

Immunohistochemical Expression of *p53* and *Ki-67* Genes in Gastric Cancer and Precancerous Lesions in the Patients with *Helicobacter pylori* Infection

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Received 2016 August 08; Revised 2016 August 21; Accepted 2016 August 21.

Abstract

Background: *Helicobacter pylori* (*H. pylori*) infection is considered as a carcinogen for gastric mucosa. The mechanism by which *H. pylori* are involved in gastric carcinogenesis is still unclear.

Objectives: The current study aimed to investigate the effects of *H. pylori* infection on immunohistochemical expression of *p53* and *Ki-67* genes in gastric specimens of patients with intestinal metaplasia (IM), dysplasia (DYS) and gastric cancer (GC).

Methods: In the study, 42 cases with IM, 38 with DYS and 42 with GC were selected from pathology archive of Ali-ebne-Abitaleb hospital, Zahedan, Iran; *p53* and *Ki-67* immunohistochemical study was done, accordingly. Kruskal-Wallis and Mann-Whitney U tests were used to compare the groups. Level of significance was set at $P < 0.05$.

Results: In IM specimens, expression of *p53* and *Ki-67* was significantly higher in the cases with *H. pylori* infection than those not infected with *H. pylori*. In DYS specimens, in the group with *H. pylori* infection, the expression of *p53* was significantly higher while expression of *Ki-67* was significantly lower than the group without *H. pylori* infection. In GC specimens, the expression of *p53* was significantly higher in the group with *H. pylori* infection compared to the group without *H. pylori* infection. Expression of *Ki-67* in the specimens with *H. pylori* infection was not significantly different from those without *H. pylori* infection.

Conclusions: The study results showed that *p53* expression increased in IM, DYS and GC cases with *H. pylori* infection compared to those without *H. pylori* infection. The results also suggested that, *Ki-67* expression was different in precancerous stages of gastric carcinogenesis, based on the infection with *H. pylori*.

Keywords: Immunohistochemistry, Gastric Cancer, *p53*, *Ki-67*, *Helicobacter pylori*

1. Background

Gastric carcinoma (GC) is the fourth most common cancer and the second most common cause of tumor related deaths worldwide (1, 2). Despite its decreasing incidence, GC remains a major health problem (3, 4). So that, approximately 700,000 people die of GC every year (5). About 70% of cancer deaths occur in the developing countries (3). There are variations in GC geographical distribution from place to place and from one generation to another (1). The highest prevalence rates are reported in East Asia, Eastern Europe and South America, and the lowest prevalence in North America and a large part of Africa (3). In Asia, the countries with high prevalence include Japan, China, South Korea and countries with low prevalence include India, Pakistan and Thailand. GC is the most common malignancy in Iran. In the North-West of Iran, the

highest prevalence is in Ardabil. In South-East Iran the GC is the second intestinal-gastrointestinal malignancy (6). Its etiology is not fully elucidated yet. Epidemiologic data indicate that environmental factors are important in the pathogenesis of GC (7). These factors include *Helicobacter pylori* (*H. pylori*) infection, high salt intake, high consumption of nitrous compounds, cigarette smoking, gastric surgeries, gastric ulcer and blood group A as a predisposing genetic factor. Among these, *H. pylori* infection is established as a necessary factor in the development of GC (8-10). Although many studies show that *H. pylori* infection is closely associated with gastric carcinogenesis, the mechanisms and processes of carcinogenesis by *H. pylori* are largely unknown (11).

Helicobacter pylori are Gram-negative spiral-shaped bacteria and can colonize the gastric mucosa for a long

time (12). Following the bacterial colonization and gastric mucosal damage, epithelial cell proliferation increases in response to injury. This event may cause increment of DNA damage, enhancement of malignancy and finally development of GC (13).

Gastric carcinogenesis is a multi-step process, ranging from chronic gastritis to atrophy; loss of gastric glands, intestinal metaplasia (IM), dysplasia (DYS) and ultimately gastric cancer (2, 14).

Chronic gastritis is the inflammation of gastric mucosa. It is characterized by elementary lesions whose extent and distribution are related to their etiology and host responses. Morphological changes in gastric mucosa of gastritis consist of epithelial degeneration, neutrophilic infiltration, mononuclear inflammatory cell infiltration, atrophy, parietal cell alterations and finally IM (15).

IM is defined as replacement of the gastric epithelial mucosa by an epithelium resembling the small intestine (16, 17). IM and DYS are considered as gastric precancerous lesion. They are created as a result of long-term and progressive gastric mucosa damages (18).

DYS is characterized microscopically by a set of histological alterations. The diagnosis relies on cytological parameters such as cytoplasmic mucin depletion, cellular crowding, pleomorphism, stratification, loss of cellular and nuclear polarity, nuclear hyperchromatism, increased nuclear to cytoplasmic (N/C) ratio and increased mitotic activity. Architectural abnormalities comprise glandular disarrangement or budding with irregular branching, dilatation and intraluminal folding. Morphologically, DYS differs from normal gastric mucosa, and it is shown that DYS is associated with a higher risk of gastric carcinogenesis (19).

Several biological markers are tested as potential predictors of the GC outcome and some of them are essential to develop malignancy. The *p53* gene is one of the tumor suppressor genes that play a complex role in the cell cycle regulation. Mutation of this gene is very common and associated with carcinogenesis, which results in the loss of gene function (9).

Recently, Shadifar et al. (11) showed that *H. pylori* infection has a direct role on *p53* mutagenesis in patients with IM, DYS and GC. Tumor suppressor gene or *p53* produces the *P53* protein, which is located in the cell nucleus (2). Increase of *P53* protein production is very common in human cancers (20, 21).

The *p53* gene activates when DNA is damaged. It leads to cell cycle arrest in G₁-phase (2) and repairs the damaged DNA, or cell apoptosis (22). If *p53* gene is damaged, it can allow a cell to go from normal into uncontrolled growth, thus leads to carcinogenicity (23). The wild-type *P53* protein has a very short half-life of about 5 to 20 min-

utes and does not accumulate inside cells for a long time (24). The *p53* damaged gene can cause production of a protein with long half-life that remains within the cell for a long time. Therefore, it can be detected using immunohistochemistry method (25). It is shown that accumulation of *P53* protein increased during the process of gastro carcinogenicity with the progression from gastritis toward IM, DYS and GC (26).

Another marker that is implicated in the progression of cancer is the *Ki-67* antigen. At first, it was recognized by Gerdes et al. (cited in Inwald et al. (27)) in the early 1980s. The *Ki-67* or Nuclear antigen is related to cell proliferation process. It is expressed at different phases of the cell cycle (14). Cells express the *Ki-67* antigen during G₁, S, G₂, and M phases, but this antigen cannot be observed in the resting phase or G₀ phase (14, 28, 29). The expression of *Ki-67* increases in many tumors (29). Forones et al. (30) demonstrated that the *Ki-67* proliferating index increased during the process of gastro carcinogenicity from IM to GC.

