



# Phenolic Compounds and Flavonoids in Various Extracts of *Ferula tadshikorum* and *Ferula sumbul* Grown in vitro Conditions

Dilafruz Jamalova Ne'matilla Qizi <sup>1,\*</sup>, Gulsaur Kurbaniyazova Tanirbergen Kizi <sup>1</sup>, Imene Caliscan Tatar <sup>2</sup>, Atoeva Rukhsora Odilovna <sup>3</sup>, Nargiza Achilova Tuxtanazarovna <sup>4</sup>, Azima Saitova Kaljanovna <sup>5</sup>, Rabiga Esemuratova Xoshmuratovna <sup>5</sup>, Boon Chin Tan <sup>6</sup>, Azizbek Aliyor o'g'li Tog'ayev <sup>7</sup>, Ulugbek Kodirov Hamrokulovich <sup>1</sup>, Ziyoviddin Yusupov Olimjon Ugli <sup>1</sup>

<sup>1</sup> Institute of Botany of Academy of Sciences of the Republic of Uzbekistan, Tashkent, Uzbekistan

<sup>2</sup> Institute of Biological Sciences, Faculty of Science, Universiti Malaya, Kuala Lumpur, Malaysia

<sup>3</sup> Department of Biology, Faculty of Natural Sciences and Agrobiotechnology, Bukhara State University, Bukhara, Uzbekistan

<sup>4</sup> Jizzak Politechnical Institute, Jizzakh, Uzbekistan

<sup>5</sup> Karakalpak State University Named After Berdakh, Nukus, Uzbekistan

<sup>6</sup> Centre for Research in Biotechnology for Agriculture (CEBAR), Universiti Malaya, Kuala Lumpur, Malaysia

<sup>7</sup> Termez University of Economics and Service, Termez, Uzbekistan

\*Corresponding Author: Institute of Botany of Academy of Sciences of the Republic of Uzbekistan, Tashkent, Uzbekistan. Email: dilafruz.bel.91@mail.ru

Received: 17 June, 2025; Revised: 1 November, 2025; Accepted: 9 November, 2025

## Abstract

**Background:** Flavonoids, which are bioactive compounds found in medicinal plant extracts, are recognized for their significant therapeutic properties. To efficiently isolate these compounds from fresh plant materials, it is essential to optimize extraction methods. The most widely used approach involves solvent extraction using solvents such as methanol, ethanol, acetone, and hexane. This study aimed to determine the optimal extraction methods for isolating phenolic compounds and flavonoids from callus and regenerating plants of two medicinal species of the genus *Ferula* cultivated in vitro with or without indole-3-butyric acid (IBA) and 6-benzylaminopurine (BAP). This study focuses on *Ferula tadshikorum* and *Ferula sumbul*, with extracts analyzed quantitatively using high performance liquid chromatography (HPLC). The results revealed the presence of flavonoids such as kaempferol, quercetin, p-coumaric acid, and cinnamic acid at varying concentrations across different solvents. Notably, extracts from *F. sumbul* plants grown with Murashige and Skoog (MS) medium containing IBA and BAP exhibited higher levels of p-coumaric acid (9.588 µg/mL) in methanol, and kaempferol (9.595 µg/mL) in hexane. For *F. tadshikorum*, ethanol extraction yielded a significant amount of p-coumaric acid (29.9 µg/mL). Quercetin and cinnamic acid were detected in minimal quantities in both extracts, whereas kaempferol was notably higher (2.61 µg/mL) than in the methanol extract (0.617 µg/mL).

**Objectives:** The present study represents the first attempt to determine the phenolic compounds in regenerating plants obtained through in vitro methods.

