



CAR-NK Cells as Promising Immune Therapeutics: Platforms and Current Progress

Maryam Samareh Salavatipour¹, Shirin Tavakoli¹, Shima Tavoosi², Masoume Nodehi ¹, Amir Hossein Baghsheikhi³, Mohammad Vaezi ⁴, Javad Verdi¹, Soheila Rahgozar⁵, Maryam Barkhordar^{4, **}, Mohammad Ahmadvand^{4, *}

¹ Department of Applied Cell Sciences, School of Advanced Technologies in Medicine, Tehran University of Medical Sciences, Tehran, Iran

² Department of Biology, Faculty of Sciences, University of Isfahan, Hezar Jerib Ave, Azadi, Iran

³ Department of Biology, Science and Research Branch, Islamic Azad University, Tehran, Iran

⁴ Cell Therapy and Hematopoietic Stem Cell Transplantation Research Center, Research Institute for Oncology, Hematology and Cell Therapy, Tehran University of Medical Sciences, Tehran, Iran

⁵ Department of Cell and Molecular Biology & Microbiology, Faculty of Biological Science and Technology, University of Isfahan, Hezar Jarib Street, Isfahan 81746-73441, Iran

* Corresponding Author: Cell Therapy and Hematopoietic Stem Cell Transplantation Research Center, Research Institute for Oncology, Hematology and Cell Therapy, Tehran University of Medical Sciences, Tehran, Iran. Email: ahmadvand.mohamad64@yahoo.com

** Corresponding Author: Cell Therapy and Hematopoietic Stem Cell Transplantation Research Center, Research Institute for Oncology, Hematology and Cell Therapy, Tehran University of Medical Sciences, Tehran, Iran. Email: barkhordarm.n@gmail.com

Received 2024 January 27; **Revised** 2024 February 19; **Accepted** 2024 March 17.

Abstract

Context: Adoptive T-cell therapy with chimeric antigen receptor (CAR) has shown tremendous progress in hematological cancers. However, some obstacles, such as high price tag, cytokine release syndrome, inability to penetrate solid tumors, and manufacturing complexity limit the wide application of this therapy. Natural killer (NK) cells can kill target cells via mechanisms similar to those of CD8+ cytotoxic T cells; therefore, CAR-NK cell therapy is a promising strategy for cancer treatment.

Evidence Acquisition: In this manuscript, all articles published in English regarding CAR-NKs and their application for the treatment of different types of cancers were collected from several databases, including PubMed, Scopus, and Google Scholar, using related keywords such as "Cancer, CAR construction, NK cells, and CAR-NK cells".

Results: Compared with CAR-T cells, CAR-NK cells have several advantages, including less toxicity, a high potential opportunity for universal off-the-shelf manufacturing, increased infiltration into solid tumors, overcoming resistant tumor microenvironment, and absence of graft-versus-host disease (GVHD).

Conclusions: In this review, we discuss NK cell biology, the source of CAR-NK cells, CAR structure, advances, challenges, and ways to overcome these challenges in CAR-NK cell therapy. Furthermore, we have summarized and highlighted some preclinical and clinical studies in this field.

Keywords: Chimeric Antigen Receptor, NK Cells, Immunotherapy, Cancer

1. Context

1.1. Natural Killer Cells: Phenotype, Biology, and Receptors

Natural killer (NK) cells, which are a critical part of the innate immune system, show intense cytotoxic responses against virus-infected cells and altered self-cells such as tumor cells without prior sensitization. For the first time, NK cells have been observed in human

peripheral blood (1) and rodent splenocytes (2), whereas additional populations of NK cells have been identified to reside in multiple lymphoid and non-lymphoid tissues, including the lymph nodes (LNs), bone marrow (BM), gut, tonsils, skin, liver, and lungs (3), where they play specialized immune functions.

Natural Killer cells represent 5% to 20% of all circulating lymphocytes in humans (4), which are characterized by the expression of neural cell adhesion

molecules (NCAM or CD56) and a lack of CD3 surface antigens.

Unlike T cells, NK cells lack an antigen-specific T-cell receptor (TCR); therefore, the cytotoxic or regulatory phenotypes of NK cells are determined by the expression of various activating and inhibiting receptors and interaction with their related ligands (5). The intracellular domain of activating receptors such as NKG2A, NCR (NKP44, NKP46, NKP30), immunoglobulin-like receptors (KIR), and CD16 contains an immunoreceptor tyrosine-based activation motif (ITAM) that is phosphorylated after interaction with their ligands, resulting in activating signal transduction.

Unlike T cells, NK cells lack an antigen-specific TCR; therefore, the cytotoxic or regulatory phenotypes of NK cells are determined by the expression of various activating and inhibiting receptors and their interaction with their related ligands. Activating receptors play a crucial role in controlling NK cell function. The intracellular domain of these receptors contains an ITAM that is phosphorylated after the interaction of receptor-ligand complexes and results in activating signal transduction (6). Typical activating receptors include NKG2D and natural cytotoxicity receptors (NCR: NKP44, NKP46, NKP30) KIR, and CD16.

In contrast, NK cells are negatively regulated by inhibitory receptors that present immunoreceptor tyrosine-based inhibitory motifs (ITIM) in their cytoplasmic tails. NK cells contain two types of inhibitory receptors: HLA-specific and non-HLA-specific receptors (PD-1, SIGLEC7, and IRP60).

In addition to activating KIRs, different kinds of KIR family receptors are known to induce inhibitory feedback upon interaction with class I specific HLA molecules such as HLA-A, HLA-B, and HLA-C, whereas NKG2A recognizes non-classical HLA-E molecules. According to the initial observation of Ljunggren and Karre et al., NK cells target cells lacking HLA-I expression, whereas HLA-I-sufficient cells are spared (7). Such ligands are expressed in all nucleated cells that prevent NK cell stimulation, whereas due to the missing-self recognition phenomenon, downregulation of HLA-I on tumor cells leads to increased targeting by NK cells.

1.2. Natural Killer Cell-Based Cancer Immunotherapy

Cancer is the leading cause of death and a major public health problem that is increasing all over the world (8). The most common treatments are surgery, chemotherapy, and radiation, which have difficulty completely eradicating cancer cells. Inspired by the natural capacity of our immune system to recognize

and prevent tumor progression, emerging immunotherapy-based accessory immune cells such as T and NK cells are revolutionizing the clinical management of multiple tumors (8). There are several key advantages to using NK cells in immunotherapy. First, NK cells do not require a specific antigen expressed on a particular HLA allotype, making them antigens non-specific. Instead, they can recognize numerous ligands that can trigger a cytolytic response. Second, NK cells can be readily isolated and expanded outside the body, allowing their use in adoptive or autologous cell therapy. Third, NK cells have a shorter lifespan than T cells, eliminating the need for a suicide vector to prevent excessive expansion of the transferred cells (1).

Recently, T-cell transfer therapy, especially chimeric antigen receptor (CAR) T-cell therapy, as a type of immunotherapy, has become a central focus to boost the immune system's ability to fight tumor cells. While CAR-T cell therapy has shown potential clinical benefits in cancer treatment, a major hurdle to CAR-T cell therapy, such as cytokine release syndrome (CRS), neurotoxicity, on-target/off-tumor toxicity, tumor lysis syndrome (TLS), and anaphylaxis has also been reported. As an alternative to CAR-T cell therapy, CAR-NK cells not only overcome these limitations but also present additional major advantages (9).

1.3. Chimeric Antigen Receptor

Chimeric Antigen Receptor (CARs) are genetically engineered receptors that re-direct NK cells with a high ability to target certain proteins in cancer cells. For example, Her2 and CD19 are the most expressed targets for breast and B cell malignancies and are widely targeted by CAR-NK cells.

As represented in Figure 1, functional CAR molecules consist of 3 regions: Extracellular, transmembrane, and intracellular domains. The extracellular antigen-binding domain is made up of the single chain fragment variant (SCFV) with a linker between the heavy and light chains, as well as the hinge region such as CD8, CD28, DAP12, IgG1, and IgG4 that causes CAR stability and flexibility to easily access the target. Extracellular and intracellular regions join each other by a transmembrane zone that transduces activation signals to intracellular activation signals.

As represented in Figure 1, functional CAR molecules consist of 3 regions: Extracellular, transmembrane, and intracellular domains. The extracellular antigen-binding domain is made up of the single chain fragment variant (SCFV) with a linker between the heavy and light chains, as well as the hinge region such as CD8, CD28, DAP12, IgG1, and IgG4 that causes CAR stability and

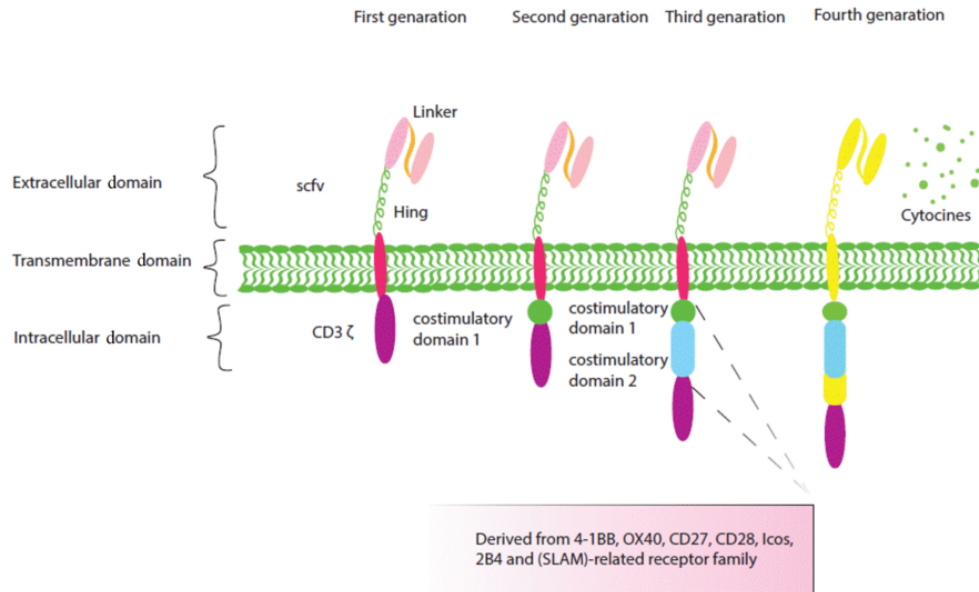


Figure 1. Structure of the first, second, and fourth generations of chimeric antigen receptors (CARs). Chimeric antigen receptors are composed of an extracellular antigen-binding domain, a transmembrane domain, and an intracellular domain. The extracellular region contains a chain fragment of the antibody variable region (ScFv) or a functional domain of a specific ligand. The intracellular region consists of a signaling domain (first generation) and a stimulatory domain (second generation) or two (third generation). Fourth-generation CARs are designed to express molecules such as cytokines.

flexibility to easily access the target. Extracellular and intracellular regions join each other by a transmembrane zone that transduces activation signals to intracellular activation signals.