Since GC is a multifactorial disease with significant geographical variations, confused pathogenesis and controversies in the results among different geographic regions determined by numerous environmental and host genetic factors (31), the molecular basis of GC is investigated in numerous studies. The lack of data in the studied geographical region and population and also lack of enough knowledge about the role of *H. pylori* infection in the process of GC were major motivations behind the current study. Hence, the contribution of this paper is to evaluate the role of *H. pylori* infection as a gastric cancer risk factor on *p53* and *Ki-67* immunohistochemical expressions in cases of IM, DYS and GC.

2. Objectives

The current study aimed to evaluate the effects of *H. pylori* infection on immunohistochemical expression of *p53* and *Ki-67* in gastric specimens of patients with IM, DYS and GC.

3. Methods

3.1. Study Design

This descriptive-analytic study was conducted on 122 gastric specimens selected from the pathologic files archive of Ali-ebne-Abitaleb hospital, Zahedan University of Medical Sciences (ZAUMS), Zahedan, Iran. All samples had been obtained by endoscopic biopsy from patients admitted from 2010 to 2015. The inclusion criteria of the selection of archival histological specimens were suitable formalin fixed paraffin-embedded tissue from patients

with gastric cancer and precancerous lesion and a complete clinicopathological data. The exclusion criteria were the samples from patients with immune disorders, recent recipient of steroids and anti-*H. pylori* drugs.

The project was approved by the ethics committee of Zahedan University of Medical Sciences (IR.ZAUMS.REC.1394.53).

The study specimens were classified according to the histological diagnosis: 42 IM (20 cases with *H. pylori* infection and 22 without *H. pylori* infection), 38 DYS (19 cases with *H. pylori* infection and 19 without *H. pylori* infection) and 42 GC samples (19 cases with *H. pylori* infection and 23 without *H. pylori* infection).

Selected formalin-fixed paraffin-embedded tissue blocks were cut into three micrometer thick sections mounted on glass slides for histological and immunohistochemical evaluations.

Histological evaluations of the specimens were done based on H&E sections and all selected samples were re-examined by an expert pathologist to ensure prior diagnosis.

3.2. Detection of *Helicobacter pylori* Infection in Gastric Biopsy Specimens

Helicobacter pylori in the gastric tissue were detected by histological examination; using H&E, modified Giemsa and carbol fuchsin staining methods. Specimens in which all three assays were positive were classified as the group with *H. pylori* infection specimens and when the results of all three tests were negative, they were considered as group not infected with *H. pylori* specimens.

3.3. Immunohistochemical Analysis

Immunohistochemical expression of *p53* and *Ki-67* genes in biopsy samples of patients with IM, DYS and GC were performed using the monoclonal mouse anti-human antibodies *p53* (RTU-*p53*-DO7, Novocastra, England), and *Ki-67* (RTU-*Ki-67*-MMI, Novocastra, England), respectively. Time, temperature and storage conditions were according to the manufacturer's instructions. The tissue samples were cut into three micrometer sections using the fully automated Leica RM2255 instrument (Leica, Germany) and mounted on HistoGrip (CEDARLANE, Canada) coated slides. The sections were deparaffinized and rehydrated. Then they were transferred into sodium citrate buffer (pH: 6.0) and boiled at 120°C for a total of 20 minutes. After that, the sections were left to cooldown to room temperature for 15 minutes. Immunostaining was precisely described in the authors' previous study (21).

Positive and negative controls were performed at the same time for each section. The positive controls were

respectively appendix tissue for *Ki-67*, and a specimen of colon cancer; previously shown to express *p53* for *p53* immunostainings. Negative control was obtained by incubation of slides in Tris-buffered saline (TBS) buffer and omitting the primary antibodies. Slides were evaluated under a light microscope at high-power magnification (X400) and scoring was done by two expert histologists blindly.

3.4. Evaluation of Immunostaining

The intensity of staining for *p53* and *Ki-67* were scored as follows: 0 for negative; 1 for weak or Mild (yellow or light-brown nuclear stain); 2 for moderate (dark-yellow, medium-brown stain or a mixture of light and dark stain) and 3 for strong (dark-brown stain in most of the nuclei) (10). The extent of staining was classified according to: 0, < 5%; 1, > 5% - 25%; 2, > 25% - 50%; 3, > 50% - 75%; 4, > 75%, and the final score was calculated by multiplying the percentage of positive cells by staining intensity. Generally, the final results were within the range of 0 - 12, scores higher than four were considered positive expression and less than four were considered as negative expression (32).

3.5. Statistical Analysis

Statistical analyses were performed using SPSS ver. 16. Statistical differences between independent groups were assessed by Kruskal-Wallis and Mann-Whitney U tests. All data are presented as the mean \pm standard error of the mean (SEM). $P < 0.05$ were considered significant.

4. Results

4.1. Clinicopathological Data

A total of 122 cases (77 males and 45 females) with the age range of 18 - 92 years were included in this study; 47.54% of the cases had *H. pylori* infection and 52.45% were without *H. pylori* infection. Table 1 shows the demographic profile and *H. pylori* status of the specimens.

4.2. The *p53* and *Ki-67* Expression

The expression of the *p53* and *Ki-67* genes was compared between the samples with and without *H. pylori* infection at different pathological stages (Table 2). Specific staining with both monoclonal anti-*p53* and -*Ki-67* was exclusively confined to the nuclei of the malignant cells. The study data showed that in patients with IM, average *p53* expression (5.70 ± 0.64 for cases with *H. pylori* infection vs. 3.40 ± 0.51 for those without *H. pylori* infection; $P = 0.013 < 0.05$; Table 3); and *Ki-67* gene were significantly higher in specimens with *H. pylori* infection compared to those without *H. pylori* infection (4.55 ± 0.44 vs. 2.95 ± 0.38 ; $P = 0.014 < 0.05$; Table 3). Figure 1A-D shows immunohistochemical

Table 1. Demographic Features of Patients With Gastric Cancer and Precancerous Lesions

Variable	Intestinal Metaplasia	Dysplasia	Gastric Cancer
Age range, y	22 - 81	25 - 86	18 - 92
Gender, No. (%)			
Male	21 (50)	33 (86.8)	23 (54.7)
Female	21 (50)	5 (13.2)	19 (45.2)
Helicobacter pylori infection			
Positive	20 (47.61)	19 (50)	19 (45.23)
Negative	22 (52.38)	19 (50)	23 (54.76)
No. (%)	42 (34.43)	38 (31.15)	42 (34.43)

Table 2. The Expression of *p53* and *Ki-67* in Gastric Cancer and Precancerous Gastric Lesions

Group No. (%)	N	Helicobacter pylori	No. Infection	<i>p53</i> Positive, No. (%)	P Value	<i>Ki-67</i> Positive, No. (%)	P Value
IM	42	+	20	12 (60.0)	0.014	11 (55)	0.005
		-	22	5 (22.72)		3 (13.63)	
DYS	38	+	19	11(57.89)	0.103	5 (26.31)	0.049
		-	19	6 (31.57)		11 (57.89)	
GC	42	+	19	13(68.42)	0.014	14 (73.68)	0.207
		-	23	7(30.43)		14 (60.86)	

Abbreviations: IM, intestinal metaplasia; DYS, dysplasia; GC, gastric cancer.

