



Evaluation of *Toxoplasma gondii* Antibodies in Addicted and Non-Addicted Women in Zahedan, Southeast of Iran

Javid Dehghan Haghghi  ¹, Asiyeh Hosseini  ¹, Reza Shafiei  ², Ahmad Mehravaran  ³, Ebrahim Alijani  ³ and Hadi Mirahmadi  ^{4,*}

¹Department of Community Medicine, Faculty of Medicine, Zahedan University of Medical Sciences, Zahedan, IR Iran

²Vector-Borne Diseases Research Center, North Khorasan University of Medical Sciences, Bojnurd, IR Iran

³Clinical Immunology Research Center, Zahedan University of Medical Sciences, Zahedan, IR Iran

⁴Infectious Diseases and Tropical Medicine Research Center, Resistant Tuberculosis Institute, Zahedan University of Medical Sciences, Zahedan, IR Iran

*Corresponding author: Infectious Diseases and Tropical Medicine Research Center, Resistant Tuberculosis Institute, Zahedan University of Medical Sciences, Zahedan, IR Iran.
Email: hmirahmadi59@gmail.com

Received 2020 June 14; Revised 2020 August 12; Accepted 2020 August 23.

Abstract

Background: *Toxoplasma gondii* might lead to behavioral changes in patients with toxoplasmosis. Since addicts are considered as individuals with behavioral, social, and psychiatric changes, we need to evaluate the prevalence of toxoplasmosis in addicts.

Objectives: This cross-sectional study was conducted to measure *Toxoplasma* antibodies using the enzyme-linked immunosorbent assay (ELISA) in addicted women who were kept at the Zahedan Welfare Center in 2018.

Patients and Methods: Ninety-six addicted women with high-risk behaviors and history of narcotic drug consumption as the case group and 96 non-addicted women (without a history of narcotic drug consumption) as the control group participated in this study. They had no history of alcohol consumption. The type of consumed narcotic drugs and the way of the consumption of narcotic drugs were indicated in the participants. After taking blood samples and separating the sera, the opium rapid strips were used to confirm the addiction status of the participants. Then, all sera were evaluated for *Toxoplasma* IgG and IgM antibodies using ELISA.

Results: All sera showed positive results for addition using opium rapid strips. The ELISA results indicated that the seroprevalence of *Toxoplasma* IgM in addicted women was 0.96%. Also, the seroprevalence of *Toxoplasma* IgG was identified as 37.5% in addicted women and 27.1% in the control group.

Conclusions: According to the low difference in the rate of *Toxoplasma* IgG between addicts and controls, and the low rate of *Toxoplasma* IgM, the relationship between addiction and toxoplasmosis was not proven in this study. Presumably, different factors can be involved, so more studies are needed to elucidate the reasons for the high seroprevalence of toxoplasmosis in addicts.

Keywords: Toxoplasmosis, Addicts, Enzyme-Linked Immunosorbent Assay (ELISA), Immunoglobulin M (IgM), Immunoglobulin G (IgG), Iran

1. Background

Toxoplasma gondii is a unicellular protozoan that is classified in the genus *Toxoplasma*. The *Toxoplasma* parasite infects all cells with nuclei in different hosts, including mammals, humans, and birds. This parasite leads to the induction of an infectious disease in the hosts, which is known as toxoplasmosis (1-3).

Acute toxoplasmosis occurs in the hosts due to the presence and replication of a fast-growing form of the *Toxoplasma* parasite, which is named as the tachyzoite. Provoking the innate and adaptive immune system of the host decreases the metabolism of the tachyzoite and changes it to bradyzoite. The bradyzoites are often enclosed in a cyst in different tissues, including the muscle and brain (4, 5).