The intracellular region, which mainly mediates CAR-NK cell cytotoxicity, includes activation signaling domains, and CAR generation is determined by the number of these domains (10). CARs are identified by their different generation. The first generation of CARs has only one activating signal; for example, the first CAR-NK cell generation that overcomes leukemia resistance consists of only a CD3 zeta(ζ) signaling moiety (11).

1.4. Different Sources of NK Cells for CAR-NK

As displayed in Figure 2, allogeneic NK cells for immunotherapy can be derived from different sources, such as NK cell lines, induced pluripotent stem cells (iPSCs), cord blood (CB), hematopoietic stem cells (HSCs), and peripheral blood (PB). Obtaining large numbers of NK cells and being time-consuming are the common limitations of NK cell sources while cell lines overcome these problems (12).

Many NK cells, including KHYG-1, NK92, NKL, NKG, and YT cells with notable advancement have been

studied.

1.5. NK92

The NK92 cell line not only shows high anti-tumor activity by direct cytotoxicity but also could be an ideal CAR carrier because of its easy modification. The first CAR-NK92 cells targeting HER2 were reported by Uherek et al. Other studies also confirmed the efficacy and safety of CAR-based NK92 cells in preclinical and clinical trials by targeting hematological malignancies such as acute myeloid leukemia (CD33), lymphoma (CD19), myeloma (CSI), and other solid tumors such as glioblastoma (EGFR), ovarian cancer (mesothelin), and prostate cancer (EPCAM) (13).

1.6. Peripheral Blood

The most available source of NK cells is peripheral blood, which can be easily isolated from healthy donors (allogeneic PB-NK) or from the patient itself (autologous PB-NK). Using autologous NK cells is typically not impressive, as they are functionally inhibited after encountering self-MHC antigens; therefore, for immunotherapy purposes, allogeneic PB-NK cells could be more effective (14). Furthermore, because of the low

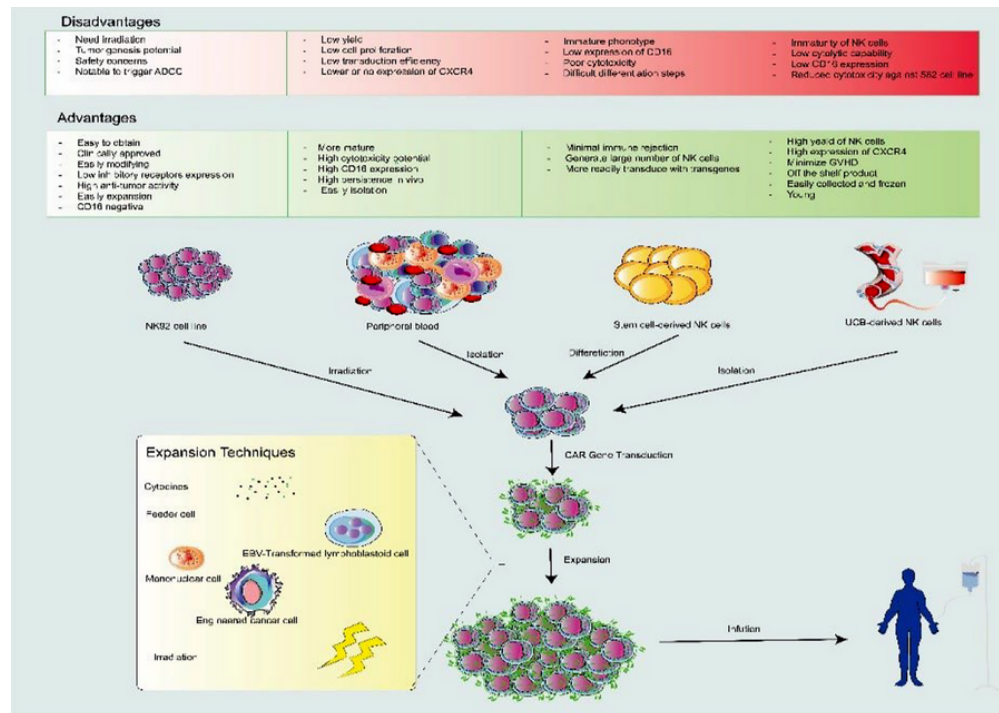


Figure 2. Sources and generation procedure of CAR-NK cells. Isolated or established natural killer (NK) cells from various sources (e.g., PB, UCB, NK cell line, and Stem cell-derived NK cells) can be activated and genetically modified with CAR-expressing vectors (e.g., lentivirus or retrovirus) and, then, expanded by some techniques (e.g., using cytokines, irradiation, feeder cells such as mononuclear cell, EBV transformed lymphoblastoid cells or engineered cancer cell). UCB, umbilical cord blood; PB, peripheral blood; CAR-NK, chimeric antigen receptor-natural killer cells.

number of NK cells after isolation, they are routinely expanded by various expansion protocols, which is another difficulty because prolonged culture causes telomerase shortening and subsequently reduced cytotoxicity (15).

1.7. Umbilical Cord Blood-Derived NK Cells (UCB-Derived NK Cells)

UCB-derived NK cells as a potential "off the shelf" product have recently been noticed (16). The NK cells in UCB can be easily collected and frozen with high proliferation potential and number compared to PB-isolated NK cells. For instance, only 10% of one cord blood unit is sufficient to generate more than 109 NK cells after 2 weeks (17). In addition, according to a study, more than 100 doses of NK cells were collected just from a single UCB unit for clinical usage. Although a minority of the preclinical studies applied UCB as an NK cell source, the safety and potency of CB-NK cells were demonstrated in UCB-NK trials (12). A recent phase I/II study showed that CAR-NK targeting CD19-expressing

malignancies can persist for more than 270 days in vivo. This group also demonstrated that most 11 patients with relapsed/refractory CD19-positive cancer reacted CAR-NK without any toxic effects (18).

1.8. Stem Cell-Derived NK cells

The other potential source for CAR-NK generation is pluripotent stem cells, including induced pluripotent stem cells (iPSCs) and human embryonic stem cells (hESCs), which have emerged as favorable sources for the generation of off-the-shelf and engineered cell products with high antitumor function (19). For CAR-NK cell therapy, the major sources of donor NK are peripheral blood (PB) and umbilical cord blood (UCB), which have their challenges. Although in vitro expansion from a single donor may be adequate for a limited number of patients, it is not enough for an off-the-shelf allogeneic commercial product (20).

Despite these advantages, there are still some challenges ahead in iPSC-CAR-NK cells in clinical applications, including the stop of CAR-iPSC-NK cell

proliferation in vivo after exogenous administration of cytokines and the immunogenic and malignant transformation potential of these cells (12).

1.9. CAR Transduction Platform in NK cells

Efficient and secure genetic delivery systems are crucial for the successful production of genetically modified cells; therefore, different approaches, including viral and nonviral platforms, are applied for CAR delivery (21). Notably, the considerable difference between these technologies is the duration of CAR expression stability.

1.10. Retroviruses

For decades, various studies have shown highly efficient CAR transduction into pre-stimulated NK cells by using a single dose of retroviral vectors ranging from 27 % to 52% (22). However, Imamura et al. achieved more impressive transduction (approximately 70%) of IL15 expression using a retroviral vector-based method. In addition, long-term persistence of retrovirally CAR-engineered NK cells has been reported. According to a recent clinical trial targeting lymphoid tumors (non-Hodgkins lymphoma and CLL), 7 of 11 patients completely recovered after infusion of retrovirus-transduced anti-CD19 CAR cord blood NK cells, and remarkably after a 1-year follow-up, expanded CAR-NK cells were detectable in the peripheral blood of treated patients (18).

1.11. Lentivirus

As with retroviruses, lentiviruses have also been greatly exploited for decades, with the difference that lentiviruses are not dependent on cell cycle progression and can transduce cycling and ever-non-cycling cells with higher transduction efficacy. Lentiviruses also possess certain other benefits such as low immunogenicity and the ability to integrate transgenes into the host genome, which can result in the permanent expression of the transgene (23).

1.12. Non-viral Approaches

NK cells have demonstrated a higher level of resistance to viral infection than T cells. This resistance may be attributed to the inherent ability of NK cells to protect against viral infections (24). In addition, alternative methods such as transposon vectors and electroporation of DNA or RNA plasmids have gained interest due to their safety, outstanding design

flexibility, ease of producing therapeutic cells on a large scale at a reasonable speed, and low cost (25).

1.13. Non-viral Electroporation (m-RNA Electroporation)

Electroporation, a commonly used transient technique, involves the use of electric pulses to make the cell membrane permeable and introduce genetic material into the cells. Various studies have relied on electroporation (26). Electroporated cargo mostly includes CAR-encoding mRNA or plasmids. Despite the satisfactory effectiveness of electrotransfection in delivering CAR-encoded DNA, various studies have indicated a potential danger of growth inhibition and mortality in NK cells, which hampers the application of DNA. In contrast, when DNA-electroporated NK cells were compared with mRNA-electroporated NK92 cells after 24 h, it was found that the DNA-electroporated NK92 cells showed 2- or 3-times lower cell viability (27). Therefore, more focus has been placed on mRNA-based electroporation because of its superior transfection efficiency (up to 95%) (28).

1.14. Nanoparticles

Viral transduction and electroporation techniques have been successful in achieving high levels of gene expression in NK cells. However, these methods are time-consuming and require specialized equipment and laboratory setups. In contrast, mRNA-carrying nanoparticles, which can be easily synthesized in the lab, have shown promise in CAR cell technology (29). These nanoparticles can be made from various materials such as polymers or ionizable lipids. Kim et al. used polymeric nanoparticles complexed with PDNA EGFR CAR to efficiently transfect NK92MI cells and demonstrated anti-oncogenic activity in a breast cancer model (30). Ionizable lipid nanoparticle platforms are particularly attractive because of their ability to provide stable formulation, low toxicity rates, and effective endogenous cellular internalization (31).

1.15. Transposons

The use of transposons has gained significant attention over traditional methods because of several advancements. These include their high effectiveness, ability to transduce CAR transgenes at specific locations (32), transfection of more than 10 kb gene fragments, lower immunogenicity, and cost-effectiveness (33).

Transposons consist of mobile plasmids with 2 terminal inverted repeats (TIRL) that carry the enzymatic gene and the desired sequence. The

transposase enzyme facilitates the cutting and pasting of these desired elements into the host genome (34). Two commonly used transposons are Sleeping Beauty (SB) and PiggyBac (PB), which have shown promise in therapeutic cell manufacturing. SB-based CAR-modified cells are currently being clinically tested because of their potential and safer integration profile. However, generating CAR-NK cells using transposons is more challenging than generating primary NK cells. Nonetheless, there have been studies using transposon systems to generate CAR-NK cells (35).