Table 3. Comparing the Level of *p53* and *Ki-67* Expression in Gastric Cancer and Precancerous Lesions Based on *Helicobacter pylori* Infection

Group No. (%)	N	Helicobacter pylori	No. Infection	<i>p53</i> Positive, (Mean \pm SEM)	P Value	<i>Ki-67</i> Positive, (Mean \pm SEM)	P Value
IM	42	+	20	5.70 \pm 0.64	0.013	4.55 \pm 0.44	0.014
		-	22	3.40 \pm 0.51		2.95 \pm 0.38	
DYS	38	+	19	6.52 \pm 0.60	0.011	3.47 \pm 0.47	0.027
		-	19	4.47 \pm 0.25		5.21 \pm 0.51	
GC	42	+	19	7.89 \pm 0.85	0.001	6.73 \pm 0.60	0.097
		-	23	3.65 \pm 0.59		4.95 \pm 0.73	

Abbreviations: IM, intestinal metaplasia; DYS, dysplasia; GC, gastric cancer; N, number of cases in each group.

staining of *p53* and *Ki-67* positive cells in the groups including specimens with and without *H. pylori* infection.

In DYS, only *p53* expression was significantly higher in the group with *H. pylori* infected compared to that of the group without *H. pylori* infected (6.52 \pm 0.60 vs. 4.47 \pm 0.25; $P = 0.011 < 0.05$; Table 3; Figure 2A and B); while the expression of *Ki-67* in the tissues infected with *H. pylori* was lower than that of the specimens not infected with *H. pylori* (3.47 \pm 0.47 vs. 5.21 \pm 0.51; $P = 0.027 < 0.05$; Table 3; Figure 2C and D).

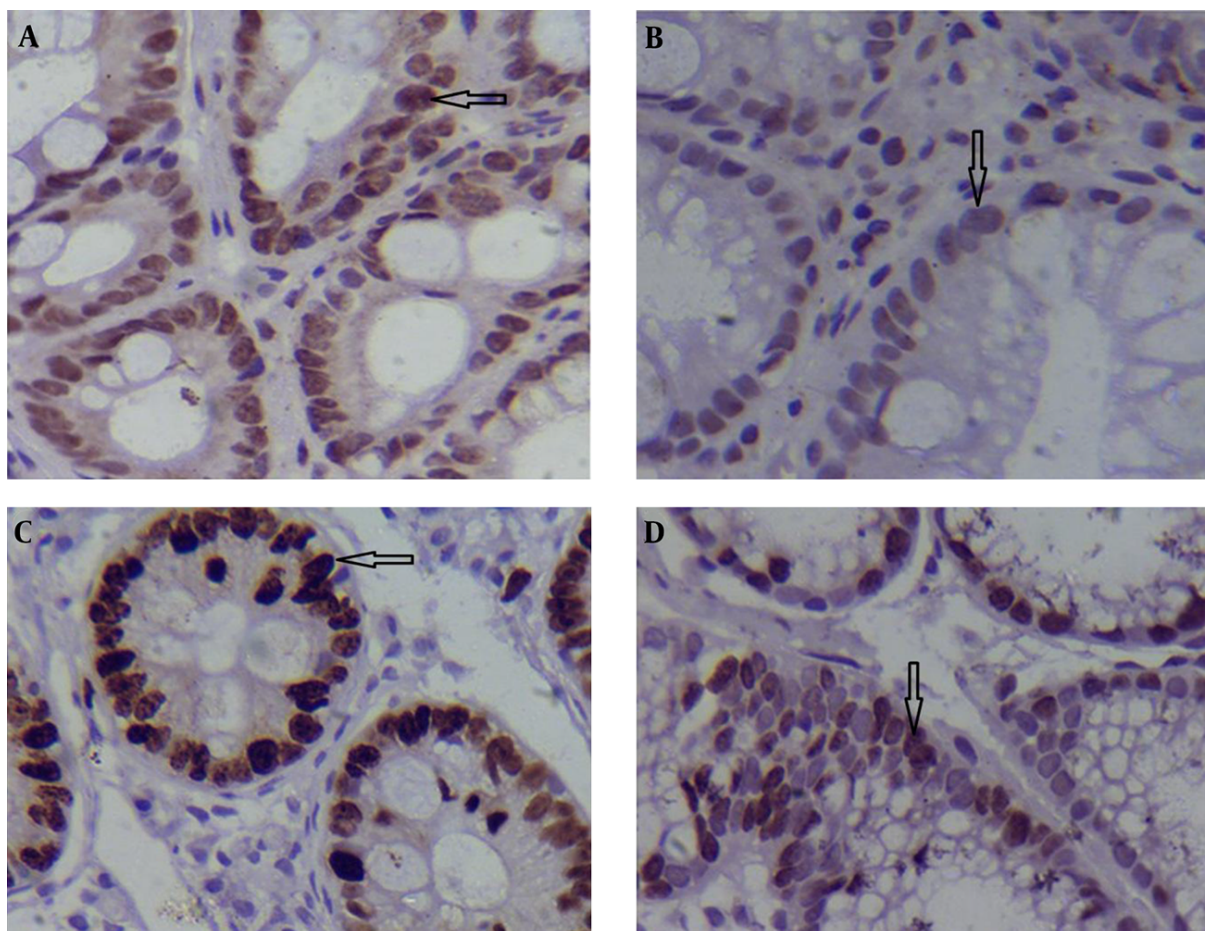
In GC, there was a significant different between the tissues infected and not infected with *H. pylori* regarding the expression of *p53* (7.89 \pm 0.85 vs. 4.13 \pm 0.69; $P = 0.001 < 0.05$ Table 3; Figure 3A and B). There was no significant difference in *Ki-67* expression between the groups with and

without *H. pylori* infection (6.73 \pm 0.60 vs. 4.95 \pm 0.73; $P = 0.097 > 0.05$; Table 3; Figure 3C and D).

