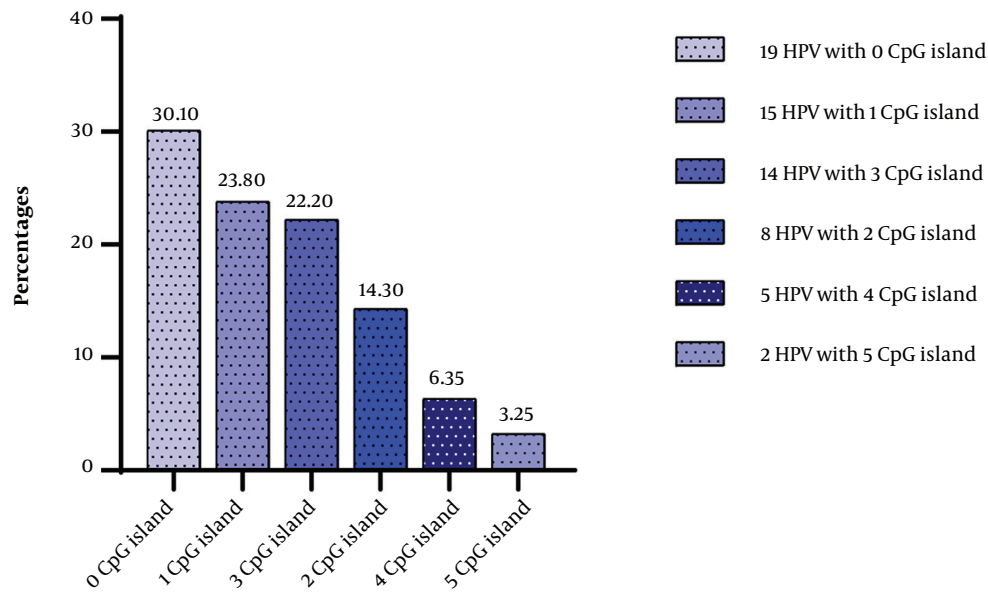
## 4. Results

### 4.1. Distribution of the Number of CpG Islands

The distribution of the number of CpG islands in all the analyzed genomes is presented in [Figure 1](#) below. It is noteworthy that a significant portion of the encountered viruses lacked CpG islands (30.1%). Conversely, viruses with the highest number of CpG islands constituted only 3.25% of the total number of viruses in the study.

### 4.2. Profile of HPV Without CpG Islands

The strains listed in [Table 1](#) were found to have no CpG islands when their genomes were examined. Mucosal tropic viruses were found in all of these genotypes. The viruses without detected CpG islands belonged to 5 *Alphapapillomavirus* species (Alpha HPV 6, Alpha HPV 7, Alpha HPV 9, Alpha HPV 10, Alpha HPV 11). There are no significant types in the Alpha HPV 10 species; these species are generally regarded as low-risk viruses commonly found in *condyloma acuminata* and laryngeal papillomatosis. There are no major CpG islands in the genomes of subtypes HPV 13, HPV 44, and HPV 73. We found HPV type 16 and 6 subtypes in *Alphapapillomavirus* 9, which contains the high-risk genotypes responsible for mucosal lesions. These include HPV 31, HPV 33, HPV 35, HPV 52, HPV 58, and HPV 67. HPV 18, which includes 6 subtypes, such as HPV 39, HPV 45, HPV 56, HPV 68, HPV 70, and HPV 97, manifests as *Alphapapillomavirus* 7, which also has high-risk genotypes often responsible for mucosal lesions. *Alphapapillomavirus* 11 species and its major type, HPV 34, were also found, as well as *Alphapapillomavirus* 6 species and its major type, HPV 53. Results indicate the viral genotypes of HPV according to their oncogenic potential, which did not show any CpG islands. It can be seen that high-risk viruses represent about 69% of the viruses without CpG islands in their genomes, while low-risk oncogenic viruses are also observed with a frequency of 15.79%. However, we also observed viruses whose oncogenic risk is not well-defined that showed an absence of CpG islands, notably viruses (67, 97, and 34).