1.16. Crisper-cas9

The clustered regulatory interspaced short palindromic repeats (CRISPR) associated protein (CAS9) system has greatly advanced biomedical research by providing a powerful tool for genome editing (36). This technology involves introducing cas9 as a nuclease along with guide RNA (sgRNA) to a specific location in the genome, causing double-strand breaks. The desired gene can, then, be integrated through endogenous DNA repair mechanisms, such as homologous recombination (HR) or non-homologous recombination (NHEJ). Initially, this technology was used to knock out different genes; for example, as CD38 is expressed both on NK cells and AML cells, in one study to disrupt CD38 of NK cells. These cells were electroporated with sg RNA-cas9 to prevent fratricide of NK cells when used along with the anti-CD38 antibody daratumumab (36). Another study successfully knocked out approximately 80% of the NKG2A-encoding killer cell lectin-like receptor C1 (KLRC1) locus in primary NK cells using CRISPR/CAS9 (37). In recent times, the technique of CRISPR/CAS9 has also been used to incorporate CRISPR/CAS9 plasmid, which has caused NK cells to be resistant to this technology, which is a major issue for gen-editing-based immunotherapy using CRISPR/CAS9. Thus far, numerous approaches have been used in an attempt to create a secure and effective delivery system (38). However, when the expression levels of the CAR gene can vary when it is delivered through transposon-based random insertion, electroporation of cas9 single guide RNA complexes has been established as a method to deliver large-sized CRISPR/CAS9 plasmids into NK cells (39).

1.17. Trogocytosis

Trogocytosis refers to the exchange of membrane patches between immune cells and target cells. When an NK cell interacts with a target cell, a strong immune synapse is formed, enabling the transfer of small membrane patches between two cells. As a result,

surface molecules from the target cell can be detected on the surface of NK cells. This process has been used to modify the expression of certain molecules on NK cells, enhancing their ability to destroy target cells (40).

Transfer of the chemokine receptor CCR7 from engineered K562 cells to human NK cells through trogocytosis was intentionally facilitated. This process, as observed in a study conducted by Somanchi et al., led to an increase in CCR7 expression in 80% of NK cells after being co-cultured for 1 h. Additionally, this resulted in improved migration to lymph nodes (41).

1.18. Microfluidic-Based Cell Squeezing

Devices that use microfluidic technologies to squeeze cells have been quickly developed to effectively deliver large and small molecules into different types of cells. This method works by mechanically modifying the cells, temporarily making their cellular membranes permeable. Unlike electroporation, this technique has a minimal negative effect on the functionality of immune cells (42). In the future, this approach is anticipated to be a valuable strategy for delivering substances without viruses. However, more research is needed to understand how this technique specifically affects immune cells such as NK cells (43).

1.19. CAR-NK vs. CAR-T and CAR-M for Cancer Immunotherapy

In recent years, considerable progress has been made in CAR-T cell immunotherapy, particularly for the treatment of hematological malignancies. To date, 6 CAR-T cell therapies, namely Kymriah, Yescarta, Breyanzi, Tecartus, Carvykti, and Abecma have been approved by the FDA. Despite impressive results, there are unwanted disadvantages and obstacles associated with CAR-T therapy (44). The key challenges faced in clinical applications are as follows: First, its manufacturing processes are intricate and centralized and, thus, require a high cost, which reduces patients' access to treatment. The next issue is to reduce autologous T-cell efficacy in heavily treated patients (45). Although allogeneic CAR-T cells can solve this challenge, GvHD and host allograft rejection are the two main limitations. Unlike the successful effect of CAR-T in hematological cancers, it has minimal function in solid cancers (46). Common CAR-T side effects are cytokine release syndrome (CRS) and neurotoxicity (47). Target antigen loss provides a great opportunity for tumor immune escape, which occurs after CAR T-cell therapy. The exhaustion phenotype and limited durability of CAR-T cells suppress their function. Because of these deficiencies, the clinical use of CAR-T is limited. Hence,

to solve these challenges, the use of alternative cell sources such as NK cells and macrophages is an ideal and promising option because of their desirable properties. Therefore, we compared the potential benefits of CAR-NK and CAR-M cells with those of CAR-T cells.

1.20. CAR-NK Cell Therapy: Advantages and Limitations

The great achievements in CAR-T cell therapies have led to the development of subsequent generations of NK-based cell therapies (48). Given the above limitations, CAR-NK cells may be superior to CAR-T cells, especially in cancer immunotherapy, as shown in several clinical trials. CAR structures for NK cells can be designed similar to CAR in CAR-T cells, and T cell technology is easily applied to NK cells.

1.21. Advantages

Evidence-based CAR-NK cell therapy is an innovative strategy with great potential for cancer immunotherapy (49). CD8+ T cells and NK cells use the same mechanisms to kill infected and cancer cells, but they have drastically different mechanisms of target recognition (50). Both kill target cells through lysosomal granule release and the Fas ligand and produce cytokines. The significant advantages offered by CAR-NK cells over CAR-T cells make them a novel candidate in the context of cancer immuno-cell therapy (51). First, they can quickly identify and kill their aims without prior sensitization. Although they do not require HLA matching, they cannot cause GvHD; therefore, they might be generated on an off-the-shelf platform for wider clinical use. As a result, NK cells can be generated from existing cell lines as well as from allogeneic NK cells that have mismatched major histocompatibility complex (MHC) molecules (52). As a result, there are broad cell sources used to construct CAR-NK, but CAR-T cells cannot be used for allogeneic T therapy because of GvHD (52). Second, triggered NK cells can destroy targeted cells not only through the CAR pathway but also through other pathways, such as the release of proinflammatory cytokines (perforin and granzymes) and another mechanism using antibody-dependent cellular cytotoxicity (ADCC) through Fas ligand (FasL) expression and TNF-related apoptosis-inducing ligand (TRAIL). Third, the main mechanisms of tumor escape from CAR-T cells occur through decreased MHC-I, whereas CAR-NK cells maintain their innate cytolytic capacity via self-missing recognition (53). Fourth, studies have shown that, unlike the activation of CAR-T cells that release proinflammatory cytokines (including IL-1, IL-2, IL-6, and IL-15), activated NK cells

produce different cytokines, such as IFN- γ , TNF- α , interleukin-3, and GM-CSF; therefore, CRS and neurotoxicity rarely occur during CAR-NK cell therapy.

1.22. Limitations

Most obstacles associated with CAR-T cell therapy are exerted by CAR-NK cells, such as target antigen selection, CAR design, diversity of antigens, post-infusion challenge generation, such as NK cells moving to tumor locations, and a belligerent tumor microenvironment (52). First, as discussed above, CAR structures in NK cells are similar to those of CAR-T cells, although they are low-quality selections for use in NK cells (54). Second, transduction methods, including viral transfection and electroporation, have low efficiency (55). Third, a lack of cytokine support leads to limited durability in vivo. Although desirable, this can reduce the influence of CAR-NK cell immunotherapy (21). Fourth, in ex vivo-expanded human NK cells, unlike T cells, the number of NK cells in the blood is limited; therefore, to reach a large scale, the use of expansion techniques is required. Fifth, the tumor microenvironment is a major problem in reducing the ability of CAR-NK cell therapy to eliminate target cells, including immunosuppressive cell types.

1.23. Pre-clinical Studies

Preclinical studies have demonstrated very promising anti-tumor results for CAR-NK cells in hematological and solid tumor cells. Among multiple sources of NK cells for CAR modification, the NK-92 cell line is largely used in both hematological and solid tumors. Some preclinical studies have investigated the efficacy and safety of CAR-NK cells against tumor antigens such as HER2, CD20, CD19, and CD244 (56).

1.24. Preclinical Studies of Hematologic Cancers

The majority of preclinical and clinical success on CAR-NK cells has been performed on hematologic malignancies (57). NK cell CAR-specific targets, including CD19, CD138, CD20, and Flt3, have been successfully used to treat hematologic malignancies (58). To direct cytotoxicity against B-cell malignancies, NK cells have been genetically manufactured to represent CARs. CD19-CAR-NK cells can be generated from iPSCs, PB, and UCB, which have been shown to eliminate B-cell malignancies in vitro (59). The most common purpose of hematological malignancies is the CD19 antigen (B cell malignancies). NK-92 cells have shown effective cytotoxic activity against several lymphoblastic cells expressing CD19 (11). Dual-targeting

Table 1. Preclinical Studies of CAR-NK Cells

Malignancy	Target	Source of NK Cells	Result	Reference
B-cell malignancies	CD19 CD20 Flt3	NK-92, PB-NK or CB-NK NK-92 NK-92	NK92-CD19-CD3ζ cells have cytotoxic effects against B-lineage malignancy. Transfection of CD19-41BB-mRNA into NK cells with electroporation technology illustrated that 61.3% of cells stably present anti-CD19 CAR, following that, optical in vivo imaging proved that CAR-NK cell therapy is effective in reducing the growth rate of leukemia xenografts. In vitro, NK-92 cells carrying anti-CD19 CAR demonstrated excellent cytolytic activity against previously resistant CD19-positive B-ALL and primary chronic lymphocytic leukemia (CLL) cell lines. CAR-NK-92 cells with CD20 and Flt3 expression in immunotherapy against B-cell tumors that lead to degranulation of NK cells and selective cytotoxicity. In vitro, anti-CD20 CAR-adjusted and expanded peripheral blood NK cells reduce the tumor burden of CD20+ B-NHL cells in vitro. Anti-CD20 CAR-NK cells have higher anti-tumor toxicity than anti-CD20 antibodies in chronic lymphocytic leukemia (CLL) cells.	(61, 64)
T-cell malignancies	CD3 CD5 CD7	NK-92 NK-92 NK-92	CAR-adjusted NK cells or NK-92 cell lines to target CD3, CD5, and CD7 revealed important anti-tumor activity against T-cell malignancies.	(60, 65)
Multiple myeloma	CD138 CS1 BCMA NKG2D	NK-92 NK-92 NK-92 PB-NK	CS1-CAR NK-92 cells enhanced IFN-γ production and cytotoxic activity in response to primary multiple myeloma (MM) tumor cells with high expression CS1. CD138-CAR-NK-92 eliminates myeloma cells and prolongs the survival rate of MM mice. The therapy with BCMA-CAR-NK cells with co-expressing CXCR4 reduced the in vivo progression of MM and the mice's survival was extended.	(66, 67)
AML	NKG2D	PB-NK	NK cells derived from peripheral blood with NKG2D-specific CAR exhibited their ability to lyse AML tumor cells in vitro and in a xenograft model.	(68)
Glioblastoma	HER2 EGFR and/or EGFRvIII	NK-92 NK-92, NKL, KHYG1 or YTS	Intravenous injection of EGFRvIII-directed CAR-NK cells with overexpressing of CXCR4 receptor, mice was treated and prolonged their survival. Bispecific CAR-NK-92 cells by EGFR and EGFRvIII-driven were injected intracranially in xenograft mouse glioma models, prolonged the mouse survival and enhanced cytotoxicity and IFN-γ secretion. Also, ErbB2(HER2)-CAR-NK-92/5.28. z cells with CD3ζ and CD28ζ signaling domains have shown that they can eradicate ErbB2-positive glioblastoma cells in vitro.	(69, 70)
Ovarian cancer	HLA-G CD24 αFR CD133 CD44 Mesothelin	PB-NK NK-92 NK-92 NK-92 iPSC-NK or NK-92	αFR (folate receptor)-targeted CAR-NK-92 cells showed cytotoxicity specific to antigen and significantly increased survival in animal models. MSLN-CAR NK cells derived from both iPSC and NK-92 cell lines illustrated the effective elimination of MSLN-positive OC cells (OVCAR-3 and SK-OV-3).	(32, 71, 72)
Breast cancer	EpCAM B7-H6 TF HER2 EGFR and/or EGFRvIII	NK-92 NK-92 NK-92 NK-92 or PB-NK	NK-92/31.28. z cells were applied against EpCAM and demonstrated potent cytotoxicity and cytokine secretion. B7-H6 and TF as recent targets in CAR-NK immunotherapy enhanced tumor-killing capacity of specific CAR-NK-92 cells. The NK-92-scFv (FRP5)-zeta cell line presenting a chimera HER2 antigen receptor in breast cancer with HER-2 mutation resulting in vivo antitumor activity. EGFR-CAR-NK cells could lyse TNBC in vitro and curtailing tumor growth. Dual-specific CAR-NK-92 cell, capable of recognizing both EGFR and EGFRvIII, has shown high cytotoxic and produce IFN-γ to battle breast cancer cells.	(73, 74)