In the IM, DYS and GC there was a trend of progressive increase in positivity of *p53* expression in the group with *H. pylori* infection; while *Ki-67* expression in DYS specimens with *H. pylori* infection was lower than that of the IM group. In the group without *H. pylori* infection, from IM toward DYS the expression of the *p53* and *Ki-67* increased but in GC group the intensity of expression of both antibodies decreased (Table 3).

5. Discussion

The current study investigated the *p53* and *Ki-67* expression in the gastric epithelial lesions based on the infection with *H. pylori*. The study results showed that expres-

Figure 1. Immunostaining of Cases With Intestinal Metaplasia

p53 expression in the tissue infected with *H. pylori* (A) and not infected with *H. pylori* (B); Ki-67 expression in cases with *H. pylori* infection (C) and those without *H. pylori* infection (D) (arrows); magnification $\times 400$.

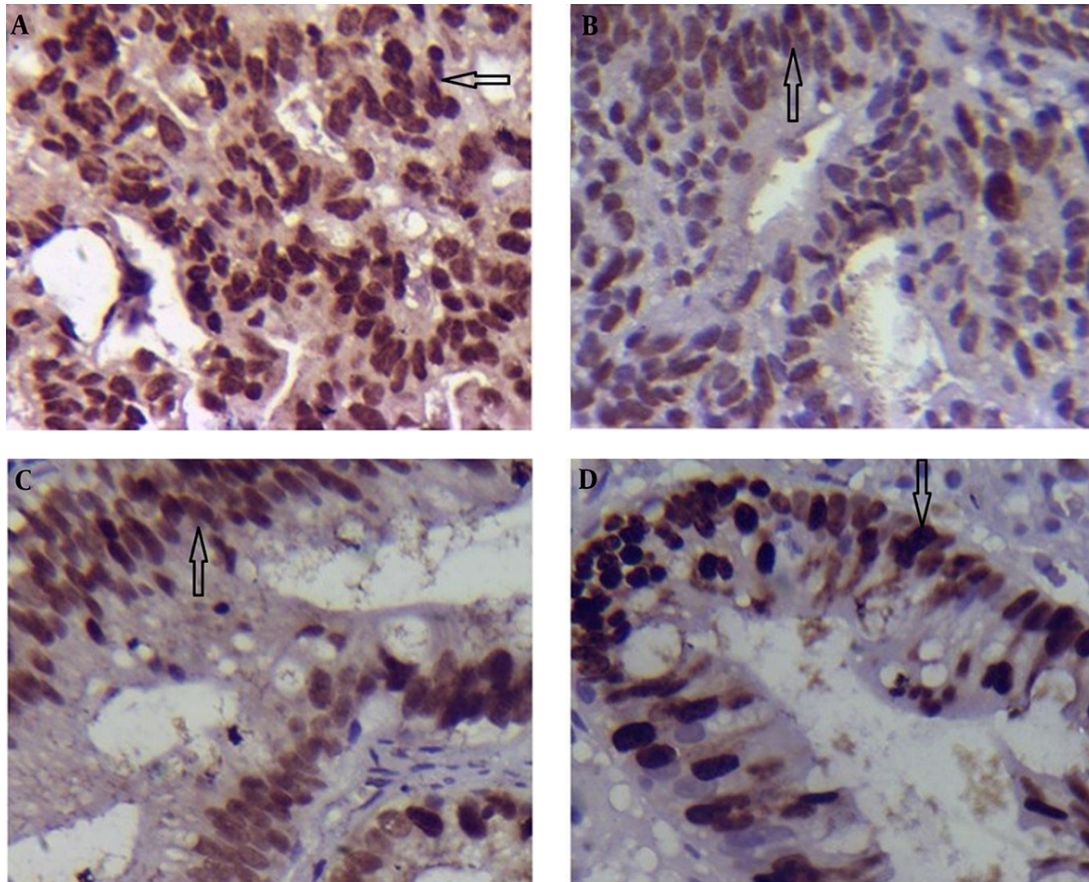
sion of *p53* in the group with *H. pylori* infection was higher than that of the group without *H. pylori* infection (IM, DYS and GC). In addition, it was shown that from the IM to the GC, the *p53* expression progressively increased in the group with *H. pylori* infection. In the current study, *H. pylori* were detected in 47.61% of IM, 50% of DYS and 45.23% of GC specimens. The lower prevalence of *H. pylori* in GC samples compared to that of the precancerous lesions justified as the modified gastric mucosa may become inhospitable for the continued infection with *H. pylori* and its colonization (10, 33).

The study data showed that in the IM group, the *p53* expression in the specimens infected with *H. pylori* was higher than that of the specimens not infected with *H. pylori*. Zhang et al. (34) also showed that in the metaplasia phase, *p53* expression in the group with *H. pylori* infection

was higher than those of the group without *H. pylori* infection and normal mucosa. Similarly, Yang et al. (35) found that in IM, the expression of *p53* in the specimens with *H. pylori* infection was higher than that of the specimens without *H. pylori* infection. In another study by Forones et al. (30), *p53* was negative in all of the IM specimens, and they suggested that mutation of *p53* probably occurs in the latter stages of gastric carcinogenesis.

In the study, in DYS specimens the expression of *p53* in those with *H. pylori* infection was higher than that of specimens without *H. pylori* infection. The findings were similar to the results reported by Zhang et al. (34) that *p53* expression was higher in the specimens with *H. pylori* infection.

The current study showed that, in the GC group, the *p53* expression in the tissues infected with *H. pylori* was higher than that of the tissues not infected with *H. pylori*. In line