**Methods:** In vitro regenerated plants of *F. tadshikorum* and *F. sumbul*, cultured on MS basal medium supplemented with IBA (0.5 mg/mL) and BAP (0.5 mg/mL), were ground into a fine powder in liquid nitrogen and then dried in a drying oven at 50°C before chemical analyses. The bioactive compounds were extracted by soaking the powders in methanol, ethanol, and hexane overnight. The extracts were filtered and evaporated under a vacuum at 35 - 36°C. The resultant slurry was partitioned with equal volumes of ethyl acetate (EA) and water to remove excess polar compounds. The EA fraction was vacuum-dried, and the mass of the crude extract was recorded. The extract was subsequently dissolved in methanol, ethanol, and hexane to achieve a concentration of 1 mg/mL. Finally, the solution was filtered through a 0.45 µm PTFE filter (German Acrodisc 13 CR) before analysis by HPLC.

**Results:** We then analyzed methanol, ethanol, and hexane extracts from in vitro *F. tadshikorum* and *F. sumbul* plants grown under various combinations of hormones and hormone-free conditions using HPLC. When comparing extracts prepared with 3 solvents from 2 different samples of the *F. sumbul*, plants obtained in a combination of IBA and BAP hormones showed a higher content of p-coumaric acid in methanol extraction (9.58 µg/mL) and a higher content of kaempferol in the hexane extraction (9.59 µg/mL). During the extraction of callus and plant regenerants from in vitro *F. tadshikorum* plants using methanol and ethanol, a higher content of p-coumaric acid (29.9 µg/mL) was observed in the ethanol extract than in methanol. Quercetin and cinnamic acid were detected in very low quantities in both the extracts. Notably, kaempferol content was higher (2.61 µg/mL) than in the methanol extract (0.617 µg/mL).

**Conclusions:** This study focused on the declining wild medicinal herbs in natural populations, providing an opportunity for systematic in vitro propagation year-round. This approach aims to establish these herbs as alternative sources for extracting biologically active secondary metabolites such as flavonoids.

**Keywords:** Flavonoids, in vitro, p-Coumaric Acid, Kaempferol, Medicinal Plants

## 1. Background

*Ferula* L., a medicinal plant species, is rich in phenolic compounds, coumarins, terpenoids, flavonoids, and essential oils, all of which demonstrate notable biological activities (1-4). Among these species, *Ferula tadshikorum* Pimenov, native to Uzbekistan and Tajikistan, is particularly diverse in its medicinal properties. Its essential oils possess mild antioxidant and antimicrobial activities, similar to those found in other sulfur-containing *Ferula* oils (5). This plant is recognized for its expectorant and anticonvulsant effects, particularly under conditions such as exudative diathesis, pulmonary tuberculosis, otitis media, and lymphadenitis (6). *Ferula* species are traditionally used to treat vitiligo, tuberculosis, joint pain, parasitic infections, gastrointestinal inflammation, and as antidotes for toxic substances. In Central Asia, gum resin is used as an anthelmintic, insecticidal, and anticonvulsant (7), as well as for treating certain nervous diseases and viral infections (8) of the reproductive system. *Ferula sumbul* (Kaufm.) Hook. f., common in Central Asia (Uzbekistan, Tajikistan), remains relatively understudied in terms of flavonoid composition. Phytochemical screening of various root extracts has revealed the presence of triterpenoids, flavonoids, coumarins, phenols, alkaloids, proteins, and carbohydrates (9). Its roots are traditionally used to relieve anxiety, serve as a sedative for stress and neurosis, provide relief for hysteria and other nervous disorders, and act as mild gastrointestinal stimulants (9, 10). Additionally, they are employed in treating kidney and stomach diseases (8).

A group of polyphenolic biologically active compounds (BAC), including flavonoids, exhibits a

diverse range of pharmacological actions. Flavonoids, such as flavonols, flavones, and anthocyanins, are secondary metabolites produced by plants in response to environmental stresses such as cold, drought, heat, salinization, ultraviolet radiation, and pathogenic microorganisms (11, 12). Recently, there has been an increase in the number of pharmacopoeial plants containing flavonoids and other polyphenolic compounds, highlighting the importance of phytochemical research and the search for new plant sources with significant pharmacological effects (13).