CAR approaches targeting CD19 and CD138 antigens used to produce CAR-NK-92 cells showed in vitro cytotoxicity and purposed tumor killing (60).

1.25. Preclinical Studies of Solid Cancers

Although CAR-T-cell immunotherapy has revealed considerable progress in treating hematologic malignancies, solid tumor limitations such as immunosuppressive TME and poor perfusion to the tumor have not been solved (61). In preclinical studies, CAR-NK cells can use promising immunotherapy strategies, such as overexpression of the chemokine receptors CXCR4 or CXCR1, which provide particular chemotaxis to remedy these limitations (62). Thus, multiple solid tumors such as glioblastoma, breast cancer, colon cancer, hepatocellular carcinoma, pancreatic cancer, and head and neck squamous cell carcinoma (HNSCC) are the researched options for CAR-NK-cell preclinical study (63). All preclinical studies are summarized in Table 1.

1.26. Clinical Studies

Confirmation of the safety and effectiveness of non-engineered NK cells and reassuring evidence from preclinical investigations of CAR-NK treatments support the clinical benefits of CAR-NK cells in patients. Since the initial recorded CAR-NK cell trial in 2009 (NCT00995137), there have been 60 registered trials on clinicaltrials.gov. Most of the clinical studies are related to recent years (mainly between the years of 2021-2023), as 15, 14, and 9 clinical trials have been registered on clinicaltrials.gov in 2021, 2022, and 2023, respectively. Among the registered studies, 10 trials had an unknown status, and 2 trials were withdrawn. Although 4 studies have completed status on clinicaltrials.gov, there is a lack of accessible information regarding these trials. The majority of related clinical trials focus on hematologic cancers, particularly B-cell lineage leukemia/lymphoma. Despite CD19 being the most common targeted antigen, CD20, CD22, CD2, NKG2D, BCMA, CD5, CD70, CD33, and CD123 are other markers used for targeting lymphoproliferative tumors.

In this regard, during phase 1 and 2 clinical trials, CB-derived CD19-CAR-NK using K562/mbIL-21/41BBL was administered to 11 volunteers with r/r CD19-positive B-cell malignancies. Following the lymphadenopathy

regimen, the injection of engineered cells was performed at only 1 of 3 dosages (10×10^4 , 10×10^5 , or 10×10^6 CAR-NK cells/kg) of a single infusion. The findings demonstrated that 73% of the patients had objective responses with no serious side effects. In addition, at all applied doses, responses were quick and visible after 30 days of infusion (18). Despite the favorable results of this trial, Karadimitris in 2020 presented a high percentage of transient clinical responses to CB-derived CAR-NK cell-based immunotherapy (75). In addition, preliminary findings from a phase I clinical study [NCT04245722] that evaluated the effect of iPSC-derived CD19-directed NK cells (FT596) on 20 patients with r/r B-cell lymphomas (BCLs) and CLL have been published. To increase the efficacy of NKs, the cells were modified for 3 antigens: CD19, CD16, and IL-15 receptors and used in combination with monoclonal antibodies. Data, while exhibiting (no evidence of immune effector cell-associated neurotoxicity syndrome, GVHD, and cytokine release syndrome), displayed an objective response in 11 patients from all 17 evaluable subjects (76). Another equivalent study used a single dose of FT596 (30×10^6 cells) in a female patient aged 76 years with diffuse large B-cell lymphoma. The results illustrated the early clinical benefits of FT596, such as safety, no GVHD, no neurologic toxicity, no dose-limiting toxicities, and no remarkable adverse reactions related to cell therapy. According to the 2014 Lugano criteria, the measurement of the tumoral response at the one-month mark revealed a partial response, with a decrease of more than 70% in ^{18}F -Glu absorption and a reduction of more than 50% in tumoral size. Furthermore, a phase I trial of this protocol [NCT04245722] is in progress, with 285 individuals expected to participate (77).

Notably, 17 clinical trials are related to solid tumors, including brain, lung, prostate, colorectal, ovarian, prostate, pancreatic, and other types. They mostly selected HER2, NKG2D, mesothelin, CCR, Claudin6, ROBO, DLL3, PSMA, and CD70 markers to target tumor cells using CAR constructs. One promising target for CAR-NK treatment of solid malignancies is the Robo-1 antigen. A case report study intravenously infused 1×10^9 Robo-1-specific CAR-NK cells into a 46-year-old man with pancreatic ductal adenocarcinoma followed up every two weeks. The evaluation showed 5 months of controlled pancreatic lesions, no significant adverse outcomes, and an overall survival time of 8 months. Based on the noted research and preclinical experiments, 3 phase I/II clinical studies [NCT03941457, NCT03940820, and NCT03931720] in China are recruiting for the investigation of ROBO-1-specific CAR-

NK cells in solid tumors (57). In a similar strategy, PD-L1 antigen was considered for immunotherapy of solid cancers, so PD-L1 targeting CAR-NK combined with IL-15 superagonist (N-803) and pembrolizumab is presently currently appraised for gastroesophageal junction and HNSCC tumor participants in the form of a phase II clinical trial (63). Another trial [NCT03415100] was conducted to assess the safety and feasibility of local administration of allogeneic or autologous NKG2DL-engineered CAR-NK cells in 3 subjects with metastatic colon cancer. Two patients achieved a decrease in the production of ascites and a remarkable drop in the number of tumor cells in ascites specimens; however, the third patient, who had liver metastases, indicated a complete metabolic response and rapid tumor regression of liver lesions (78). Furthermore, the FDA approved the FT536 product (by Fate Therapeutics) for patients with advanced solid malignancies in January 2022 (NCT05395052). This product is an allogeneic iPSC-derived CAR-NK cell line targeting the alpha-3 domain of MHC class I chain-related proteins A (MICA) and B (MICB). A comprehensive overview of all clinical trials is presented in Table 2.

1.27. CAR-NK Cell Treatment in the Foreseeable Future and Schedules to Overcome Their Limitations

The major challenges of optimal CAR-NK cell therapy for cancers include immunosuppressive TME, antigen loss, cancer heterogeneity, and impaired homing and distribution of NK cells to tumor sites. These obstacles hamper the persistence and accumulation of NK cells and their cytotoxic mechanisms (79). Several studies have employed innovative solutions to overcome its inevitable limitations.

Unlike CAR-T cells, CAR-NK cells possess the capability to detect tumor antigens and mediate cytotoxic action in both CAR-dependent and CAR-independent ways. Hence, it is possible to design CARs without a co-stimulatory signaling domain and induce intrinsic mechanisms of NK cells for cell-killing rather than CAR-induced direct cytotoxicity. Engineering NK cells to express homing-promoting target factors such as chemokines and adhesion molecules is another non-signaling CAR construct. Similarly, the insertion of additional co-stimulatory domains, such as DNAX-activation proteins 10 or 12 (DAP-10 or DAP-12) and NK-specific 2B4, into the CAR structure could amplify the immunoregulatory impact of CAR-NK cells (57). Another potential method involves co-modifying CAR constructs with cytokines, antibodies, and proteases to improve the proliferation, trafficking, persistence, and deep tumor penetration of NK cells.

2. Conclusions

The emergence of "off-the-shelf" CAR-NK cells has opened a new promising window for clinical use. With the ability to tackle these challenges, this product presents a safe and available CAR-T cell alternative for immunotherapy in different types of cancers, even in HLA-unmatched patients. However, some limitations, including the response of the recipient's immune system and the short lifespan of the injected cells in the body, as well as the immunosuppressive TME, limit their potential applications.

Footnotes

Authors' Contribution: Conceptualization: M.B and M.A. Writing the original draft: M.S.S, S.T and S.T. Review and Editing: M.N, M.S.S, S.T, S.T and A.B. Validation: M.V, J.V and S.R.

Conflict of Interests Statement: The authors declare no conflict of interest.