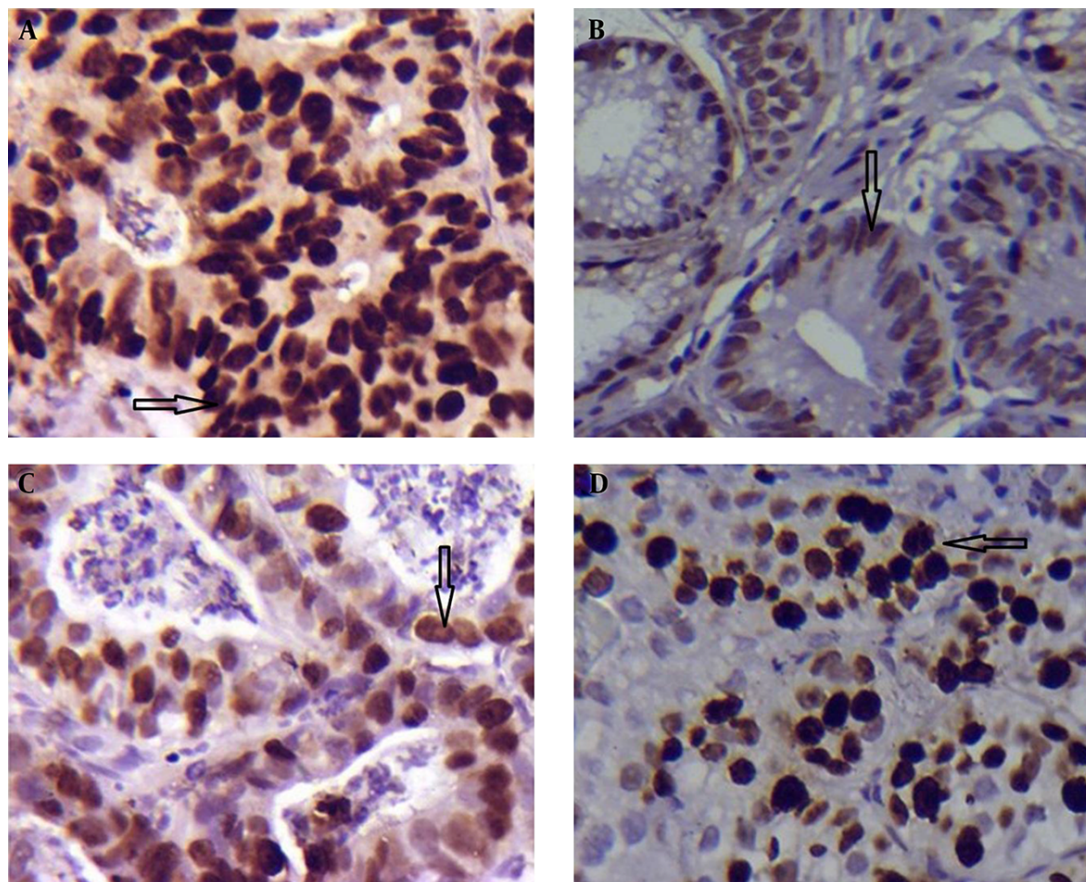
Figure 2. Immunostaining of Cases With Gastric Dysplasia

p53 expression in the cases with *H. pylori* infection (A) and those without *H. pylori* infection (B); *Ki-67* expression in cases with *H. pylori* infection (C) and those without *H. pylori* infection (D) (arrows); magnification $\times 400$.

with the current study, Salih et al. (36) reported an overexpression of *p53* in tumors infected with *H. pylori*. Li et al. (37) also reported that the GC tissue levels of *p53* in the specimens infected with *H. pylori* were higher than those not infected with *H. pylori*. In another study, Zhang et al. (34) reported no expression of *p53* in normal gastric mucosa, but there was an expression of *p53* in GC tissues. Overexpression of *p53* was observed in the specimens infected with *H. pylori* compared to those not infected with *H. pylori*. Accordingly, they concluded that *H. pylori* infection can boost *p53* mutation. Morales-Fuentes et al. (38) reported that in the group with *H. pylori* infection, 91% of cases were positive for *p53* expression while in the group without *H. pylori* infection, only 14% were *p53* positive. Kubicka et al. (39) found more *p53* mutations in patients with positive serology for *H. pylori* compared to the ones with negative test (43% vs. 14%). Yang et al. (35) showed no significant differences in *p53* expression between patients with and without

H. pylori infection in DYS and GC. Berloco et al. (40) also reported that *p53* mutations were not related to *H. pylori* infection in patients with GC in an Italian population. On the other hand, Kodama et al. (41) showed that immunoreactivity for *p53* in *H. pylori* infected group before eradication was significantly higher than that of non-infected group. After eradication of infection, the labeling index for *p53* significantly reduced in both groups. This finding suggests that, the expression of *p53* can be associated with *H. pylori* infection, and it can be concluded that *H. pylori* can be a risk factor for gastric carcinogenesis.

In the current study, the expression of *Ki-67* in IM in the specimens with *H. pylori* infection was significantly higher than that of the specimens not infected with. Similarly, Jang et al. and Cabral et al. (42, 43) found that the expression of *Ki-67* in IM was significantly higher in the group infected with *H. pylori*. This shows an agreement on the effect of *H. pylori* on the expression of *Ki-67* during the precancer-

Figure 3. Immunostaining of Cases With Gastric Cancer

p53 expression in cases with *H. pylori* infection (A) and those without *H. pylori* infection (B); *Ki-67* expression in cases with *H. pylori* infection (C) and those without *H. pylori* infection (D) (arrows); magnification $\times 400$.

ous phases. Some studies reported no statistically significant difference between *Ki-67* expression in the patients with and without *H. pylori* infection and IM (17, 44).

In DYS, expression of *Ki-67* in the group with *H. pylori* infection was lower than that of the group without *H. pylori* infection. In GC, expression of *Ki-67* was slightly higher in the specimens infected with *H. pylori* compared to those not infected with, but this difference was statistically insignificant.

Gucin et al. (14) reported that the level of *Ki-67* expression in GC specimens had a positive relationship with *H. pylori* infection. Petersson et al. (45), in specimens with *H. pylori* infection, detected more than twofold and fourfold overexpression of *Ki-67* and *p53* respectively, compared to the uninfected subjects. Sasaki et al. (46) reported that in GC specimens *Ki-67* labeling index in the group with *H. pylori* infection was higher than that of the group not infected with *H. pylori*. Van De Bovenkamp et al. (47) found

that in the antral area, the number of proliferating cells and *Ki-67* expression were significantly higher in the cases with *H. pylori* infection compared to those not infected with *H. pylori*. De Freitas et al. (48) investigated the effect of *H. pylori* infection on cell proliferation index and apoptotic index in gastric mucosa of the antrum and corpus parts. Their results showed that the expression of *Ki-67* in the antral area significantly increased in the group with *H. pylori* infection, while apoptotic index increased in the gastric corpus area. There was no significant difference in apoptotic index between the groups with and without *H. pylori* infection. In the current study, biopsies were performed on different locations of the stomach; most of them were from antral part. This, as a limiting point, may affect the results. In patients with GC, there was no difference regarding *Ki-67* expression between the groups with and without *H. pylori* infection. In this phase, *Ki-67* overexpression is observed as a result of malignancy occurrence

and this phase can be independent from *H. pylori* infection.