### Alphapapillomavirus CpG island number

**Figure 1.** Frequencies of CpG island observed

**Table 1.** Presentation of HPV with 0 CpG Island

CpG Island Number and Species	Types Species	Other Papillomavirus Type	Total Number of HPV
<b>0</b>			19
10	-	HPV 13 (X62843); HPV 44 (U31788); HPV 73 (X94165)	
9	HPV 16 (K02718)	HPV 31 (J04353); HPV 33 (M12732); HPV 35 (X74476); HPV 52 (X74481); HPV 58 (D90400); HPV 67 (D21208)	
7	HPV 18 (X05015)	HPV 39 (M62849); HPV 45 (X74479); HPV 56 (X74483); HPV 68 (X67161); HPV 70 (U21941); HPV 97 (DQ080080)	
11	HPV 34 (X74476)	-	
6	HPV 53 (X74482)	-	
<b>Total</b>			
5	4	15	

#### 4.3. Profile of HPV with a Single CpG Island

Table 2 below shows the different types of viruses encountered that had only 1 CpG island. Indeed, 7 different species of *Alphapapillomaviruses* were found, which included 5 major and 10 minor *Papillomavirus* types. A total of 15 viruses had a CpG island; among these genotypes, 46.66% were high-risk (26, 30, 51, 59, 66, 69, 82) and low-risk (6, 7, 11, 32, 43, 54, 91). The vast majority of the CpG islands were identified in the E2 genes in almost 80% of the cases. The size of the CpG clusters ranged from 224 to 410 nucleotides.

#### 4.4. CpG Islands' Positions on HPV

Figure 2 below shows the distribution pattern of the islands for different human *Papillomavirus* genes. It can be seen that 50% of the CpG islands are located on gene 2 for high-risk genotypes, while for low-risk genotypes, CpG islands were found on gene E2 in 40% of cases. One genotype with unknown oncogenic potential also harbored CpG islands on its E2 gene. No CpG islands were found in the L2 gene of the high-risk genotypes, but the only islands found in the L2 gene for viruses with a single island were found in a low-risk genotype. We were able to identify islands in the E1 gene in equal proportions for both high- and low-risk genotypes. The HPV 30 genotype, which is a high-risk genotype, showed that the CpG island encountered overlapped 3 genes (E6-E7-E1).

#### 4.5. Profile of HPV with 2 CpG Islands

The HPVs presented in Table 3 below exhibit a significant absence of major HPV types. *Alphapapillomavirus* species 2 had no major types, compared to 3 minor types. HPV species 8, 6, and 3 only showed subtypes of HPV, while HPV species 15 had a major type. Seven types were not clarified (3, 71, 74, 77, 83, 94, 106), and 1 was low-risk (40).

#### 4.6. Position of CpG Islands on Genes for Viruses with 2 Islands

Figure 3 below illustrates the various identification sites of the CpG islands and the viral types in which they were found. There was a complete absence of high-risk viruses among the viruses that had 2 CpG islands, but 1 low-risk virus was present. Almost all the encountered viruses had an unknown oncogenic potential, with nearly 100% of the genes harboring CpG islands, except for the E1 gene, where CpG islands were encountered at a frequency of 75%.

#### 4.7. Profile of HPV with 2 CpG Islands

Table 4 below displays the various *Alphapapillomavirus* species, in which we identified 3 CpG islands. We can observe that there were 5 species of *Alphapapillomavirus* that could be identified, with only 2 species presenting major HPV types (HPV 10 and HPV 14). The identified HPV subtypes numbered 12.

#### 4.8. Genomic Regions Hosting 3 CpG Islands

A total of 13 viral genotypes were found to contain 3 CpG islands distributed in different viral genes. Among these viruses, we identified 69.23% with unknown oncogenic potential (10, 27, 28, 29, 78, 84, 102, 117, 125) compared to 30.76% of low-risk viruses (42, 72, 81, 89). The range of variation in the CpG islands encountered was from 1470 to 233 nucleotides. No high-risk virus was observed among the viruses with 3 CpG islands.

#### 4.9. Position of CpG Islands on Genes for Viruses with 3 Islands

The frequency of the various CpG localization on viruses according to genes is depicted in Figure 4 below. It is clear that genes with low oncogenic risk had fewer than 45% of their genes harboring island CpGs, whereas genes with unknown oncogenic potential had viruses with more than 60% of their genes occupied by island CpGs.