Funding/Support: All authors declare that they have not received any funding/support from any organization regarding the submitted manuscript

References

- Abel AM, Yang C, Thakar MS, Malarkannan S. Natural Killer Cells: Development, Maturation, and Clinical Utilization. *Front Immunol.* 2018;**9**:1869. [PubMed ID: 30150991]. [PubMed Central ID: PMC6099181]. <https://doi.org/10.3389/fimmu.2018.01869>.
- Herberman RB, Nunn ME, Holden HT, Lavrin DH. Natural cytotoxic reactivity of mouse lymphoid cells against syngeneic and allogeneic tumors. II. Characterization of effector cells. *Int J Cancer.* 1975;**16**(2):230-9. [PubMed ID: 1080480]. <https://doi.org/10.1002/ijc.2910160205>.
- Carrega P, Ferlazzo G. Natural killer cell distribution and trafficking in human tissues. *Front Immunol.* 2012;**3**:347. [PubMed ID: 23230434]. [PubMed Central ID: PMC3515878]. <https://doi.org/10.3389/fimmu.2012.00347>.
- Langers I, Renoux VM, Thiry M, Delvenne P, Jacobs N. Natural killer cells: role in local tumor growth and metastasis. *Biologics.* 2012;**6**:73-82. [PubMed ID: 22532775]. [PubMed Central ID: PMC3333822]. <https://doi.org/10.2147/BTT.S23976>.
- Sun JC, Lanier LL. NK cell development, homeostasis and function: parallels with CD8(+) T cells. *Nat Rev Immunol.* 2011;**11**(10):645-57. [PubMed ID: 21869816]. [PubMed Central ID: PMC4408539]. <https://doi.org/10.1038/nri3044>.
- Karmakar S, Pal P, Lal G. Key Activating and Inhibitory Ligands Involved in the Mobilization of Natural Killer Cells for Cancer Immunotherapies. *Immunotargets Ther.* 2021;**10**:387-407. [PubMed ID: 34754837]. [PubMed Central ID: PMC8570289]. <https://doi.org/10.2147/JTT.S306109>.
- Ljunggren HG, Karre K. In search of the 'missing self': MHC molecules and NK cell recognition. *Immunol Today.* 1990;**11**(7):237-44. [PubMed ID: 2201309]. [https://doi.org/10.1016/0167-5699\(90\)90097-s](https://doi.org/10.1016/0167-5699(90)90097-s).
- Dong Y, Wan Z, Gao X, Yang G, Liu L. Reprogramming Immune Cells for Enhanced Cancer Immunotherapy: Targets and Strategies. *Front Immunol.* 2021;**12**:609762. [PubMed ID: 33968014]. [PubMed Central ID: PMC8097044]. <https://doi.org/10.3389/fimmu.2021.609762>.
- Lu H, Zhao X, Li Z, Hu Y, Wang H. From CAR-T Cells to CAR-NK Cells: A Developing Immunotherapy Method for Hematological Malignancies. *Front Oncol.* 2021;**11**:720501. [PubMed ID: 34422667]. [PubMed Central ID: PMC8377427]. <https://doi.org/10.3389/fonc.2021.720501>.
- Yilmaz A, Cui H, Caligiuri MA, Yu J. Chimeric antigen receptor-engineered natural killer cells for cancer immunotherapy. *J Hematol Oncol.* 2020;**13**(1):168. [PubMed ID: 33287875]. [PubMed Central ID: PMC7720606]. <https://doi.org/10.1186/s13045-020-00998-9>.
- Romanski A, Uhrek C, Bug G, Seifried E, Klingemann H, Wels WS, et al. CD19-CAR engineered NK-92 cells are sufficient to overcome NK cell resistance in B-cell malignancies. *J Cell Mol Med.* 2016;**20**(7):1287-94. [PubMed ID: 27008316]. [PubMed Central ID: PMC4929308]. <https://doi.org/10.1111/jcmm.12810>.
- Gong Y, Klein Wolterink RGJ, Wang J, Bos GMJ, Germaad WTV. Chimeric antigen receptor natural killer (CAR-NK) cell design and engineering for cancer therapy. *J Hematol Oncol.* 2021;**14**(1):73. [PubMed ID: 33933160]. [PubMed Central ID: PMC8088725]. <https://doi.org/10.1186/s13045-021-01083-5>.
- Williams BA, Law AD, Routy B, denHollander N, Gupta V, Wang XH, et al. A phase I trial of NK-92 cells for refractory hematological malignancies relapsing after autologous hematopoietic cell transplantation shows safety and evidence of efficacy. *Oncotarget.* 2017;**8**(51):89256-68. [PubMed ID: 29179517]. [PubMed Central ID: PMC5687687]. <https://doi.org/10.18632/oncotarget.19204>.
- Shah NN, Baird K, Delbrook CP, Fleisher TA, Kohler ME, Rampertaap S, et al. Acute GVHD in patients receiving IL-15/4-1BB activated NK cells following T-cell-depleted stem cell transplantation. *Blood.* 2015;**125**(5):784-92. [PubMed ID: 25452614]. [PubMed Central ID: PMC4311226]. <https://doi.org/10.1182/blood-2014-07-592881>.
- Martin-Antonio B, Sune G, Perez-Amill L, Castella M, Urbano-Ispizua A. Natural Killer Cells: Angels and Devils for Immunotherapy. *Int J Mol Sci.* 2017;**18**(9). [PubMed ID: 28850071]. [PubMed Central ID: PMC5618517]. <https://doi.org/10.3390/ijms18091868>.
- Zhao X, Cai L, Hu Y, Wang H. Cord-Blood Natural Killer Cell-Based Immunotherapy for Cancer. *Front Immunol.* 2020;**11**:584099. [PubMed ID: 33193399]. [PubMed Central ID: PMC7642255]. <https://doi.org/10.3389/fimmu.2020.584099>.
- Liu E, Tong Y, Dotti G, Shaim H, Savoldo B, Mukherjee M, et al. Cord blood NK cells engineered to express IL-15 and a CD19-targeted CAR show long-term persistence and potent antitumor activity. *Leukemia.* 2018;**32**(2):520-31. [PubMed ID: 28725044]. [PubMed Central ID: PMC6063081]. <https://doi.org/10.1038/leu.2017.226>.
- Liu E, Marin D, Banerjee P, Macapinlac HA, Thompson P, Basar R, et al. Use of CAR-Transduced Natural Killer Cells in CD19-Positive Lymphoid Tumors. *N Engl J Med.* 2020;**382**(6):545-53. [PubMed ID: 32023374]. [PubMed Central ID: PMC7101242]. <https://doi.org/10.1056/NEJMoa1910607>.
- Arias J, Yu J, Varshney M, Inzunza J, Nalvarte I. Hematopoietic stem cell- and induced pluripotent stem cell-derived CAR-NK cells as reliable cell-based therapy solutions. *Stem Cells Transl Med.* 2021;**10**(7):987-95. [PubMed ID: 33634954]. [PubMed Central ID: PMC8235144]. <https://doi.org/10.1002/sctm.20-0459>.
- Quintarelli C, Sivori S, Caruso S, Carlomagno S, Falco M, Boffa I, et al. Efficacy of third-party chimeric antigen receptor modified peripheral blood natural killer cells for adoptive cell therapy of B-cell precursor acute lymphoblastic leukemia. *Leukemia.* 2020;**34**(4):1102-15. [PubMed ID: 31745215]. <https://doi.org/10.1038/s41375-019-0613-7>.
- Matosevic S. Viral and Nonviral Engineering of Natural Killer Cells as Emerging Adoptive Cancer Immunotherapies. *J Immunol Res.*