Since in IM phase the expression of *Ki-67* was higher in the cases infected with *H. pylori*, it can be concluded that the presence of bacteria is essential for the onset of the disease (36). As observed in DYS, the expression of *Ki-67* in the group with *H. pylori* infection was lower than that of the group without infection, and despite the fact that in GC there were no significant differences in the *Ki-67* expression between the cases infected and not infected with *H. pylori*, it can be suggested that when the disease progresses from IM toward DYS and GC, the expression of *Ki-67* is affected by genetic changes and molecular events that take place at the earlier stages of carcinogenesis and these changes can be caused by the colonization of *H. pylori* (38). The most important limitation of the study was insufficient biopsy specimens in group dysplasia, lack of some paraffin blocks and atrophy of some samples. Future studies are required to take larger sample sizes and samples from the lesions in the same locations of the stomach to clarify the differences observed between the current study and those of the previous researches. The current study showed a strong association between *H. pylori* infection and intensity of *p53* expression. It suggests that *H. pylori* can be a risk factor to develop gastric cancer. Also, an association between *H. pylori* infection and overexpression of *Ki-67* was observed in earlier stages of carcinogenesis. Further studies should be conducted on this matter.

Acknowledgments

This research was approved by ethics committee of Zahedan University of Medical Sciences (IR.ZAUMS.REC.1394.53). The authors appreciate the vice chancellor of research and technology of Zahedan University of Medical Sciences that financially supported the research, and also the pathology department of Ali-ebne-Abitaleb hospital, Zahedan, Iran.

Footnotes

Authors' Contribution: Hamidreza Mahmoudzadeh Sagheb and Zahra Heidari: conceive and design of the study, supervising all the experimental designs, results analysis and interpretation, modification and improvement of final version of the manuscript; Mehdi Jahantigh: reviewing the original sections to confirm the histopathological diagnosis; Mahdieh Narouei: study design, data analysis and manuscript drafting

Funding/Support: The current study was financially supported by the vice chancellor of research and technology of Zahedan University of Medical Sciences (IR.ZAUMS.REC.1394.53).

References

- Al-Moundhri MS, Nirmala V, Al-Hadabi I, Al-Mawaly K, Burney I, Al-Nabhani M, et al. The prognostic significance of p53, p27 kipt1, p21 waf1, HER-2/neu, and Ki67 proteins expression in gastric cancer: a clinicopathological and immunohistochemical study of 121 Arab patients. *J Surg Oncol*. 2005;**91**(4):243–52. doi: [10.1002/jso.20324](https://doi.org/10.1002/jso.20324). [PubMed: [16121348](https://pubmed.ncbi.nlm.nih.gov/16121348/)].
- Poteca T, Poteca A, Sajin M, Comanescu M. Biological prognostic parameters in gastric carcinomas. *Chirurgia (Bucur)*. 2014;**109**(3):347–54. [PubMed: [24956340](https://pubmed.ncbi.nlm.nih.gov/24956340/)].
- Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. *CA Cancer J Clin*. 2011;**61**(2):69–90. doi: [10.3322/caac.20107](https://doi.org/10.3322/caac.20107). [PubMed: [21296855](https://pubmed.ncbi.nlm.nih.gov/21296855/)].
- Pinto-de-Sousa J, Silva F, David L, Leitao D, Seixas M, Pimenta A, et al. Clinicopathological significance and survival influence of p53 protein expression in gastric carcinoma. *Histopathology*. 2004;**44**(4):323–31. doi: [10.1111/j.1365-2559.2004.01852.x](https://doi.org/10.1111/j.1365-2559.2004.01852.x). [PubMed: [15049897](https://pubmed.ncbi.nlm.nih.gov/15049897/)].
- Matysiak-Budnik T, Megraud F. Helicobacter pylori infection and gastric cancer. *Eur J Cancer*. 2006;**42**(6):708–16. doi: [10.1016/j.ejca.2006.01.020](https://doi.org/10.1016/j.ejca.2006.01.020). [PubMed: [16556496](https://pubmed.ncbi.nlm.nih.gov/16556496/)].
- Mashhadi M, Nazem K, Abdollahinejad MJ. Gastric Cancer in South East of Iran. *Int J Hematol Oncol Stem Cell Res*. 2009;**3**(4):38–42.
- Howson CP, Hiyama T, Wynder EL. The decline in gastric cancer: epidemiology of an unplanned triumph. *Epidemiol Rev*. 1986;**8**:1–27. [PubMed: [3533579](https://pubmed.ncbi.nlm.nih.gov/3533579/)].
- Kulmambetova GN, Imanbekova MK, Logvinenko AA, Sukashev AT, Filipenko ML, Ramanculov EM. Association of cytokine gene polymorphisms with gastritis in a Kazakh population. *Asian Pac J Cancer Prev*. 2014;**15**(18):7763–8. [PubMed: [25292060](https://pubmed.ncbi.nlm.nih.gov/25292060/)].
- Strehl JD, Hoegel J, Hornicek I, Hartmann A, Riener MO. Immunohistochemical expression of IMP3 and p53 in inflammatory lesions and neoplastic lesions of the gastric mucosa. *Int J Clin Exp Pathol*. 2014;**7**(5):2091–101. [PubMed: [24966917](https://pubmed.ncbi.nlm.nih.gov/24966917/)].
- Zhang C, Yamada N, Wu YL, Wen M, Matsuhisa T, Matsukura N. Helicobacter pylori infection, glandular atrophy and intestinal metaplasia in superficial gastritis, gastric erosion, erosive gastritis, gastric ulcer and early gastric cancer. *World J Gastroenterol*. 2005;**11**(6):791–6. [PubMed: [15682469](https://pubmed.ncbi.nlm.nih.gov/15682469/)].
- Shadifar M, Ataee R, Ataie A, Heydari Gorgi AM, Nasri Nasrabadi N, Nouri S. Genetic and molecular aspects of Helicobacter pylori in gastritis, pre-cancerous conditions and gastric adenocarcinoma. *Gastroenterol Hepatol Bed Bench*. 2015;**8**(Suppl1):15–22. [PubMed: [26171133](https://pubmed.ncbi.nlm.nih.gov/26171133/)].
- Wroblewski LE, Peek RM, Wilson KT. Helicobacter pylori and gastric cancer: factors that modulate disease risk. *Clin Microbiol Rev*. 2010;**23**(4):713–39. doi: [10.1128/CMR.00011-10](https://doi.org/10.1128/CMR.00011-10). [PubMed: [20930071](https://pubmed.ncbi.nlm.nih.gov/20930071/)].
- Hegazi A, Hassan E, El-Atrebi KA, El-Bassyouni HT. P53 protein and Ki-67 expression in chronic gastritis patients with positive Helicobacter pylori infection. *J Genetic Engin Biotechnol*. 2011;**9**(1):73–6.
- Gucin Z, Cakmak T, Bayyurt N, Salih B. Helicobacter pylori infection and relationship with gastric epithelial cell proliferation and apoptosis. *Turk J Med Sci*. 2013;**43**:739–46. doi: [10.3906/sag-1207-12](https://doi.org/10.3906/sag-1207-12).
- Kayacetin S, Guresci S. What is gastritis? What is gastropathy? How is it classified?. *Turk J Gastroenterol*. 2014;**25**(3):233–47. doi: [10.5152/tjg.2014.7906](https://doi.org/10.5152/tjg.2014.7906). [PubMed: [25141310](https://pubmed.ncbi.nlm.nih.gov/25141310/)].
- Zheng Y, Wang L, Zhang JP, Yang JY, Zhao ZM, Zhang XY. Expression of p53, c-erbB-2 and Ki67 in intestinal metaplasia and gastric carcinoma. *World J Gastroenterol*. 2010;**16**(3):339–44. [PubMed: [20082479](https://pubmed.ncbi.nlm.nih.gov/20082479/)].
- Erkan G, Gonul I, Kandilci U, Dursun A. Evaluation of apoptosis along with BCL-2 and Ki-67 expression in patients with intestinal metaplasia. *Pathol Res Pract*. 2012;**208**(2):89–93. doi: [10.1016/j.prp.2011.12.002](https://doi.org/10.1016/j.prp.2011.12.002). [PubMed: [22277792](https://pubmed.ncbi.nlm.nih.gov/22277792/)].