#### 4.10. Profile of HPV with 4 Islands

Table 5 below shows 3 species of HPV, with 1 main type of HPV (HPV2) and 3 secondary HPV types. It can be seen that all of these viruses (100%) were viruses of unknown oncogenic potency. The range of CpG islands observed was between 1247 and 221.

#### 4.11. Profile of HPV with 5 CpG Islands

Table 6 below shows the HPV viruses in which 5 CpG islands were observed. Indeed, 2 viruses among those selected showed 5 CpG islands; they are the same species of *Alphapapillomavirus*, species 3, which presented a main genotype (HPV 61), which has a low oncogenic risk, and a secondary genotype (HPV 62), so the oncogenic power is unknown.

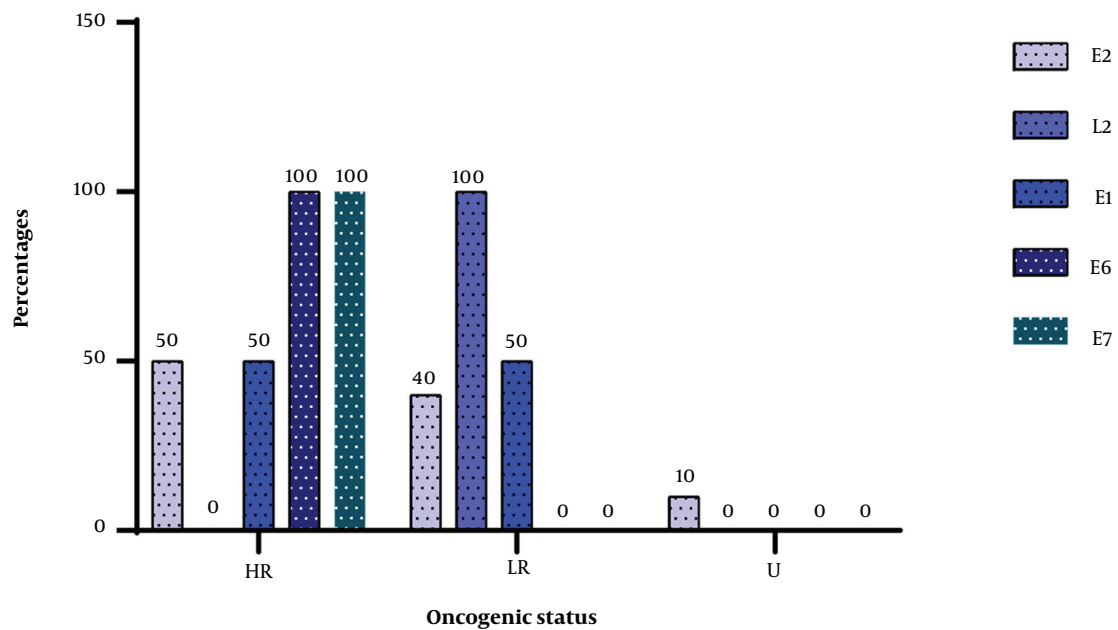
### 5. Discussion

This study aimed to assess the distribution of CpG island profiles on *Papillomavirus* genomes and comprehend the impact of these predispositions on the neoplastic dynamics of different types of *Alphapapillomavirus* genus in host cells. Preliminary results indicate that HPV genotypes exhibit epigenomic variations, resulting in different CpG islands in each HPV