Rupturing the tissue cysts and releasing the bradyzoites in infected hosts with chronic toxoplasmosis and also the presence of tachyzoites in infected hosts with acute toxoplasmosis can lead to the production of *Toxoplasma* IgG and IgM antibodies in host sera (6, 7). The identification of *Toxoplasma*-specific antibodies (IgG and IgM) and the investigation of *Toxoplasma* IgG avidity are considered the bases for the laboratory diagnosis of acute and chronic toxoplasmosis. The detection of specific antibodies against *Toxoplasma* is conducted using serological tests, including latex agglutination test, indirect fluorescence antibody test, hemagglutination test, and enzyme-linked immunosorbent assay (ELISA) (8-11). Easy access, low cost, and high sensitivity and specificity suggest ELISA as the most common

assay in the detection of *Toxoplasma*-specific antibodies.

The seroepidemiology of toxoplasmosis is estimated at 48% - 74.6% in the north, 22% - 37% in the south, 33% - 44% in the northwest, and 27% - 54% in the central areas of Iran using different serological tests (1). In recent years, some studies have revealed the presence of *Toxoplasma*-specific antibodies in blood donors, diabetic patients, pregnant women, and patients with mental disorders (1,12-17).

The *Toxoplasma* parasite might lead to the alteration of the dopaminergic signaling mediators; thus, behavioral changes may occur in patients infected with toxoplasmosis. Also, toxoplasmosis induces and progresses the central nervous system and psychiatric disorders (18,19). Since addicts are considered as individuals with behavioral, social, and psychiatric changes, we need to evaluate the prevalence of toxoplasmosis in addicts (20).

2. Objectives

This study was designed to measure *Toxoplasma* IgG and IgM antibodies using the ELISA in addicted women who were kept at the Zahedan Welfare Center in 2018.

3. Patients and Methods

A cross-sectional study was done in 2018 on 96 addicted women with high-risk behaviors who were kept at the Zahedan Welfare Center. Ninety-six healthy (non-addicted) women were selected as the negative control group. All women who participated in this study were 20-50-years-old, with histories of street life and narcotic drug consumption. None of them were addicted to alcohol products. All participants were questioned about age, education, type of consumed narcotic drug, and the way of the consumption of the narcotic drug. After filling out the questionnaires, blood samples were taken (4 mL of blood from each person) without the use of the EDTA anticoagulant. All human specimens were obtained upon the approval of the Ethics Committee of Zahedan University of Medical Sciences, Zahedan, Iran (code: IR.BUMS.REC.1396.010). Informed consent was taken from all participants in this study.

After blood sample collection, all samples were centrifuged at $1250 \times g$ for 8 min, and sera were separated. In the next step, opium rapid strips (Amazon Company, USA) were used to confirm the addiction status of the participants. To this end, 100 μ L of each serum (addicted and non-addicted groups) was poured on the opium rapid strips. The remaining sera were kept at -20°C for ELISA. All sera of addicted and non-addicted women were evaluated for the

detection of *Toxoplasma* IgG and IgM antibodies using specified ELISA kits (Pishtaz Teb Company, Iran). It has been performed based on the manufacturer's guidelines.

The SPSS software (version 20) and chi-square test were applied for analyzing the obtained data. A P value of ≤ 0.05 was considered statistically significant.

4. Results

All sera of addicted women showed positive results for addiction using opium rapid strips. However, the sera in the control group (non-addicted women) indicated negative results. Based on the kits' instructions, the observation of two marker lines on the strip was considered as a positive result.

The results showed that the seroprevalence of *Toxoplasma* IgM antibody in addicted women with high-risk behaviors was 0.96% (positive case = 1), which was not statistically comparable with the control group (positive case = 0). Also, the seroprevalence of IgG antibody was 37.5% in addicted women with high-risk behaviors and 27.1% in the control group (P value = 0.16) (Table 1).