18. Zullo A, Hassan C, Romiti A, Giusto M, Guerriero C, Lorenzetti R, et al. Follow-up of intestinal metaplasia in the stomach: When, how and why. *World J Gastrointest Oncol.* 2012;**4**(3):30–6. doi: [10.4251/wjgo.v4.i3.30](https://doi.org/10.4251/wjgo.v4.i3.30). [PubMed: [22468181](https://pubmed.ncbi.nlm.nih.gov/22468181/)].
19. Shin N, Jo HJ, Kim WK, Park WY, Lee JH, Shin DH, et al. Gastric pit dysplasia in adjacent gastric mucosa in 414 gastric cancers: prevalence and characteristics. *Am J Surg Pathol.* 2011;**35**(7):1021–9. doi: [10.1097/PAS.0b013e31821ec495](https://doi.org/10.1097/PAS.0b013e31821ec495). [PubMed: [21677540](https://pubmed.ncbi.nlm.nih.gov/21677540/)].
20. Soussi T, Wiman KG. Shaping genetic alterations in human cancer: the p53 mutation paradigm. *Cancer Cell.* 2007;**12**(4):303–12. doi: [10.1016/j.ccr.2007.10.001](https://doi.org/10.1016/j.ccr.2007.10.001). [PubMed: [17936556](https://pubmed.ncbi.nlm.nih.gov/17936556/)].
21. Heidari Z, Mahmoudzadeh Sagheb H, Asemi Rad A, Keikhaee MA. Immunocytochemical Study of p53 Protein in Exfoliated Cells of Oral Mucosa in Patients With Type 2 Diabetes. *Gene Cell Tissue.* 2015;**2**(1).
22. Levine AJ. p53, the cellular gatekeeper for growth and division. *Cell.* 1997;**88**(3):323–31. [PubMed: [9039259](https://pubmed.ncbi.nlm.nih.gov/9039259/)].
23. Greenblatt MS, Bennett WP, Hollstein M, Harris CC. Mutations in the p53 tumor suppressor gene: clues to cancer etiology and molecular pathogenesis. *Cancer Res.* 1994;**54**(18):4855–78. [PubMed: [8069852](https://pubmed.ncbi.nlm.nih.gov/8069852/)].
24. Hall PA, Lane DP. p53 in tumour pathology: can we trust immunohistochemistry?—Revisited!. *J Pathol.* 1994;**172**(1):1–4. doi: [10.1002/path.1711720103](https://doi.org/10.1002/path.1711720103). [PubMed: [7931821](https://pubmed.ncbi.nlm.nih.gov/7931821/)].
25. Soussi T. p53 alterations in human cancer: more questions than answers. *Oncogene.* 2007;**26**(15):2145–56. doi: [10.1038/sj.onc.1210280](https://doi.org/10.1038/sj.onc.1210280). [PubMed: [17401423](https://pubmed.ncbi.nlm.nih.gov/17401423/)].
26. Yamada Y, Yoshida T, Hayashi K, Sekiya T, Yokota J, Hirohashi S, et al. p53 gene mutations in gastric cancer metastases and in gastric cancer cell lines derived from metastases. *Cancer Res.* 1991;**51**(21):5800–5. [PubMed: [1933850](https://pubmed.ncbi.nlm.nih.gov/1933850/)].
27. Inwald EC, Klinkhammer-Schalke M, Hofstadter F, Zeman F, Koller M, Gerstenhauer M, et al. Ki-67 is a prognostic parameter in breast cancer patients: results of a large population-based cohort of a cancer registry. *Breast Cancer Res Treat.* 2013;**139**(2):539–52. doi: [10.1007/s10549-013-2560-8](https://doi.org/10.1007/s10549-013-2560-8). [PubMed: [23674192](https://pubmed.ncbi.nlm.nih.gov/23674192/)].
28. Niimi C, Goto H, Ohmiya N, Niwa Y, Hayakawa T, Nagasaka T, et al. Usefulness of p53 and Ki-67 immunohistochemical analysis for preoperative diagnosis of extremely well-differentiated gastric adenocarcinoma. *Am J Clin Pathol.* 2002;**118**(5):683–92. doi: [10.1309/NYAI-V9KQ-NVF8-MA8M](https://doi.org/10.1309/NYAI-V9KQ-NVF8-MA8M). [PubMed: [12428787](https://pubmed.ncbi.nlm.nih.gov/12428787/)].
29. Lazar D, Taban S, Sporea I, Dema A, Cornianu M, Lazar E, et al. Ki-67 expression in gastric cancer. Results from a prospective study with long-term follow-up. *Rom J Morphol Embryol.* 2010;**51**(4):655–61. [PubMed: [21103622](https://pubmed.ncbi.nlm.nih.gov/21103622/)].
30. Forones NM, Carvalho AP, Giannotti-Filho O, Lourenco LG, Oshima CT. Cell proliferation and apoptosis in gastric cancer and intestinal metaplasia. *Arq Gastroenterol.* 2005;**42**(1):30–4. [PubMed: [15976908](https://pubmed.ncbi.nlm.nih.gov/15976908/)].
31. Sanaat Z, Halimi M, Ghojzadeh M, Pirovi AH, Gharamaleki JV, Ziae AE, et al. Immunohistochemical Analysis of p53, Ki-67, CD44, HER-2/neu Expression Patterns in Gastric Cancer, and Their Association with One Year Survival in North-West of Iran. *Int J Hematol Oncol Stem Cell Res.* 2013;**7**(3):15–20. [PubMed: [24505530](https://pubmed.ncbi.nlm.nih.gov/24505530/)].
32. Tiwari E, Pallipady A, Misra R, Mishra S. P53 and ki67 Immunostaining in Gastric Biopsies: A Histopathological Study. *Int J Sci Stud.* 2(11):96–101.
33. Chatzipantelis P, Konstantinou P, Voros D, Smyrniotis V, Kondi-Pafiti A. Apoptosis and gastric carcinogenesis: Immunohistochemical analysis of Bax and Bcl-2 proteins. *Annals of Gastroenterology.* 2007;**20**(2).
34. Zhang Z, Yuan Y, Gao H, Dong M, Wang L, Gong YH. Apoptosis, proliferation and p53 gene expression of H. pylori associated gastric epithelial lesions. *World J Gastroenterol.* 2001;**7**(6):779–82. [PubMed: [11854900](https://pubmed.ncbi.nlm.nih.gov/11854900/)].