Quercetin and kaempferol are among the most prevalent flavonoids in plants and are known for their wide-ranging biological activities, including antioxidant, antitumor, anti-inflammatory, and anti-allergic effects (14). In cancer research, nanoquercetin has been shown to activate apoptosis in defective MCF-7 cells. Notably, quercetin can enhance the sensitivity of MCF-7 cells to the cytostatic anthracycline antibiotic doxorubicin, potentially overcoming drug resistance (15). Additionally, a treatment approach for gastric cancer has been developed using quercetin, which activates apoptosis in gastric adenocarcinoma cell lines (14). Kaempferol is a flavonoid produced in plants and is distinguished from quercetin by the absence of one hydroxyl group in its aryl structure (16). It exhibits anti-inflammatory effects through multifaceted actions on the mechanisms underlying the inflammatory processes. Kaempferol inhibits the synthesis of nitric oxide and the activity of hyaluronidase, collagenase, 15-lipoxygenase, and both cyclooxygenases (17).

Cinnamic acid and p-coumaric acid are precursors of pinocembrin and naringenin, respectively. Both flavonoids have recently garnered attention owing to

their antimicrobial, antioxidant, antitumor, and anti-inflammatory properties (18). p-Coumaric acid and its derivatives exhibit a wide range of bioactive properties, including antioxidant, antimicrobial, anticancer, anti-arthritic, anti-inflammatory, gout prevention, anti-diabetic, anti-melanogenic, skin regeneration, gastroprotective, anti-ulcer, cardioprotective, hepatoprotective, renoprotective, bone formation, anti-angiogenic, and anti-platelet properties. Given its extensive bioactivity, p-coumaric acid has potential applications in edible food, pharmaceutical, and cosmetic products. However, further studies are required to evaluate the compatibility of these products. To the best of our knowledge, this is the first study to discuss the natural occurrence, extraction, natural derivatives, synthesis of various derivatives, and therapeutic applications of p-coumaric acid (19).

## 2. Objectives

The present study aimed to isolate and quantify biologically active substances from in vitro cultures of *F. tadshikorum* and *F. sumbul* by dissolving them in various solvents. Because these species primarily contain phenolic compounds, coumarins, flavonoids, and secondary metabolites related to terpenoids, the extraction methods focused on isolating substances from these groups. This study is the first to quantify phenolic compounds in in vitro-regenerated *Ferula* species.

## 3. Methods

### 3.1. Plantlet Cultivation

Plantlets of *F. tadshikorum* and *F. sumbul* were cultivated in vitro using Murashige and Skoog (MS) medium. The media were supplemented with 0.5 mg/mL indole-3-butyric acid (IBA), 0.5 mg/mL 6-benzylaminopurine (BAP), or maintained hormone-free. Cultures were incubated at  $24 \pm 2^\circ\text{C}$  under a 16/8 h light/dark photoperiod with a light intensity of approximately 3000 lux. The plantlets were subcultured every 4 weeks to maintain vigor and growth.

### 3.2. Plant Materials

In vitro regenerated plants of *F. tadshikorum* and *F. sumbul*, cultured on MS basal medium supplemented with IBA (0.5 mg/mL) and BAP (0.5 mg/mL) (20), were

ground into a fine powder in liquid nitrogen and then dried in a drying oven at  $50^\circ\text{C}$  before chemical analyses.