**Table 2.** Presentation of HPV with 1 CpG Island

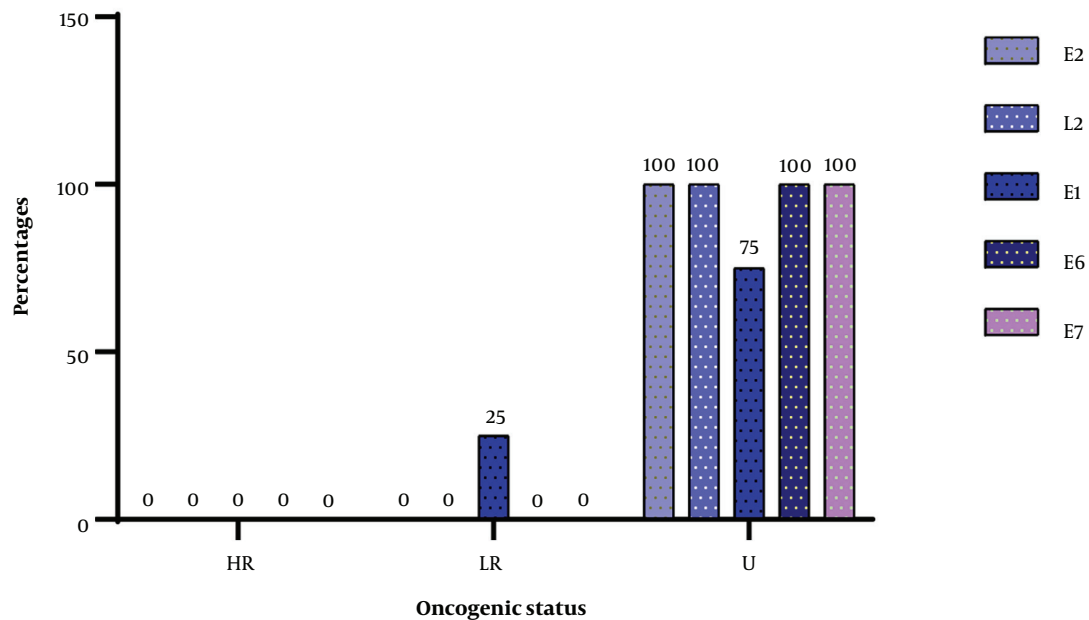
CpG Island Number and Species	Types Species	Other Papillomavirustype	Total Number of HPV
1			15
10	HPV 6 (X00203)	HPV 11 (M14119)	
8	HPV 7 (X74463)	HPV 43 (AJ620205); HPV 91 (AF419318)	
5	HPV 26 (X74472)	HPV 51 (M62877); HPV 69 (AB027020); HPV 82 (AB027021)	
1	HPV 32 (X74475)	-	
6	-	HPV 30 (X74474); HPV 66 (U31794)	
13	HPV 54 (U37488)	-	
7	-	HPV 59 (X77858); HPV (KR816168)	
<b>Total</b>			
7	5	10	

<sup>z</sup> Abbreviations: LR, low risk; HR, high risk; U, unknown.

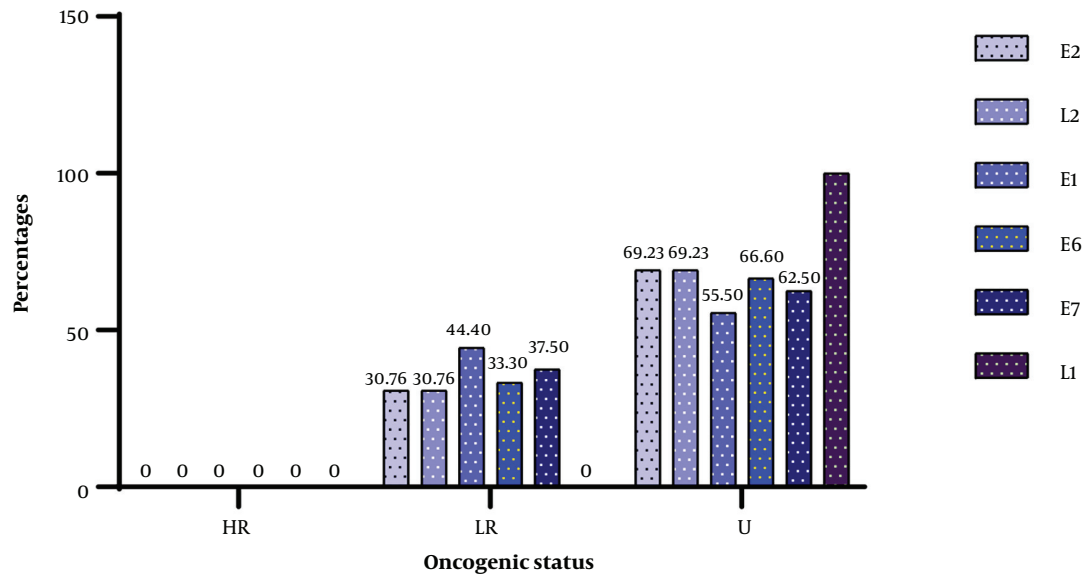
**Figure 2.** Distribution of single CpG islands on the HPV genome (LR, low risk; HR, high risk; U, unknown)