- 2018;**2018**:4054815. [PubMed ID: [30306093](#)]. [PubMed Central ID: [PMC6166361](#)]. <https://doi.org/10.1155/2018/4054815>.
22. Streltsova MA, Barsov E, Erokhina SA, Kovalenko EI. Retroviral gene transfer into primary human NK cells activated by IL-2 and K562 feeder cells expressing membrane-bound IL-21. *J Immunol Methods*. 2017;**450**:90-4. [PubMed ID: [28802832](#)]. <https://doi.org/10.1016/j.jim.2017.08.003>.
23. June CH, Blazar BR, Riley JL. Engineering lymphocyte subsets: tools, trials and tribulations. *Nat Rev Immunol*. 2009;**9**(10):704-16. [PubMed ID: [19859065](#)]. [PubMed Central ID: [PMC3412112](#)]. <https://doi.org/10.1038/nri2635>.
24. Schmidt P, Raftery MJ, Pecher G. Engineering NK Cells for CAR Therapy-Recent Advances in Gene Transfer Methodology. *Front Immunol*. 2020;**11**:611163. [PubMed ID: [33488617](#)]. [PubMed Central ID: [PMC7817882](#)]. <https://doi.org/10.3389/fimmu.2020.611163>.
25. Robbins GM, Wang M, Pomeroy EJ, Moriarity BS. Nonviral genome engineering of natural killer cells. *Stem Cell Res Ther*. 2021;**12**(1):350. [PubMed ID: [34134774](#)]. [PubMed Central ID: [PMC8207670](#)]. <https://doi.org/10.1186/s13287-021-02406-6>.
26. Karvouni M, Vidal-Manrique M, Lundqvist A, Alici E. Engineered NK Cells Against Cancer and Their Potential Applications Beyond. *Front Immunol*. 2022;**13**:825979. [PubMed ID: [35242135](#)]. [PubMed Central ID: [PMC8887605](#)]. <https://doi.org/10.3389/fimmu.2022.825979>.
27. Boissel L, Betancur M, Wels WS, Tuncer H, Klingemann H. Transfection with mRNA for CD19 specific chimeric antigen receptor restores NK cell mediated killing of CLL cells. *Leuk Res*. 2009;**33**(9):1255-9. [PubMed ID: [19147228](#)]. [PubMed Central ID: [PMC3047414](#)]. <https://doi.org/10.1016/j.leukres.2008.11.024>.
28. Chu Y, Yahr A, Huang B, Ayello J, Barth M, S. Cairo M. Romidepsin alone or in combination with anti-CD20 chimeric antigen receptor expanded natural killer cells targeting Burkitt lymphoma in vitro and in immunodeficient mice. *Oncoimmunology*. 2017;**6**(9). e1341031. [PubMed ID: [28932644](#)]. [PubMed Central ID: [PMC5599075](#)]. <https://doi.org/10.1080/2162402X.2017.1341031>.
29. Chen J, Guo Z, Tian H, Chen X. Production and clinical development of nanoparticles for gene delivery. *Mol Ther Methods Clin Dev*. 2016;**3**:16023. [PubMed ID: [27088105](#)]. [PubMed Central ID: [PMC4822651](#)]. <https://doi.org/10.1038/mtm.2016.23>.
30. Kim KS, Han JH, Park JH, Kim HK, Choi SH, Kim GR, et al. Multifunctional nanoparticles for genetic engineering and bioimaging of natural killer (NK) cell therapeutics. *Biomaterials*. 2019;**221**:119418. [PubMed ID: [31419655](#)]. <https://doi.org/10.1016/j.biomaterials.2019.119418>.
31. Huang J, Yuen D, Minter JD, Johnston APR. Opportunities for innovation: Building on the success of lipid nanoparticle vaccines. *Curr Opin Colloid Interface Sci*. 2021;**55**:101468. [PubMed ID: [34093062](#)]. [PubMed Central ID: [PMC8164502](#)]. <https://doi.org/10.1016/j.cocis.2021.101468>.
32. Li Y, Hermanson DL, Moriarity BS, Kaufman DS. Human iPSC-Derived Natural Killer Cells Engineered with Chimeric Antigen Receptors Enhance Anti-tumor Activity. *Cell Stem Cell*. 2018;**23**(2):181-192 e5. [PubMed ID: [30082067](#)]. [PubMed Central ID: [PMC6084450](#)]. <https://doi.org/10.1016/j.stem.2018.06.002>.
33. Tsai HC, Pietrobon V, Peng M, Wang S, Zhao L, Marincola FM, et al. Current strategies employed in the manipulation of gene expression for clinical purposes. *J Transl Med*. 2022;**20**(1):535. [PubMed ID: [36401279](#)]. [PubMed Central ID: [PMC9673226](#)]. <https://doi.org/10.1186/s12967-022-03747-3>.
34. Magnani CF, Tettamanti S, Alberti G, Pisani I, Biondi A, Serafini M, et al. Transposon-Based CAR T Cells in Acute Leukemias: Where are We Going? *Cells*. 2020;**9**(6). [PubMed ID: [32471151](#)]. [PubMed Central ID: [PMC7349235](#)]. <https://doi.org/10.3390/cells9061337>.
35. Hudecek M, Izsvak Z, Johnen S, Renner M, Thumann G, Ivics Z. Going non-viral: the Sleeping Beauty transposon system breaks on through to the clinical side. *Crit Rev Biochem Mol Biol*. 2017;**52**(4):355-80. [PubMed ID: [28402189](#)]. <https://doi.org/10.1080/10409238.2017.1304354>.
36. Savic N, Schwank G. Advances in therapeutic CRISPR/Cas9 genome editing. *Transl Res*. 2016;**168**:15-21. [PubMed ID: [26470680](#)]. <https://doi.org/10.1016/j.trsl.2015.09.008>.
37. Bexte T, Alzubi J, Reindl LM, Wendel P, Schubert R, Salzmann-Manrique E, et al. CRISPR-Cas9 based gene editing of the immune checkpoint NKG2A enhances NK cell mediated cytotoxicity against multiple myeloma. *Oncoimmunology*. 2022;**11**(1):2081415. [PubMed ID: [35694192](#)]. [PubMed Central ID: [PMC9176243](#)]. <https://doi.org/10.1080/2162402X.2022.2081415>.
38. van Haasteren J, Li J, Scheideler OJ, Murthy N, Schaffer DV. The delivery challenge: fulfilling the promise of therapeutic genome editing. *Nat Biotechnol*. 2020;**38**(7):845-55. [PubMed ID: [32601435](#)]. <https://doi.org/10.1038/s41587-020-0565-5>.
39. Jo DH, Kaczmarek S, Shin O, Wang L, Cowan J, McComb S, et al. Simultaneous engineering of natural killer cells for CAR transgenesis and CRISPR-Cas9 knockout using retroviral particles. *Mol Ther Methods Clin Dev*. 2023;**29**:173-84. [PubMed ID: [37063482](#)]. [PubMed Central ID: [PMC10102412](#)]. <https://doi.org/10.1016/j.omtm.2023.03.006>.
40. Joly E, Hudrisier D. What is trogocytosis and what is its purpose? *Nat Immunol*. 2003;**4**(9):815. [PubMed ID: [12942076](#)]. <https://doi.org/10.1038/ni0903-815>.
41. Somanchi SS, Somanchi A, Cooper LJ, Lee DA. Engineering lymph node homing of ex vivo-expanded human natural killer cells via trogocytosis of the chemokine receptor CCR7. *Blood*. 2012;**119**(22):5164-72. [PubMed ID: [22498742](#)]. [PubMed Central ID: [PMC3418772](#)]. <https://doi.org/10.1182/blood-2011-11-389924>.
42. DiTommaso T, Cole JM, Cassereau L, Bugge JA, Hanson JLS, Bridgen DT, et al. Cell engineering with microfluidic squeezing preserves functionality of primary immune cells in vivo. *Proc Natl Acad Sci U S A*. 2018;**115**(46):E10907-14. [PubMed ID: [30381459](#)]. [PubMed Central ID: [PMC6243275](#)]. <https://doi.org/10.1073/pnas.1809671115>.
43. Sharei A, Zoldan J, Adamo A, Sim WY, Cho N, Jackson E, et al. A vector-free microfluidic platform for intracellular delivery. *Proc Natl Acad Sci U S A*. 2013;**110**(6):2082-7. [PubMed ID: [23341631](#)]. [PubMed Central ID: [PMC3568376](#)]. <https://doi.org/10.1073/pnas.1218705110>.
44. Kilgour MK, Bastin DJ, Lee SH, Ardolino M, McComb S, Visram A. Advancements in CAR-NK therapy: lessons to be learned from CAR-T therapy. *Front Immunol*. 2023;**14**:1166038. [PubMed ID: [37205115](#)]. [PubMed Central ID: [PMC10187144](#)]. <https://doi.org/10.3389/fimmu.2023.1166038>.
45. Tomasik J, Jasinski M, Basak GW. Next generations of CAR-T cells - new therapeutic opportunities in hematology? *Front Immunol*. 2022;**13**:1034707. [PubMed ID: [36389658](#)]. [PubMed Central ID: [PMC9650233](#)]. <https://doi.org/10.3389/fimmu.2022.1034707>.
46. Mazinani M, Rahbarizadeh F. New cell sources for CAR-based immunotherapy. *Biomark Res*. 2023;**11**(1):49. [PubMed ID: [37147740](#)]. [PubMed Central ID: [PMC10163725](#)]. <https://doi.org/10.1186/s40364-023-00482-9>.
47. Sengsayadeth S, Savani BN, Oluwole O, Dholaria B. Overview of approved CAR-T therapies, ongoing clinical trials, and its impact on clinical practice. *EJHaem*. 2022;**3**(Suppl 1):6-10. [PubMed ID: [35844299](#)]. [PubMed Central ID: [PMC9175669](#)]. <https://doi.org/10.1002/jha2.338>.
48. Ruppel KE, Fricke S, Kohl U, Schmiedel D. Taking Lessons from CAR-T Cells and Going Beyond: Tailoring Design and Signaling for CAR-NK Cells in Cancer Therapy. *Front Immunol*. 2022;**13**:822298. [PubMed ID: [35371071](#)]. [PubMed Central ID: [PMC8971283](#)]. <https://doi.org/10.3389/fimmu.2022.822298>.