### 3.3. Extraction and Purification of Bioactive Compounds

For extraction, 500 mg of each plant sample was weighed and placed into flasks. An appropriate solvent was added (e.g., methanol), and the mixture was vortexed for 1 - 2 minutes. The samples were then sonicated for 5 minutes and stored overnight at  $4^\circ\text{C}$ . The next day, samples were centrifuged at 10,000 rpm for 10 minutes at room temperature. The supernatant was collected, and the residue was re-extracted by adding fresh solvent and repeating the vortexing, sonication, storage, and centrifugation steps. The combined extracts were evaporated under vacuum at  $35 - 36^\circ\text{C}$  until complete removal of solvent and water. Ethyl acetate (EA) and distilled water (1:1) were added to the residue and thoroughly mixed. The mixture was transferred to a separatory funnel and allowed to separate into two phases. The lower aqueous phase was discarded, and the upper organic phase containing flavonoids was collected. The organic phase was evaporated under vacuum again, and the residue was weighed. Finally, methanol was added in equal proportion to the residue mass, and the extract was stored refrigerated until further analysis. Finally, the solution was filtered through a  $0.45\text{ }\mu\text{m}$  PTFE filter (German Acrodisc 13 CR) before analysis by high performance liquid chromatography (HPLC).

### 3.4. High Performance Liquid Chromatography Analysis

An injection volume of  $20\text{ }\mu\text{L}$  was applied to each sample, and the eluent was monitored at 280 nm using an HPLC system (Shimadzu) equipped with a C18 column (Shim-pack C18, 5  $\mu\text{m}$ , 4.6 mm  $\times$  250 mm) (Shimadzu, Japan). The system included a Shimadzu LC-20AT pump with a low-pressure gradient formation unit (LGPE), diode array detector (SPD-M20A), and Shimadzu Sil-20AHT autosampler operating at a flow rate of 1.5 mL/min. The solvent system consisted of (A) 0.5% (v/v) acetic acid in water, and (B) 0.5% (v/v) acetic acid in methanol. The elution gradient was as follows: 0 - 10% (B) (0 - 0.01 min); 20 - 60% (B) (0.01 - 2 min); 60 - 80% (2 - 15 min); 100% (B) (15 - 30 min); 100 - 10% (B) (30 - 35 min); and 10 - 0% (35 - 40 min). Each run lasted 40 min, and three runs were conducted for each sample. Only peak areas with a standard deviation of less than 5% were

**Table 1.** Retention Times and Concentrations of Compounds in *Ferula sumbul* Extracted Using Different Solvents<sup>a</sup>

Peak	Standard	Formula	Methanol		Ethanol		Hexane	
			Rt (min)	Concentration (µg/mL)	Rt (min)	Concentration (µg/mL)	Rt (min)	Concentration (µg/mL)
1	Quercetin	C <sub>15</sub> H <sub>10</sub> O <sub>7</sub>	11.859	0.67 ± 0.02	11.846	0.526 ± 0.07	11.884	0.592 ± 0.05
2	Kaempferol	C <sub>15</sub> H <sub>10</sub> O <sub>6</sub>	36.453	0.465 ± 0.03	36.021	2.28 ± 0.47	35.588	9.52 ± 0.21
3	p-Coumaric acid	C <sub>9</sub> H <sub>8</sub> O <sub>3</sub>	9.435	9.58 ± 0.09	9.364	3.68 ± 0.2	9.432	0.65 ± 0.06
4	Cinnamic acid	C <sub>9</sub> H <sub>8</sub> O <sub>2</sub>	12.432	0.28 ± 0.02	12.315	0.13 ± 0.016	12.346	0.21 ± 0.017

Abbreviation: Rt, retention time.

<sup>a</sup>Values are expressed as mean ± standard deviation.

recorded. Quercetin (98 - 99% for HPLC, Sigma Aldrich, USA), kaempferol (98 - 99% for HPLC, Sigma Aldrich, USA), p-coumaric acid (0320-0595, Germany), and cinnamic acid (HWI Group, Germany) were used as standards (21).

### 3.5. Statistical Analysis

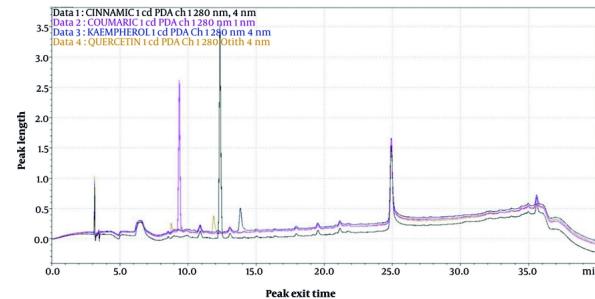
The experiments were conducted in triplicates and expressed as percentages. The data were analyzed statistically by analysis of variance (ANOVA) followed by Duncan's multiple-range test at a significance level of P < 0.05.