Table 1. The Frequency of *Toxoplasma* IgM and IgG Antibody Titers in Addicted Women with High-Risk Behaviors in Comparison with the Control Group^a

| Characteristics | Values | Positive for Anti- <i>Toxoplasma</i> Antibodies | P Value |
|-----------------------|-----------|---|---------|
| Group | | | 0.16 |
| Addicted women | 96 (100) | 36 (37.5) | |
| Controls | 96 (100) | 26 (27.1) | |
| Age group | | | |
| 20 - 29 | | | 0.015 |
| Addicted women | 28 (29.1) | 2 (7.1) | |
| Controls | 27 (28.1) | 9 (33.3) | |
| 30 - 39 | | | 0.22 |
| Addicted women | 42 (43.7) | 28 (66.7) | |
| Controls | 42 (43.7) | 33 (78.6) | |
| 40 - 50 | | | 0.001 |
| Addicted women | 26 (27.2) | 20 (76.9) | |
| Controls | 27 (28.2) | 8 (29.6) | |

^aValues are expressed as No. (%).

The frequency of *Toxoplasma* IgG antibody titer in addicted women in comparison with the control group (according to age) showed that in the age range of 20 - 29 years, the positivity rate of *Toxoplasma* IgG in the control group was more than that in the addicted group. The study

results showed that there is no significant difference in the positivity rates of *Toxoplasma*-IgG antibody titer among addicted women and non-addicted women in age group 30-39 years ($P = 0.22$) (Table 1), when the high positivity rate of *Toxoplasma*-IgG titer were seen in the sera of addicted women in the age groups of 20 - 29 years ($P = 0.015$) and 40-50 years ($P = 0.001$) (Table 1).

The frequency of *Toxoplasma* IgG antibody titer in addicted women according to the way of the consumption of narcotic drugs was the highest in addicted women who were consuming the narcotic drug via both oral and inhalation ways. The result of the current study revealed the fact that the titer of *Toxoplasma*-IgG antibody in addicted women who consumed the narcotic drug via oral or inhalation routes were higher in comparison to the other routes of uses ($P = 0.03$). Also, women who were addicted to Heroin and Hashish had a higher level of *Toxoplasma*-IgG antibody in comparison to other drug users ($P = 0.008$) (Table 2). The healthy women were not selected as the control group in this section.

Data analysis showed that there is no significant difference in the *Toxoplasma*-IgG antibody level in addicted women with criminal or prison history when compared to others ($P = 0.19$) (Table 2). Addicted women with high-risk behaviors with no criminal or prison history considered as the control group.

The obtained data were not significant regarding the relationship of *Toxoplasma* IgG antibody titer and the education level in addicted women ($P = 0.53$) (Table 2).

5. Discussion

The seroprevalence of toxoplasmosis has been investigated in diabetic patients, pregnant women, and patients with mental disorders in recent years (1, 12-16). However, the seroepidemiological studies of toxoplasmosis in addicts are rare due to the specific psychological, behavioral, and social conditions of addicts and the possibility of the presence of different infectious diseases including hepatitis and acquired immunodeficiency syndrome (AIDS) in these individuals. According to the relationship between latent toxoplasmosis infection and risky behaviors leading to death, the indication of toxoplasmosis seroepidemiology is an important issue that should be considered more, especially in addiction treatment centers (20, 21).

The correlation between toxoplasmosis and ethanol consumption has been examined in ethanol consumers in a study in 2017. The study revealed a positive relationship between the presence of *Toxoplasma* IgG antibody and the high consumption of psychoactive substances such as ethanol in the consumers (20). However, they did not point out the possible causes of this correlation. Our obtained

results regarding the rate of *Toxoplasma* IgG antibody titer in addicted women were not significant in comparison to the control group. Presumably, the type of the consumed narcotic drug is effective on the rate of toxoplasmosis in addicts.