49. Marofi F, Al-Awad AS, Sulaiman Rahman H, Markov A, Abdelbasset WK, Ivanovna Enina Y, et al. CAR-NK Cell: A New Paradigm in Tumor Immunotherapy. *Front Oncol*. 2021;11:673276. [PubMed ID: 34178661]. [PubMed Central ID: PMC8223062]. <https://doi.org/10.3389/fonc.2021.673276>.
50. Maalej KM, Merhi M, Inchakalody VP, Mestiri S, Alam M, Maccalli C, et al. CAR-cell therapy in the era of solid tumor treatment: current challenges and emerging therapeutic advances. *Mol Cancer*. 2023;22(1):20. [PubMed ID: 36717905]. [PubMed Central ID: PMC9885707]. <https://doi.org/10.1186/s12943-023-01723-z>.
51. Pang Z, Wang Z, Li F, Feng C, Mu X. Current Progress of CAR-NK Therapy in Cancer Treatment. *Cancers (Basel)*. 2022;14(17). [PubMed ID: 36077853]. [PubMed Central ID: PMC9454439]. <https://doi.org/10.3390/cancers14174318>.
52. Pan K, Farrukh H, Chitpeu V, Xu H, Pan CX, Zhu Z. CAR race to cancer immunotherapy: from CAR T, CAR NK to CAR macrophage therapy. *J Exp Clin Cancer Res*. 2022;41(1):19. [PubMed ID: 35361234]. [PubMed Central ID: PMC8969382]. <https://doi.org/10.1186/s13046-022-02327-z>.
53. Pfefferle A, Huntington ND. You Have Got a Fast CAR: Chimeric Antigen Receptor NK Cells in Cancer Therapy. *Cancers (Basel)*. 2020;12(3). [PubMed ID: 32192067]. [PubMed Central ID: PMC7140022]. <https://doi.org/10.3390/cancers12030706>.
54. Wang W, Jiang J, Wu C. CAR-NK for tumor immunotherapy: Clinical transformation and future prospects. *Cancer Lett*. 2020;472:175-80. [PubMed ID: 31790761]. <https://doi.org/10.1016/j.canlet.2019.11.033>.
55. Khawar MB, Sun H. CAR-NK Cells: From Natural Basis to Design for Kill. *Front Immunol*. 2021;12:707542. [PubMed ID: 34970253]. [PubMed Central ID: PMC8712563]. <https://doi.org/10.3389/fimmu.2021.707542>.
56. Habib S, Tariq SM, Tariq M. Chimeric Antigen Receptor-Natural Killer Cells: The Future of Cancer Immunotherapy. *Ochsner J*. 2019;19(3):186-7. [PubMed ID: 31528126]. [PubMed Central ID: PMC6735593]. <https://doi.org/10.31486/toj.19.0033>.
57. Wrona E, Borowiec M, Potemski P. CAR-NK Cells in the Treatment of Solid Tumors. *Int J Mol Sci*. 2021;22(11). [PubMed ID: 34072732]. [PubMed Central ID: PMC8197981]. <https://doi.org/10.3390/ijms22115899>.
58. Wu X, Matosevic S. Gene-edited and CAR-NK cells: Opportunities and challenges with engineering of NK cells for immunotherapy. *Mol Ther Oncolytics*. 2022;27:224-38. [PubMed ID: 36420307]. [PubMed Central ID: PMC9676278]. <https://doi.org/10.1016/j.omto.2022.10.011>.
59. Klingemann H. Are natural killer cells superior CAR drivers? *Oncoimmunology*. 2014;3. e28147. [PubMed ID: 25340009]. [PubMed Central ID: PMC4203506]. <https://doi.org/10.4161/onci.28147>.
60. Chen KH, Wada M, Firor AE, Pinz KG, Jares A, Liu H, et al. Novel anti-CD3 chimeric antigen receptor targeting of aggressive T cell malignancies. *Oncotarget*. 2016;7(35):56219-32. [PubMed ID: 27494836]. [PubMed Central ID: PMC5302909]. <https://doi.org/10.18632/oncotarget.11019>.
61. Li H, Song W, Li Z, Zhang M. Preclinical and clinical studies of CAR-NK-cell therapies for malignancies. *Front Immunol*. 2022;13:992232. [PubMed ID: 36353643]. [PubMed Central ID: PMC9637940]. <https://doi.org/10.3389/fimmu.2022.992232>.
62. Wlodarczyk M, Pyrzynska B. CAR-NK as a Rapidly Developed and Efficient Immunotherapeutic Strategy against Cancer. *Cancers (Basel)*. 2022;15(1). [PubMed ID: 36612114]. [PubMed Central ID: PMC9817948]. <https://doi.org/10.3390/cancers15010117>.
63. Zhang L, Meng Y, Feng X, Han Z. CAR-NK cells for cancer immunotherapy: from bench to bedside. *Biomark Res*. 2022;10(1):12. [PubMed ID: 35303962]. [PubMed Central ID: PMC8932134]. <https://doi.org/10.1186/s40364-022-00364-6>.
64. Muller T, Uherek C, Maki G, Chow KU, Schimpf A, Klingemann HG, et al. Expression of a CD20-specific chimeric antigen receptor enhances cytotoxic activity of NK cells and overcomes NK-resistance of lymphoma and leukemia cells. *Cancer Immunol Immunother*. 2008;57(3):411-23. [PubMed ID: 17717662]. [PubMed Central ID: PMC1029838]. <https://doi.org/10.1007/s00262-007-0383-3>.
65. Chen KH, Wada M, Pinz KG, Liu H, Lin KW, Jares A, et al. Preclinical targeting of aggressive T-cell malignancies using anti-CD5 chimeric antigen receptor. *Leukemia*. 2017;31(10):2151-60. [PubMed ID: 28074066]. [PubMed Central ID: PMC5629371]. <https://doi.org/10.1038/leu.2017.8>.
66. Jiang H, Zhang W, Shang P, Zhang H, Fu W, Ye F, et al. Transfection of chimeric anti-CD138 gene enhances natural killer cell activation and killing of multiple myeloma cells. *Mol Oncol*. 2014;8(2):297-310. [PubMed ID: 24388357]. [PubMed Central ID: PMC5528539]. <https://doi.org/10.1016/j.molonc.2013.12.001>.
67. Chu J, Deng Y, Benson DM, He S, Hughes T, Zhang J, et al. CS1-specific chimeric antigen receptor (CAR)-engineered natural killer cells enhance in vitro and in vivo antitumor activity against human multiple myeloma. *Leukemia*. 2014;28(4):917-27. [PubMed ID: 24067492]. [PubMed Central ID: PMC3967004]. <https://doi.org/10.1038/leu.2013.279>.
68. Du Z, Ng YY, Zha S, Wang S. piggyBac system to co-express NKG2D CAR and IL-15 to augment the in vivo persistence and anti-AML activity of human peripheral blood NK cells. *Mol Ther Methods Clin Dev*. 2021;23:582-96. [PubMed ID: 34853803]. [PubMed Central ID: PMC8609108]. <https://doi.org/10.1016/j.omtm.2021.10.014>.
69. Zhang C, Burger MC, Jennewein L, Genßler S, Schönfeld K, Zeiner P, et al. Erbb2/HER2-Specific NK Cells for Targeted Therapy of Glioblastoma. *JNCI: J National Cancer Institute*. 2016;108(5). <https://doi.org/10.1093/jnci/djv375>.
70. Genssler S, Burger MC, Zhang C, Oelsner S, Mildnerberger I, Wagner M, et al. Dual targeting of glioblastoma with chimeric antigen receptor-engineered natural killer cells overcomes heterogeneity of target antigen expression and enhances antitumor activity and survival. *Oncoimmunology*. 2016;5(4). e1119354. [PubMed ID: 27141401]. [PubMed Central ID: PMC4839317]. <https://doi.org/10.1080/2162402X.2015.1119354>.
71. Jan CI, Huang SW, Canoll P, Bruce JN, Lin YC, Pan CM, et al. Targeting human leukocyte antigen G with chimeric antigen receptors of natural killer cells convert immunosuppression to ablate solid tumors. *J Immunother Cancer*. 2021;9(10). [PubMed ID: 34663641]. [PubMed Central ID: PMC8524382]. <https://doi.org/10.1136/jitc-2021-003050>.
72. Ao X, Yang Y, Li W, Tan Y, Guo W, Ao L, et al. Anti-alphaFR CAR-engineered NK-92 Cells Display Potent Cytotoxicity Against alphaFR-positive Ovarian Cancer. *J Immunother*. 2019;42(8):284-96. [PubMed ID: 31261167]. [PubMed Central ID: PMC6735933]. <https://doi.org/10.1097/CJL.0000000000000286>.
73. Sahn C, Schonfeld K, Wels WS. Expression of IL-15 in NK cells results in rapid enrichment and selective cytotoxicity of gene-modified effectors that carry a tumor-specific antigen receptor. *Cancer Immunol Immunother*. 2012;61(9):1451-61. [PubMed ID: 22310931]. [PubMed Central ID: PMC11029748]. <https://doi.org/10.1007/s00262-012-1212-x>.
74. Hu Z. Tissue factor as a new target for CAR-NK cell immunotherapy of triple-negative breast cancer. *Sci Rep*. 2020;10(1):2815. [PubMed ID: 32071339]. [PubMed Central ID: PMC7028910]. <https://doi.org/10.1038/s41598-020-59736-3>.
75. Karadimitris A. Cord Blood CAR-NK Cells: Favorable Initial Efficacy and Toxicity but Durability of Clinical Responses Not Yet Clear. *Cancer Cell*. 2020;37(4):426-7. [PubMed ID: 32289266]. <https://doi.org/10.1016/j.ccell.2020.03.018>.
76. Bachanova V, Ghobadi A, Patel K, Park JH, Flinn IW, Shah P, et al. Safety and Efficacy of FT596, a First-in-Class, Multi-Antigen Targeted, Off-the-Shelf, iPSC-Derived CD19 CAR NK Cell Therapy in Relapsed/Refractory

- B-Cell Lymphoma. *Blood*. 2021;**138**(Supplement 1):823. <https://doi.org/10.1182/blood-2021-151185>.
77. Bachanova V, Cayci Z, Lewis D, Maakaron JE, Janakiram M, Bartz A, et al. Initial Clinical Activity of FT596, a First-in-Class, Multi-Antigen Targeted, Off-the-Shelf, iPSC-Derived CD19 CAR NK Cell Therapy in Relapsed/Refractory B-Cell Lymphoma. *Blood*. 2020;**136**(Supplement 1):8. <https://doi.org/10.1182/blood-2020-141606>.
 78. Xiao L, Cen D, Gan H, Sun Y, Huang N, Xiong H, et al. Adoptive Transfer of NKG2D CAR mRNA-Engineered Natural Killer Cells in Colorectal Cancer Patients. *Mol Ther*. 2019;**27**(6):1114-25. [PubMed ID: [30962163](#)]. [PubMed Central ID: [PMC6554529](#)]. <https://doi.org/10.1016/j.ymthe.2019.03.011>.
 79. Rodriguez-Garcia A, Palazon A, Noguera-Ortega E, Powell DJ, Guedan S. CAR-T Cells Hit the Tumor Microenvironment: Strategies to Overcome Tumor Escape. *Front Immunol*. 2020;**11**:1109. [PubMed ID: [32625204](#)]. [PubMed Central ID: [PMC7311654](#)]. <https://doi.org/10.3389/fimmu.2020.01109>.

Table 2. Clinical Studies of CAR-NK Cells in Cancers ^a

Clinical Trial Identifier	Status	Clinical Trial Phase	Disease		Antigen	NK Source	Number Enrolled	Interventions	Dosage	Starting Date	Location
NCT00995137	Completed	Phase I	B-Lineage	ALL	CD19	Unknown	14	Anti-CD19 CAR-NK cells	Unknown	October 2009	United States
NCT01974479	Suspended for an interim review of (CAR) CD19 research strategy	Phase I	B-Lineage	ALL	CD19	Allogeneic	20	Anti-CD19 redirected NK cells	0.5-5×10 ⁷ /kg, and up to 1×10 ⁸ /kg	September 1, 2013	Singapore
NCT02839954	Unknown	Phase I/II	MUC1 positive r/r solid tumor		MUC1	Unknown	10	Anti-MUC1 CAR-pNK cells	Unknown	July 21, 2016	China
NCT02944162	Unknown	Phase I/II	AML		CD33	NK-92	10	Anti-CD33 CAR-NK cells	Unknown	October 2016	China
NCT03056339	Completed	Phase I/II	r/r B Lymphoid Malignancies		CD19	CB	44	iC9/CAR19/IL15-Transduced CB-NK Cells	1×10 ⁶	June 21, 2017	United States
NCT03383978	Recruiting	Phase I	Glioblastoma		HER2	NK-92	42	NK-92/5.28.z	1×10 ⁷ ; 1×10 ⁸	December 1, 2017	Germany
NCT03415100	Unknown	Phase I	Metastatic Solid Tumors		NG2D-L	Autologous or allogeneic	30	CAR-NK cells targeting NG2D ligands	Unknown	January 2, 2018	China
NCT03656705	Enrolling by invitation	Phase I	NSCLC								
NK-92	5	CCCR-NK92 cells	1×10 ⁷ ; 1×10 ⁸		September 29, 2018	China					
NCT03692663	Recruiting	Early Phase I	mCRPC		PSMA	Unknown	9	TABP EIC	0.5, 10, and 30 × 10 ⁶	December 1, 2018	China
NCT03824964	Unknown	Early Phase I	r/r B cell lymphoma		CD19/CD22	Unknown	10	Anti-CD19/CD22 CAR-NK Cells	50 - 600 × 10 ³ /kg	February 1, 2019	unknown
NCT03692637	Unknown	Early Phase I	Epithelial ovarian cancer		Mesothelin	Autologous	30	anti-Mesothelin CAR-NK Cells	0.5 - 3 × 10 ⁶ /kg	March 2019	unknown
NCT03692767	Unknown	Early Phase I	r/r B cell lymphoma		CD22	Unknown	9	Anti-CD22 CAR-NK Cells	50 - 600 × 10 ³ /kg	March 2019	unknown
NCT03690310	Unknown	Early Phase I	r/r B-Cell Lymphoma		CD19	Unknown	9	Anti-CD19 CAR-NK Cells	50 - 600 × 10 ³ /kg	March 2019	unknown
NCT03940833	Unknown	Phase I/II	MM		BCMA	NK-92	20	BCMA CAR-NK 92 cells	Unknown	May 2019	China
NCT03931720	Unknown	Phase I/II	Malignant Tumor		ROBO1	Unknown	20	BiCAR-NK/T cells (ROBO1 CAR-NK/T cells)	Unknown	May 2019	China
NCT03940820	Unknown	Phase I/II	solid tumors		ROBO1	Unknown	20	ROBO1 CAR-NK cells	Unknown	May 2019	China
NCT03941457	Unknown	Phase I/II	Pancreatic Cancer		ROBO1	Unknown	9	BiCAR-NK cells (ROBO1 CAR-NK cells)	Unknown	May 2019	China
NCT03579927	Withdrawn (lack of Funding)	Phase I/II	B-Cell Lymphoma		CD19 CD28-zeta-2A	CB	0	CD19-CD28-zeta-2A-iCasp9-IL15-transduced CB-CAR-NK cells	Unknown	October 3, 2019	United States
NCT04639739	Not yet recruiting	Early Phase I	NHL		CD19	Unknown	9	Anti-CD19 CAR-NK	2 × 10 ⁶ /kg, 6 × 10 ⁶ /kg, 2 × 10 ⁷ /kg	December 17, 2020	China
NCT05215015	Recruiting	Early Phase I	AML		CD33 and CLL1	Unknown	18	Anti-CD33/CLL1 CAR-NK	2 × 10 ⁵ ; 3×10 ⁶ ; 18×10 ⁸	November 30, 2020	China
NCT04623944	Recruiting	Phase I	Hematological Malignancies or Dysplasia		NGK2D	Allogeneic PBMC	90	CAR-NK cells targeting NGK2D ligands	1×10 ⁸ NK cells (2 ×10 ⁶ /kg for patients <50 kg), 1.5×10 ⁸ NK cells (3 ×10 ⁶ /kg for patients < 50 kg)	September 21, 2020	United States
NCT04245722	Active, not recruiting	Phase I	B-cell Lymphoma and CLL		CD19	iPSC	98	FT596 + Cyclophosphamide + Fludarabine + Rituximab + Obinutuzumab + Bendamustine	Unknown	March 19, 2020	United States
NCT04747093	Unknown	Phase I/II	B cell malignancies		CD19	Induced-T Cell Like NK	12	Induced-T Cell Like NK (CAR-ITNK cells)	Unknown	January 29, 2021	China
NCT04796675	Recruiting	Phase I	hematological malignancies.		CD19	CB	27	CAR-NK-CD19 Cells	0.01 × 10 ⁷ , 0.1 × 10 ⁷ , 1 × 10 ⁷	April 10, 2021	China
NCT04887012	Recruiting	Phase I	B-NHL		CD19	Unknown	25	Anti-CD19 CAR-NK	Unknown	May 1, 2021	China
NCT05020678	Recruiting	Phase I	B-cell Malignancies		CD19	Allogenic PBMC	150	Anti-CD19 CAR-NK Cells	3 × 10 ⁸ , 6 × 10 ⁶ /kg for patients < 50 kg	August 20, 2021	United States
NCT05248048	Recruiting	Early Phase I	metastatic colorectal cancer		NGK2D	Unknown	9	NGK2D CAR-NK Cell	Unknown	September 13, 2021	China
NCT05008536	Recruiting	Early Phase I	MM		BCMA	CB	27	Anti-BCMA CAR-NK Cells	1 - 3 × 10 ⁶ /kg, 3 - 6×10 ⁶ /kg, 0.6 - 1.2×10 ⁷ /kg	October 1, 2021	China
NCT05247957	Terminated	Phase I	AML		NGK2D	CB	9	NGK2D CAR-NK	2 × 10 ⁶ /kg, 6 × 10 ⁷ /kg, 18 × 10 ⁶ /kg	October 13, 2021	China
NCT05 379647	Recruiting	Phase I	B-Cell Malignancies		CD19	allogeneic	24	QN-019a (allogeneic CAR-NK cells targeting CD19)+ Anti-CD20 Monoclonal Antibodies	Unknown	November 4, 2021	China
NCT05020015	Recruiting	Phase 2	r/r B-cell NHL		CD19	CB	242	TAK-007	200 × 10 ⁶ , 800 × 10 ⁶	November 22, 2021	Japan