## 4. Results and Discussion

Flavonoids are essential active compounds in medicinal plant extracts and offer significant health benefits (22, 23). Therefore, it is crucial to optimize the flavonoid extraction method to effectively isolate these compounds from fresh plant materials. The most commonly used method is extraction using solvents (methanol, ethanol, acetone, EA, chloroform, ether, and water) (24). Quercetin (hydrate) and kaempferol are soluble in organic solvents such as ethanol, dimethyl sulfoxide (DMSO), and dimethylformamide (DMF). Quercetin (hydrate) has a solubility of approximately 2 mg/mL in ethanol and 30 mg/mL in DMSO and DMF, respectively. In comparison, the solubility of kaempferol in these solvents is approximately 11 mg/mL in ethanol, 10 mg/mL in DMSO, and 3 mg/mL in DMF. As kaempferol is slightly soluble in water, it can also be extracted using a combination of organic solvents (mainly ethanol, methanol, and acetone). Similarly, cinnamic acid, a white crystalline compound, exhibits slight solubility in water but dissolves easily in various organic solvents such as benzene, diethyl ether, and acetone, while remaining insoluble in hexane (25). In this study, we

focused on optimizing the extraction methods for quercetin, kaempferol, p-coumaric acid, and cinnamic acid. We then analyzed methanol, ethanol, and hexane extracts from *in vitro* *F. tadzhikorum* and *F. sumbul* plants grown under different conditions, including IBA (0.5 mg/mL), BAP (0.5 mg/mL), and hormone-free MS medium (20) using HPLC (Table 1).

We first determined the retention time (Rt) and peak profiles for four standards: Quercetin, kaempferol, p-coumaric acid, and cinnamic acid. The Rt values of quercetin, kaempferol at 36.453 min, p-coumaric acid at 9.435 min, and cinnamic acid were 11.859, 36.453, 9.435, and 12.432 min, respectively (Figure 1).



**Figure 1.** Chromatogram of standards (quercetin, kaempferol, p-coumaric acid and cinnamic acid)

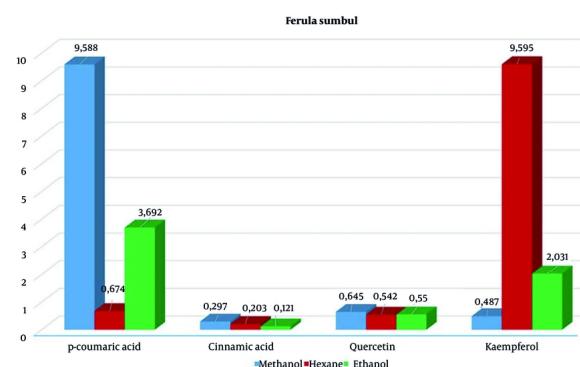
When comparing extracts prepared with 3 solvents from 2 different samples of the *F. sumbul*, plants obtained in a combination of IBA and BAP hormones (26, 27) showed a higher content of p-coumaric acid in methanol extraction (9.58 µg/mL) and a higher content of kaempferol in the hexane extraction (9.59 µg/mL) (Figure 2).