The results of a study indicated that the seroprevalence of *Toxoplasma* IgM and IgG antibodies was 0.08% and 7.7%, respectively, in intravascular narcotic drug consumers (22). Although the used samples in the mentioned study were three times in the current study, it seems that the low rate of the seroprevalence of *Toxoplasma* IgG titer refers to the way of drug consumption in addicts, which was only intravascular. In contrast, another study indicated that the intravascular injection of narcotic drugs is a major criterion in the increased rate of toxoplasmosis infection in addicts. The rate of *Toxoplasma* IgG titer was reported as 37.2% in addicts with intravascular injection in the mentioned study (23). Since our study reported the rate of *Toxoplasma* IgG titer as 37.5% in addicts that used different narcotic drugs in different ways, it seems that the type of narcotic drug and the way of consumption are not effective in the level of *Toxoplasma* IgG antibody titer in addicts.

The relationships between age, the type of narcotic drugs, and the way of the consumption of narcotic drugs and *Toxoplasma* IgG and IgM antibodies have not been investigated in previous studies; thus, the achieved results according to these factors in our study were not compared to the previous results. The results of our study showed a raised titer of *Toxoplasma* IgG antibody in the age range of 20 - 29 and 40 - 50. Therefore, the age can be considered an effective factor in the level of *Toxoplasma* IgG titer in addicts. Also, based on our results, the consumption of narcotic drugs in several ways, such as oral and inhalation, can increase the level of *Toxoplasma* IgG titer in addicts. Since our study showed the highest level of the *Toxoplasma* IgG titer in the addicts with heroin consumption, the effect of the type of the used narcotic drugs should be further investigated in future studies.

5.1. Conclusions

According to the low difference in the rate of *Toxoplasma* IgG between addicts and controls, and the low rate of *Toxoplasma* IgM, the possible relationship between addiction and toxoplasmosis was not revealed in this study. Presumably, different factors can be involved, so more studies are needed to elucidate the reason for the high seroprevalence of toxoplasmosis in addicts.

Acknowledgments

We are grateful to the Central Welfare of Zahedan for the support of sampling in this work.

Table 2. The Frequency of *Toxoplasma* IgG Antibody Titer in Addicted Women with High-Risk Behaviors According to Education Level, Way and Type of the Consumption of Narcotic Drug, and Criminal or Prison History^a

| Characteristics | Values | Positive for Anti- <i>Toxoplasma</i> Antibodies | P Value |
|--|-----------|---|---------|
| The way of consumption of narcotic drugs | | | |
| Oral | 17 (17.7) | 2 (11.8) | 0.03 |
| Inhalation | 53 (55.2) | 21 (39.6) | |
| Oral and inhalation | 26 (27.1) | 13 (50.0) | |
| The type of the consumed narcotic drug | | | |
| Opium | 29 (30.2) | 4 (13.8) | 0.008 |
| Heroin | 10 (10.5) | 6 (60) | |
| Hashish | 4 (4.2) | 0 (0) | |
| Crystal methamphetamine | 24 (25) | 12 (50) | |
| Opium and other narcotic drugs | 14 (14.5) | 7 (50) | |
| Crystal methamphetamine and other narcotic drugs | 15 (15.6) | 7 (46.7) | |
| Criminal or prison record | | | |
| Yes | 20 (20.8) | 10 (50) | 0.19 |
| No | 76 (79.2) | 26 (34.2) | |
| Education level | | | |
| Illiterate | 23 (23.9) | 8 (34.8) | 0.53 |
| Primary education | 20 (20.9) | 10 (50) | |
| Secondary education | 27 (28.1) | 11 (40.7) | |
| Diploma | 20 (20.9) | 6 (30) | |
| Higher than diploma | 6 (6.2) | 1 (16.7) | |

^aValues are expressed as No. (%).

Footnotes

Authors' Contribution: Study concept and design: Javid Dehghan Haghghi and Hadi Mirahmadi. Analysis and interpretation of data: Ahmad Mehravar and Ebrahim Aljani. Drafting of the manuscript: Asiyeh Hosseini and Reza Shafiei. Statistical analysis: Hadi Mirahmadi.

Conflict of Interests: The authors declare that this manuscript was approved by all authors in its form and no competing interest exists.

Ethical Approval: The ethical approval code was IR.BUMS.REC.1396.010.