**Table 3.** HPV with 2 CpG Islands

CpG Island Number and Species	Types Species	Other <i>Papillomavirus</i> Type	Total Number of HPV
<b>2</b>			<b>8</b>
2	-	HPV 3 (X74462); HPV 77 (Y15175); HPV 94 (AJ620211)	
8	-	HPV 40 (X74478)	
15	HPV 71 (AB040456)	-	
6	-	HPV 74 (U40822)	
3	-	HPV 83 (AF151983)	
14	-	HPV 106 (DQ080082)	
<b>Total</b>			
6	1	7	

**Figure 3.** Distribution of CpG islands according to the genes of viruses with 2 CpG islands (LR, low risk; HR, high risk; U, unknown)**Table 4.** Presentation of HPV with 3 CpG Islands

CpG Island Number and Species	Types Species	Other <i>Papillomavirus</i> Type	Total Number of HPV
<b>3</b>			<b>14</b>
2	HPV 10 (X74465)	HPV 28 (U31783); HPV 29 (U31784); HPV 78 (KC138720); HPV 117 (GQ246950); HPV 125 (FN547152)	
4	-	HPV 27 (X73373)	
1	-	HPV 42 (M73236)	
3	-	HPV 72 (X94164); HPV 81 (AJ620209); HPV 84 (AF293960); HPV 89 (AF436128); HPV 102 (DQ080083)	
14	HPV 90 (AY057438)	-	
<b>Total</b>			
5	2	12	



**Figure 4.** Distribution of CpG islands according to the genes of viruses with 3 CpG islands (LR, low risk; HR, high risk; U, unknown)

**Table 5.** Presentation of HPV with 4 CpG Islands

CpG Island Number and Species	Types Species	Other Papillomavirus Type	Total Number of HPV
4			4
2	-	HPV 160 (AB745694)	
4	HPV 2 (X55964)	HPV 57 (X55965)	
3	-	HPV 114 (GQ244463)	
<b>Total</b>			
3	1	3	

**Table 6.** Presentation of HPV with 5 CpG Islands

CpG Island Number and Species	Types Species	Other Papillomavirus type	Total Number of HPV
5			2
3	HPV 61 (U31793)	HPV 62 (AY395706)	
<b>Total</b>			
1	1	1	

genotype or subgenotype. This variation may be attributed to different mucocutaneous tropisms of various human *Papillomavirus* genotypes, potentially influencing the CpG island positions within the genome. These findings align with the work of Chen and colleagues, who observed differences in CpG island profiles in hepatitis B virus (7).

However, some HPVs did not exhibit CpG islands. This absence of CpG islands in certain HPV viruses is predominantly observed in high-risk HPV viruses, suggesting that the absence of CpG islands in these viruses

could be a crucial biomarker for their identification. It is also possible that the absence of these CpG islands is a result of the virus's oncoproteins altering the host environment. Previous research by XIA and subsequent studies by Thomas and Sudhir proposed the hypothesis that genomic methylation may be influenced by host-specific defenses (8, 9).

Regarding their oncogenic potential, high-risk viruses accounted for approximately 69% of viruses lacking CpG islands in their genomes, while low-risk oncogenic



viruses accounted for 15.79% of all viruses. This increased absence of CpG islands in high-risk viruses could be a direct consequence of their evolutionary lineage, as demonstrated in a study by Mohita and Perumal, which concluded that CpG depletion among *Papillomaviruses* and polyomaviruses is linked to the evolutionary lineage of the infected host (10). Silvia and collaborators showed a positive correlation between observed and expected CpG values, with mucosal high-risk (HR) virus types exhibiting the smallest O/E ratios (11).