**Table 2.** Retention Times and Concentrations of p-Coumaric Acid, Cinnamic Acid, Quercetin, and Kaempferol in *Ferula tadshikorum* Extracted Using Different Solvents<sup>a</sup>

Peak	Standard	Formula	Methanol		Ethanol	
			Rt (min)	Concentration (µg/mL)	Rt (min)	Concentration (µg/mL)
1	Quercetin	C <sub>15</sub> H <sub>10</sub> O <sub>7</sub>	11.859	0.657 ± 0.03	11.846	0.726 ± 0.07
2	Kaempferol	C <sub>15</sub> H <sub>10</sub> O <sub>6</sub>	36.453	1.604 ± 0.01	36.021	2.63 ± 0.32
3	p-Coumaric acid	C <sub>9</sub> H <sub>8</sub> O <sub>3</sub>	9.435	2.44 ± 0.04	9.364	29.9 ± 0.64
4	Cinnamic acid	C <sub>9</sub> H <sub>8</sub> O <sub>2</sub>	12.432	0.262 ± 0.04	12.315	0.33 ± 0.003

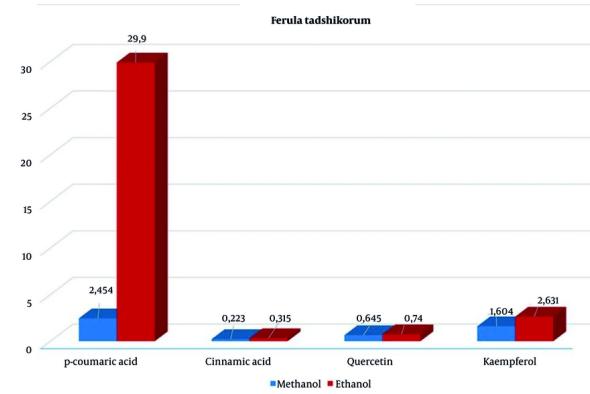
Abbreviation: Rt, retention time.

<sup>a</sup> Values are expressed as mean ± standard deviation.



**Figure 2.** Statistical chart showing p-coumaric acid, cinnamic acid, quercetin and kaempferol concentrations in *Ferula sumbul* extracts.

In contrast, plants grown in a hormone-free medium had lower levels of these compounds, with p-coumaric acid at 7.706 µg/mL and kaempferol at similar levels (Figure 2). However, the hexane extract contained significant amounts of cinnamic acid (4.09 µg/mL) and kaempferol (9.536 µg/mL). During the extraction of callus and plant regenerants from in vitro *F. tadshikorum* plants using methanol and ethanol, a higher content of p-coumaric acid (29.9 ± 0.64 µg/mL) was observed in the ethanol extract, compared to 2.44 ± 0.04 µg/mL in the methanol extract (Table 2). Quercetin (0.657 ± 0.03; 0.726 ± 0.07) and cinnamic acid (0.262 ± 0.04; 0.33 ± 0.003) were detected in very low quantities in both extracts. Notably, kaempferol content in the ethanol extract (2.63 µg/mL) was higher than in the methanol extract (1.604 µg/mL) (Figure 3).



**Figure 3.** Statistical chart of compounds in *Ferula tadshikorum* regenerants extracted with methanol and ethanol

## 5. Conclusions

This study demonstrated that solvent type and in vitro culture conditions significantly influence the extraction efficiency of phenolic compounds and flavonoids from *F. tadshikorum* and *F. sumbul*. Among the tested solvents, ethanol yielded the highest concentration of p-coumaric acid in *F. tadshikorum* (29.9 ± 0.64 µg/mL), while hexane was the most effective for extracting kaempferol from *F. sumbul* (9.59 ± 0.21 µg/mL). Quercetin and cinnamic acid were detected in low concentrations across all samples. Plants cultured with hormonal supplementation (IBA and BAP) generally produced higher levels of bioactive compounds compared to hormone-free media. These findings suggest that optimizing solvent selection and hormonal conditions is crucial for maximizing the recovery of valuable phytochemicals from *Ferula* species. Furthermore, in vitro propagation of *Ferula* species offers a sustainable alternative to wild

harvesting, helping to conserve natural populations that are increasingly threatened by overexploitation and habitat loss.

## Acknowledgements

The authors also express their sincere gratitude to the Centre for Research in Biotechnology for Agriculture (CEBAR), Universiti Malaya, for their valuable support in conducting this research.