35. Yang Z, Shu X, Chen L, Chen J, Xie Y, Lu NH. Expression of p53-MDM2 feedback loop related proteins in different gastric pathologies in relation to Helicobacter pylori infection: implications in gastric carcinogenesis. *Clin Res Hepatol Gastroenterol.* 2012;**36**(3):235–43. doi: [10.1016/j.clinre.2011.11.009](https://doi.org/10.1016/j.clinre.2011.11.009). [PubMed: [22306053](https://pubmed.ncbi.nlm.nih.gov/22306053/)].
36. Salih BA, Gucin Z, Bayyurt N. A study on the effect of Helicobacter pylori infection on p53 expression in gastric cancer and gastritis tissues. *J Infect Dev Ctries.* 2013;**7**(9):651–7. doi: [10.3855/jidc.2993](https://doi.org/10.3855/jidc.2993). [PubMed: [24042100](https://pubmed.ncbi.nlm.nih.gov/24042100/)].
37. Li JH, Shi XZ, Lv S, Liu M, Xu GW. Effect of Helicobacter pylori infection on p53 expression of gastric mucosa and adenocarcinoma with microsatellite instability. *World J Gastroenterol.* 2005;**11**(28):4363–6. [PubMed: [16038035](https://pubmed.ncbi.nlm.nih.gov/16038035/)].
38. Morales-Fuentes GA, Zarate-Osorno A, Quinonez-Urrego EE, Antonio-Manrique M, Martinez-Garcia CL, Figueroa-Barojas P, et al. [p53 expression in the gastric mucosa of patients infected with Helicobacter pylori]. *Rev Gastroenterol Mex.* 2013;**78**(1):12–20. doi: [10.1016/j.rgm.2012.11.001](https://doi.org/10.1016/j.rgm.2012.11.001). [PubMed: [23374541](https://pubmed.ncbi.nlm.nih.gov/23374541/)].
39. Kubicka S, Claas C, Staab S, Kuhnle F, Zender I, Trautwein C, et al. p53 mutation pattern and expression of c-erbB2 and c-met in gastric cancer: relation to histological subtypes, Helicobacter pylori infection, and prognosis. *Dig Dis Sci.* 2002;**47**(1):14–21. [PubMed: [11837710](https://pubmed.ncbi.nlm.nih.gov/11837710/)].
40. Berloco P, Russo F, Cariola F, Gentile M, Giorgio P, Caruso ML, et al. Low presence of p53 abnormalities in H pylori-infected gastric mucosa and in gastric adenocarcinoma. *J Gastroenterol.* 2003;**38**(1):28–36. doi: [10.1007/s005350300003](https://doi.org/10.1007/s005350300003). [PubMed: [12560919](https://pubmed.ncbi.nlm.nih.gov/12560919/)].
41. Kodama M, Fujioka T, Murakami K, Okimoto T, Sato R, Watanabe K, et al. Eradication of Helicobacter pylori reduced the immunohistochemical detection of p53 and MDM2 in gastric mucosa. *J Gastroenterol Hepatol.* 2005;**20**(6):941–6. doi: [10.1111/j.1440-1746.2005.03880.x](https://doi.org/10.1111/j.1440-1746.2005.03880.x). [PubMed: [15946145](https://pubmed.ncbi.nlm.nih.gov/15946145/)].
42. Jang TJ, Kim JR. Proliferation and apoptosis in gastric antral epithelial cells of patients infected with Helicobacter pylori. *J Gastroenterol.* 2000;**35**(4):265–71. [PubMed: [10777155](https://pubmed.ncbi.nlm.nih.gov/10777155/)].
43. Cabral MM, Oliveira CA, Mendes CM, Guerra J, Queiroz DM, Rocha GA, et al. Gastric epithelial cell proliferation and cagA status in Helicobacter pylori gastritis at different gastric sites. *Scand J Gastroenterol.* 2007;**42**(5):545–54. doi: [10.1080/00365520601014034](https://doi.org/10.1080/00365520601014034). [PubMed: [17454874](https://pubmed.ncbi.nlm.nih.gov/17454874/)].
44. Hoshi T, Sasano H, Kato K, Ohara S, Shimosegawa T, Toyota T, et al. Cell damage and proliferation in human gastric mucosa infected by Helicobacter pylori—a comparison before and after H pylori eradication in non-atrophic gastritis. *Hum Pathol.* 1999;**30**(12):1412–7. [PubMed: [10667417](https://pubmed.ncbi.nlm.nih.gov/10667417/)].
45. Petersson F, Borch K, Franzen LE. Gastric epithelial proliferation and p53 and p21 expression in a general population sample: relations to age, sex, and mucosal changes associated with H. pylori infection. *Dig Dis Sci.* 2002;**47**(7):1558–66. [PubMed: [12141817](https://pubmed.ncbi.nlm.nih.gov/12141817/)].
46. Sasaki A, Kitadai Y, Ito M, Sumii M, Tanaka S, Yoshihara M, et al. Helicobacter pylori infection influences tumor growth of human gastric carcinomas. *Scand J Gastroenterol.* 2003;**38**(2):153–8. [PubMed: [12678331](https://pubmed.ncbi.nlm.nih.gov/12678331/)].
47. Van De Bovenkamp JH, Korteland-Van Male AM, Buller HA, Einerhand AW, Dekker J. Infection with Helicobacter pylori affects all major secretory cell populations in the human antrum. *Dig Dis Sci.* 2005;**50**(6):1078–86. [PubMed: [15986858](https://pubmed.ncbi.nlm.nih.gov/15986858/)].
48. de Freitas D, Urbano M, Goulao MH, Donato MM, Baldaia C, Martins MI, et al. The effect of Helicobacter pylori infection on apoptosis and cell proliferation in gastric epithelium. *Hepatogastroenterology.* 2004;**51**(57):876–82. [PubMed: [15143938](https://pubmed.ncbi.nlm.nih.gov/15143938/)].