However, viruses with an undetermined carcinogenic risk, such as HPV 67, HPV 97, and HPV 34, were also found to lack CpG islands. The presence of these viruses with a similar profile to known oncogenic risk viruses suggests their potential oncogenicity within host cells. This observation is logical since genotypes 67 and 97 are subtypes of HPV 16 and HPV 18, which are known to have oncogenic potential. Some authors have isolated these genotypes from high-grade intraepithelial lesions (12,13).

In some viruses, only 1 CpG island was found, and it was present in 46.66% of cases among high- and low-risk genotypes. In 80% of these cases, the CpG island was predominantly found in the E2 genes. The presence of CpG islands in E2 may result from the regulatory role this gene plays in viral replication. The viral E2 protein, a sequence-specific DNA-binding protein, regulates the *Papillomavirus* replication cycle and can function as a transcriptional activator or regulator of viral gene expression (14). Several other genes also have CpG islands, indicating the presence of methylation sites within these genes. Given that methylation is a critical phenomenon in the genome, the presence of these CpG islands within these genes serves as tangible evidence of the virus's regulatory quality, although it is reversible. CpG islands are essential for protecting adjacent housekeeping genes from de novo DNA methylation and maintaining them in a transcriptionally active state (15).

In this study, we found that the size of CpG islands varied with the number of islands encountered. Larger CpG islands were more likely to be found in viruses with multiple islands. This variation may be linked to their oncogenic potential, as suggested by the study conducted by Xue and colleagues, who demonstrated that the lengths and QS heterogeneity of CpG islands differ according to the clinical phases of infection. These data could partly explain the different clinical features observed during various infection phases, warranting further investigation (16).

### 5.1. Conclusions

This study aimed to assess the distribution of CpG islands in the genomes of *Alphapapillomaviruses*. It reveals

that within this group of *Papillomaviruses*, CpG islands are notably absent in viruses with a clear oncogenic potential. This pattern of CpG island distribution in viruses could serve as a critical element for identifying oncogenic HPVs. The absence of CpG islands appears to represent an evolutionary characteristic of HPV-related molecular processes. Notably, the more capable an HPV is of causing damage to host tissues (being oncogenic), the fewer CpG islands it possesses. This observation may indeed serve as an evolutionary hallmark for *Alphapapillomaviruses*, which are characterized by the complete absence of CpG islands. The findings of this study could provide a foundational basis for further research on the role of CpG islands in determining HPV oncogenic potential.

### Acknowledgments

The authors express their sincere gratitude to all researchers who have contributed to the publication of HPV sequences in the GenBank.

### Footnotes

**Authors' Contribution:** All authors cooperated equally in this study.

**Conflict of Interests:** The authors declare no conflicts of interest.

**Data Availability:** The data presented in this study are available upon request from the corresponding author. Our institution is currently working on establishing a data repository.

**Funding/Support:** This research received no external funding or support.

### References

1. McBride AA. Human papillomaviruses: Diversity, infection and host interactions. *Nat Rev Microbiol.* 2022;**20**(2):95-108. [PubMed ID: 34522050]. <https://doi.org/10.1038/s41579-021-00617-5>.
2. De Sanjose S, Brotons M, Pavon MA. The natural history of human papillomavirus infection. *Best Pract Res Clin Obstet Gynaecol.* 2018;**47**:2-13. [PubMed ID: 28964706]. <https://doi.org/10.1016/j.bpobgyn.2017.08.015>.
3. Egawa N, Egawa K, Griffin H, Doorbar J. Human papillomaviruses; epithelial tropisms, and the development of neoplasia. *Viruses.* 2015;**7**(7):3863-90. [PubMed ID: 26193301]. [PubMed Central ID: PMC4517131]. <https://doi.org/10.3390/v7072802>.
4. Ma L, Cong X, Shi M, Wang XH, Liu HY, Bian ML. Distribution of human papillomavirus genotypes in cervical lesions. *Exp Ther Med.* 2017;**13**(2):535-41. [PubMed ID: 28352328]. [PubMed Central ID: PMC5348701]. <https://doi.org/10.3892/etm.2016.4000>.