Clinical Trial Identifier	Status	Clinical Trial Phase	Disease	Antigen	NK Source	Number Enrolled	Interventions	Dosage	Starting Date	Location
NCT05137275	Recruiting	Early Phase I	Locally Advanced or Metastatic Solid Tumors	5T4	Allogeneic	56	Anti-5T4 CAR-raNK Cells	3.0×10^9 , 6.0×10^9 , 9.0×10^9	November 24, 2021	China
NCT05182073	Recruiting	Phase I	r/r Multiple Myeloma	BCMACD38	iPSC	168	CAR-NK cells with BCMA expression	Unknown	November 24, 2021	United States
NCT05213195	Recruiting	Phase I	Colorectal Cancer	NG2D	Unknown	38	NG2D CAR-NK	Unknown	December 10, 2021	China
NCT04847466	Recruiting	Phase 2	Gastric or Head and Neck Cancer	PD-L1	Unknown	55	Irradiated PD-L1 CAR-NK Cells	2×10^9	December 14, 2021	United States
NCT05008575	Recruiting	Phase I	AML	CD33	Unknown	27	Anti-CD33 CA- NK cells	6×10^8 , 12×10^8 , 18×10^8	December 23, 2021	China
NCT05194709	Recruiting	Early Phase I	advanced solid tumors	5T4)	Unknown	40	Anti-5T4 CAR-NK cells	3×10^9 , 4×10^9	December 30, 2021	China
NCT05410041	Recruiting	Phase I	B-cell malignant tumors	CD19	Unknown	15	CAR-NK-CD19 Cells	1×10^7 /kg, 2×10^7 /kg, 3×10^7 /kg	May 25, 2022	China
NCT05395052	Active, not recruiting	Phase I	advanced solid tumors	MHC) class I related proteins A (MICA) and B (MICB)	iPSC	5	FT536	Unknown	May 31, 2022	United States
NCT05410717	Recruiting	Phase I/II	advanced solid tumors (ovarian cancer and others)	CLDN6	Autologous	40	Claudin6 targeting CAR-NK cells	Unknown	June 1, 2022	China
NCT05563545	Completed	Phase I	ALL	CD19	Unknown	2	CAR-NK-CD19 Cells	1×10^7 /kg, 2×10^7 /kg, 3×10^7 /kg	July 21, 2022	China
NCT05507593	Recruiting	Phase I	SCLC	DLL3	Unknown	18	DLL3-CAR-NK cells	1×10^7 , 1×10^8 , 1×10^9	September 1, 2022	China
NCT05472558	Recruiting	Phase I	DLBCL	CD19	CB	48	Anti-CD19 CAR-NK	2×10^6 /kg, 4×10^6 /kg, 8×10^6 /kg	September 10, 2022	China
NCT05487651	Recruiting	Phase I	B-cell NHL or leukemia	CD19	Allogeneic	36	CD19-CAR-aNKT cells	1×10^7 /m ² , 3×10^7 /m ² , 1×10^8 /m ²	October 1, 2022	United States
NCT05570188	Withdrawn	Phase I/II	B Cell Hematologic Malignancies	CD19	PBMC	0	Anti-CD19 UCAR-NK cells	5×10^6 /kg, $1-2 \times 10^7$ /kg, $2-5 \times 10^7$ /kg	October 1, 2022	China
NCT05574608	Recruiting	Early Phase I	AML	CD123	Allogenic	12	CD123-CAR-NK cells	1×10^9 , $1-2 \times 10^7$ /kg	October 1, 2022	China
NCT05092451	Recruiting	Phase I/II	B-Cell Lymphoma/MDS AML	CD70	CB	94	CAR-70/IL15-transduced CB-NK cells	Unknown	November 1, 2022	United States
NCT05652530	Recruiting	Early Phase I	MM	BCMA	Allogeneic	19	BCMA CAR-NK	Unknown	November 13, 2022	China
NCT05645601	Recruiting	Phase I	B-cell Hematologic Malignancies	CD19	Allogenic	12	CD19-CAR-NK	5×10^6 /kg, 2×10^7 /kg	December 1, 2022	China
NCT05654038	Recruiting	Phase I/II	B-cell lymphoblastic Leukemia/Lymphoma	CD19	Unknown	30	Anti-CD19 UCAR-NK cells	$5-10 \times 10^6$ /kg, $1-2 \times 10^7$ /kg, $2-5 \times 10^7$ /kg	December 8, 2022	China
NCT05667155	Recruiting	Phase I	DLBCL	CD19 and CD70	CB	48	CB dualCAR-NK19/70	2×10^6 /kg, 4×10^6 /kg, 8×10^6 /kg	December 15, 2022	China
NCT05673447	Not yet recruiting	Early Phase I	Diffuse Large B Cell Lymphoma	CD19	Allogeneic	12	anti-CD19 CAR-NK cells	2×10^8 , 1×10^9 , 1.5×10^9	January 1, 2023	China
NCT05842707	Recruiting	Phase I/II	B-cell lymphoma	CD19/CD70	CB	48	Dual CAR-NK19/70 cell	Unknown	January 18, 2023	China
NCT05528341	Recruiting	Phase I	r/r solid tumors	NG2D	NK92 cells	20	NG2D-CAR-NK92 cells	0.5×10^6 /kg, 2×10^6 /kg	January 26, 2023	China
NCT05336409	Recruiting	Phase I	B-cell malignancies	CD19	Unknown	75	Anti-CD19 CAR-NK cells	Unknown	January 24, 2023	United States
NCT05739227	Recruiting	Early Phase I	r/r B-cell hematologic malignancies	CD19	Allogeneic	12	CD19-CAR-NK cells	Unknown	March 1, 2023	China
NCT05734898	Recruiting	unknown	r/r AML	NG2D	Unknown	30	NG2D CAR-NK	Unknown	March 3, 2023	China
NCT05776355	Recruiting	unknown	Ovarian cancer	NG2D	Unknown	18	NG2D CAR-NK	Unknown	March 20, 2023	China
NCT05703854	Recruiting	Phase I/II	renal cell carcinoma mesothelioma or osteosarcoma.	CD70 IL-15	CB	50	CAR-70/IL15-transduced CB-derived NK cells	Unknown	March 29, 2023	United States
NCT05110742	Not yet recruiting	Phase I/II	Hematological Malignancy	CD5 IL-15	CB	48	CAR-5/IL15-transduced CB-NK cells	1×10^7 , 1×10^8 , 1×10^9	June 30, 2023	United States

Abbreviations; PMBC, peripheral blood mononuclear cell; CB, cord blood; NSCLC, non-small-cell lung cancer; CCCR, chimeric costimulatory converting receptor; CLDN6, Claudin 6; AML, acute myeloblastic leukemia; rr, relapsed/refractory; NHL, non-Hodgkin lymphoma, MM, multiple myeloma, MDS, myelodysplastic syndromes; CLL, chronic lymphocytic leukemia; BCMA, B cell maturation antigen; PSMA, prostate-specific membrane antigen; PD-L1, programmed death-ligand 1; ROBO1, roundabout Guidance Receptor 1; 5T4, oncofetal trophoblast glycoprotein.

^a Only clinical studies with widely available information on www.clinicaltrials.gov were taken into consideration. ALL, acute lymphocytic leukemia.