Funding/Support: This work was supported by the Vice-Chancellors of the Zahedan University of Medical Sciences (project number: 8121).

References

- Hajsoleimani F, Ataeian A, Nourian AA, Mazloomzadeh S. Seroprevalence of *Toxoplasma gondii* in pregnant women and bioassay of IgM positive cases in Zanjan, Northwest of Iran. *Iran J Parasit.* 2012;7(2):82.
- Rahmati-Balaghaleh M, Hosseini Farash BR, Zarean M, Hatami-Pourdehno S, Mirahmadi H, Jarahi L, et al. Diagnosis of acute toxoplasmosis by IgG avidity method in pregnant women referred to health centers in south-eastern Iran. *J Parasit Dis.* 2019;43(3):517-21. doi: 10.1007/s12639-019-01120-8. [PubMed: 31406419]. [PubMed Central: PMC6667513].
- Saadatnia G, Golkar M. A review on human toxoplasmosis. *Scand J Infect Dis.* 2012;44(11):805-14. doi: 10.3109/00365548.2012.693197. [PubMed: 22831461].
- Rostami A, Riahi SM, Contopoulos-Ioannidis DG, Gamble HR, Fakhri Y, Shiadeh MN, et al. Acute *Toxoplasma* infection in pregnant women worldwide: A systematic review and meta-analysis. *PLoS Negl Trop Dis.* 2019;13(10). e0007807. doi: 10.1371/journal.pntd.0007807. [PubMed: 31609966]. [PubMed Central: PMC6822777].
- Yarovinsky F. Innate immunity to *Toxoplasma gondii* infection. *Nat Rev Immunol.* 2014;14(2):109-21. doi: 10.1038/nri3598. [PubMed: 24457485].
- Ferreira Junior A, Santiago FM, Silva MV, Ferreira FB, Macedo Júnior AG, Mota CM, et al. Production, characterization and applications for *Toxoplasma gondii*-specific polyclonal chicken egg yolk immunoglobulins. *PLoS One.* 2012;7(7). e40391. doi: 10.1371/journal.pone.0040391. [PubMed: 22808150]. [PubMed Central: PMC3395712].
- Sarkari B, Asgari Q, Bagherian N, Ashkani Esfahani S, Kalantari M, Mohammadpour I, et al. Molecular and Serological Evaluation of *Toxoplasma gondii* Infection in Reared Turkeys in Fars Province, Iran. *Jundishapur J Microbiol.* 2014;7(7). e11598. doi: 10.5812/jjm.11598. [PubMed: 25368800]. [PubMed Central: PMC4216580].