5. Jones SEF, Hibbitts S, Hurt CN, Bryant D, Fiander AN, Powell N, et al. Human papillomavirus DNA methylation predicts response to treatment using cidofovir and imiquimod in vulval intraepithelial neoplasia 3. *Clin Cancer Res*. 2017;**23**(18):5460–8. [PubMed ID: 28600473]. <https://doi.org/10.1158/1078-0432.CCR-17-0040>.
6. Niyazi M, Sui S, Zhu K, Wang L, Jiao Z, Lu P. Correlation between Methylation of Human Papillomavirus-16 L1 Gene and Cervical Carcinoma in Uyghur Women. *Gynecol Obstet Invest*. 2017;**82**(1):22–9. [PubMed ID: 26954462]. <https://doi.org/10.1159/000444585>.
7. Chen L, Shi Y, Yang W, Zhang Y, Xie Q, Li Y, et al. Differences in CpG island distribution between subgenotypes of the hepatitis B virus genotype. *Med Sci Monit*. 2018;**24**:6781–94. [PubMed ID: 30253420]. [PubMed Central ID: PMC6180904]. <https://doi.org/10.12659/MSM.910049>.
8. Xia X. Extreme genomic CpG deficiency in SARS-CoV-2 and evasion of host antiviral defense. *Mol Biol Evol*. 2020;**37**(9):2699–705. [PubMed ID: 32289821]. [PubMed Central ID: PMC7184484]. <https://doi.org/10.1093/molbev/msaa094>.
9. Leitner T, Kumar S. Where Did SARS-CoV-2 Come From? *Mol Biol Evol*. 2020;**37**(9):2463–4. [PubMed ID: 32893295]. [PubMed Central ID: PMC7454771]. <https://doi.org/10.1093/molbev/msaa162>.
10. Upadhyay M, Vivekanandan P. Depletion of CpG dinucleotides in papillomaviruses and polyomaviruses: A role for divergent evolutionary pressures. *PLoS One*. 2015;**10**(11): e0142368. [PubMed ID: 26544572]. [PubMed Central ID: PMC4636234]. <https://doi.org/10.1371/journal.pone.0142368>.
11. Galvan SC, Martinez-Salazar M, Galvan VM, Mendez R, Diaz-Contreras GT, Alvarado-Hermida M, et al. Analysis of CpG methylation sites and CGI among human papillomavirus DNA genomes. *BMC Genomics*. 2011;**12**:580. [PubMed ID: 22118413]. [PubMed Central ID: PMC3293833]. <https://doi.org/10.1186/1471-2164-12-580>.
12. Kirii Y, Matsukura T. Nucleotide sequence and phylogenetic classification of human papillomavirus type 67. *Virus Genes*. 1998;**17**(2):117–21. [PubMed ID: 9857984]. <https://doi.org/10.1023/a:1008002905588>.
13. Chen Z, Fu L, Herrero R, Schiffman M, Burk RD. Identification of a novel human papillomavirus (HPV97) related to HPV18 and HPV45. *Int J Cancer*. 2007;**121**(1):193–8. [PubMed ID: 17351898]. <https://doi.org/10.1002/ijc.22632>.
14. Nishimura A, Ono T, Ishimoto A, Dowhanick JJ, Frizzell MA, Howley PM, et al. Mechanisms of human papillomavirus E2-mediated repression of viral oncogene expression and cervical cancer cell growth inhibition. *J Virol*. 2000;**74**(8):3752–60. [PubMed ID: 10729150]. [PubMed Central ID: PMC111884]. <https://doi.org/10.1128/jvi.74.8.3752-3760.2000>.
15. Hejnar J, Hajkova P, Plachy J, Elleder D, Stepanets V, Svoboda J. CpG island protects Rous sarcoma virus-derived vectors integrated into nonpermissive cells from DNA methylation and transcriptional suppression. *Proc Natl Acad Sci U S A*. 2001;**98**(2):565–9. [PubMed ID: 11209056]. [PubMed Central ID: PMC14627]. <https://doi.org/10.1073/pnas.98.2.565>.
16. Xue Y, Wang MJ, Huang SY, Yang ZT, Yu DM, Han Y, et al. Characteristics of CpG Islands and their quasispecies of full-length hepatitis B virus genomes from patients at different phases of infection. *Springerplus*. 2016;**5**(1):1630. [PubMed ID: 27722049]. [PubMed Central ID: PMC5031574]. <https://doi.org/10.1186/s40064-016-3192-3>.