## Footnotes

**Authors' Contribution:** Conducting the entire experiments, data collection, literature search, and manuscript writing: J. D. N. and K. G. T.; Chemical analysis: J. D. N. and I. T. C.; Formal analysis: B. Ch. T., A. N. T., and R. O. A.; Data collection: A. K. S. and E. R. X.; Writing – review and editing: J. D. N., U. H. K., and Z. O. Y. All authors have read and agreed to the published version of the manuscript.

**Conflict of Interests Statement:** The authors declare no conflict of interests.

**Data Availability:** All data is provided in the manuscript and in additional files. All data are available on request.

**Funding/Support:** The present study was funded by the project “Digital Nature: Development of a Digital Platform for the Flora of Central Uzbekistan” (Institute of Botany, Academy of Sciences of the Republic of Uzbekistan, 2025 - 2029) and partly by grant No. PRIM 01-73, “Modernization of the Institute of Botany of the Academy of Sciences of the Republic of Uzbekistan”, supported by the MUNIS project, the World Bank, and the Government of the Republic of Uzbekistan.

## References

- Batra S, Kumar A, Sharma A. Pharmacognostic and phytochemical studies on *Ferula sumbul* Hook. Roots. *J Pharmacogn Phytochem*. 2017;6(4):965-8.
- Jamalova DN, Gad HA, Akramov DK, Tojibaev KS, Musayeib NMA, Ashour ML, et al. Discrimination of the Essential Oils Obtained from Four Apiaceae Species Using Multivariate Analysis Based on the Chemical Compositions and Their Biological Activity. *Plants (Basel)*. 2021;10(8). [PubMed ID: 34451574]. [PubMed Central ID: PMC8401412]. <https://doi.org/10.3390/plants10081529>.
- Zhou P, Takaishi Y, Duan H, Chen B, Honda G, Itoh M, et al. Coumarins and bicoumarin from *Ferula sumbul*: anti-HIV activity and inhibition of cytokine release. *Phytochemistry*. 2000;53(6):689-97. [PubMed ID: 10746882]. [https://doi.org/10.1016/s0031-9422\(99\)00554-3](https://doi.org/10.1016/s0031-9422(99)00554-3).
- Sayfiddinova S, Fazlieva NT, Mukhamadiyev NQ, Mukhamadiyev AN. GC-MS chemical analysis of ferula Sumbul essential oil. *Cent Asian J Med Nat Sci*. 2023;4(3):144-50.
- Sharopov FS, Khalifaev PD, Satyal P, Sun Y, Safomuddin A, Musozoda S, et al. The Chemical Composition and Biological Activity of the Essential Oil from the Underground Parts of *Ferula tadzhikorum* (Apiaceae). *Rec Nat Prod*. 2018;13(1):18-23. <https://doi.org/10.25135/rnp.65.18.02.089>.
- Toxiri M, Korsun VF, Yaremenko KV. *[Ferula Treats Tumors and Rejuvenates the Body]*. Dushanbe, Tajikistan: Tib; 2012. RU.
- Rahmonov KS. *Biology and Resources of Ferula tadzhikorum M. Pimen. in the South of Tajikistan*[Dissertation]. Dushanbe, Tajikistan: Tajik National University; 2017.
- Egamberdieva D, Mamadalieva N, Khodjimatov O, Tiezzi A. Medicinal plants from Chatkal Biosphere Reserve used for folk medicine in Uzbekistan. *Med Aromat Plant Sci Biotechnol*. 2013;7(1):56-64.
- Batra S, Kumar A, Sharma A. Quantification of Valerenic Acid in *Ferula sumbul* Roots Using UPLC. *Indian J Pharm Sci*. 2017;79(5). <https://doi.org/10.4172/pharmaceutical-sciences.1000298>.
- Bermejo Barrera J, Gonzales AG. Chemistry and sources of mono-and bicyclic sesquiterpenes from *Ferula* species. In: Bermejo Barrera J, Gijzen HJM, Gonzales AG, de Groot A, Prota G, Wijnberg JBPA, editors. *Fortschritte der Chemie organischer Naturstoffe / Progress in the Chemistry of Organic Natural Products*. Vienna, Austria: Springer; 1995. p. 1-92. [https://doi.org/10.1007/978-3-7091-9337-2\\_1](https://doi.org/10.1007/978-3-7091-9337-2_1).
- Samanta A, Das G, Das SK. Roles of flavonoids in plants. *Carbon*. 2011;100(6):12-35.
- Del Valle JC, Buide ML, Casimiro-Soriguer I, Whittall JB, Narbona E. On flavonoid accumulation in different plant parts: variation patterns among individuals and populations in the shore campion (*Silene littorea*). *Front Plant Sci*. 2015;6:939. [PubMed ID: 26579180]. [PubMed Central ID: PMC4625047]. <https://doi.org/10.3389/fpls.2015.00939>.
- Kurkin VA, Khusainova AI, Kurkina AV, Bakova NN, Bakova E. Development of a Method of Quantitative Determination of Total Flavonoids in the Leaves of *Myrtus communis*. *Russ J Bioorg Chem*. 2022;48(7):1470-5. <https://doi.org/10.1134/s1068162022070159>.
- Chiriapkin AS, Zolotykh DS, Pozdnyakov DI. [Review of Biological Activity of Flavonoids: Quercetin and Kaempferol]. *Juvenis Sci*. 2023;9(2):5-20. RU. [https://doi.org/10.32415/jscientia\\_2023\\_9\\_2\\_5-20](https://doi.org/10.32415/jscientia_2023_9_2_5-20).
- Liao H, Bao X, Zhu J, Qu J, Sun Y, Ma X, et al. O-Alkylated derivatives of quercetin induce apoptosis of MCF-7 cells via a caspase-independent mitochondrial pathway. *Chem Biol Interact*. 2015;242:91-8. [PubMed ID: 26415619]. <https://doi.org/10.1016/j.cbi.2015.09.022>.
- Chen AY, Chen YC. A review of the dietary flavonoid, kaempferol on human health and cancer chemoprevention. *Food Chem*. 2013;138(4):2099-107. [PubMed ID: 23497863]. [PubMed Central ID: PMC3601579]. <https://doi.org/10.1016/j.foodchem.2012.11.139>.
- Lee J, Kim G. Evaluation of Antioxidant and Inhibitory Activities for Different Subclasses Flavonoids on Enzymes for Rheumatoid Arthritis. *J Food Sci*. 2010;75(7). <https://doi.org/10.1111/j.1750-3841.2010.01755.x>.