8. Mazumder P, Chuang HY, Wentz MW, Wiedbrauk DL. Latex agglutination test for detection of antibodies to *Toxoplasma gondii*. *J Clin Microbiol.* 1988;26(11):2444-6. doi: 10.1128/JCM.26.11.2444-2446.1988. [PubMed: 3235675]. [PubMed Central: PMC266914].
9. Saraei M, Shojaee S, Esmaeli A, Jahani-Hashemi H, Keshavarz H. Evaluation of confounders in toxoplasmosis indirect fluorescent antibody assay. *Iran J Parasitol.* 2010;5(4):55-62. [PubMed: 22347267]. [PubMed Central: PMC3279858].
10. Jafari Modrek M, Hasanzadeh R, Foroutan M, Mirahmadi H, Rahmati-Balaghaleh M, Zarean M. Seroprevalence and molecular evaluation of *Toxoplasma gondii* in Schizophrenic patients hospitalized in Sistan and Baluchestan province, Southeast of Iran. *Trop Biomed.* 2018;36(2):422-9.
11. Chang GN, Nemzek JA, Tjostem JL, Gabrielson DA. Simple hemagglutination inhibition test for the diagnosis of toxoplasmosis. *J Clin Microbiol.* 1985;21(2):180-3. doi: 10.1128/JCM.21.2.180-183.1985. [PubMed: 3882745]. [PubMed Central: PMC271609].
12. Jafari Modrek M, Saravani R, Mousavi M, Salimi Khorashad A, Piri M. Investigation of IgG and IgM Antibodies Against *Toxoplasma gondii* Among Diabetic Patients. *Int J Infect.* 2015;2(3). doi: 10.17795/iji27595.
13. Parlog A, Schluter D, Dunay IR. *Toxoplasma gondii*-induced neuronal alterations. *Parasite Immunol.* 2015;37(3):159-70. doi: 10.1111/pim.12157. [PubMed: 25376390].
14. Miman O, Kusbeci OY, Aktepe OC, Cetinkaya Z. The probable relation between *Toxoplasma gondii* and Parkinson's disease. *Neurosci Lett.* 2010;475(3):129-31. doi: 10.1016/j.neulet.2010.03.057. [PubMed: 20350582].
15. Ramezani M, Shojaii M, Asadollahi M, Karimilavijeh E, Gharagozli K. Seroprevalence of *Toxoplasma gondii* in Iranian patients with idiopathic Parkinson's disease. *Clin Experiment Neuroimmunol.* 2016;7(4):361-5. doi: 10.1111/cen3.12329.
16. Celik T, Kamisli O, Babur C, Cevik MO, Oztuna D, Altinayar S. Is there a relationship between *Toxoplasma gondii* infection and idiopathic Parkinson's disease? *Scand J Infect Dis.* 2010;42(8):604-8. doi: 10.3109/00365541003716500. [PubMed: 20380545].
17. Zarean M, Shafiei R, Gholami M, Fata A, Rahmati Balaghaleh M, Kariminik A, et al. Seroprevalence of Anti-*Toxoplasma Gondii* Antibodies in Healthy Voluntary Blood Donors from Mashhad City, Iran. *Arch Iran Med.* 2017;20(7):441-5. [PubMed: 28745905].
18. Abdollahian E, Shafiei R, Mokhber N, Kalantar K, Fata A. Seroepidemiological Study of *Toxoplasma gondii* Infection among Psychiatric Patients in Mashhad, Northeast of Iran. *Iran J Parasitol.* 2017;12(1):117-22. [PubMed: 28761468]. [PubMed Central: PMC5522687].
19. Gaskell EA, Smith JE, Pinney JW, Westhead DR, McConkey GA. A unique dual activity amino acid hydroxylase in *Toxoplasma gondii*. *PLoS One.* 2009;4(3). e4801. doi: 10.1371/journal.pone.0004801. [PubMed: 19277211]. [PubMed Central: PMC2653193].
20. Samojlowicz D, Borowska-Solonyko A, Kruczyk M. New, previously unreported correlations between latent *Toxoplasma gondii* infection and excessive ethanol consumption. *Forensic Sci Int.* 2017;280:49-54. doi: 10.1016/j.forsciint.2017.09.009. [PubMed: 28946032].
21. Samojlowicz D, Twarowska-Malczynska J, Borowska-Solonyko A, Poniatowski LA, Sharma N, Olczak M. Presence of *Toxoplasma gondii* infection in brain as a potential cause of risky behavior: a report of 102 autopsy cases. *Eur J Clin Microbiol Infect Dis.* 2019;38(2):305-17. doi: 10.1007/s10096-018-3427-z. [PubMed: 30470966]. [PubMed Central: PMC6514116].
22. Buchy P, Follezou JY, Lien TX, An TT, Tram LT, Tri DV, et al. [Serological study of toxoplasmosis in Vietnam in a population of drug users (Ho Chi Minh city) and pregnant women (Nha Trang)]. *Bull Soc Pathol Exot.* 2003;96(1):46-7. French. [PubMed: 12784594].
23. Massoud A, Mahaki E. A study of some opportunistic agents in IV drug abusers. *Tehran Univ Med J TUMS Publ.* 1996;54(1):3-6.