18. Tungmunnithum D, Thongboonyou A, Pholboon A, Yangsabai A. Flavonoids and Other Phenolic Compounds from Medicinal Plants for Pharmaceutical and Medical Aspects: An Overview. *Medicines (Basel)*. 2018;5(3). [PubMed ID: 30149600]. [PubMed Central ID: PMC6165118]. <https://doi.org/10.3390/medicines5030093>.
19. Kaur J, Kaur R. p-Coumaric Acid: A Naturally Occurring Chemical with Potential Therapeutic Applications. *Curr Org Chem*. 2022;26(14):1333-49. <https://doi.org/10.2174/138527282666221012145959>.
20. Jamalova DN, Kurbaniyazova GT, Mustafina FU. Development of an effective method of somatic embryogenesis for obtaining biologically active substances from an endangered medicinal plant (f. Sumbul (kauffm.) hook. F.). *Eurasian J Appl Biotechnol*. 2024;3(3). <https://doi.org/10.1134/symposium.17>.
21. Ishii K, Furuta T, Kasuya Y. High-performance liquid chromatographic determination of quercetin in human plasma and urine utilizing solid-phase extraction and ultraviolet detection. *J Chromatogr B Analyt Technol Biomed Life Sci*. 2003;794(1):49-56. [PubMed ID: 12888197]. [https://doi.org/10.1016/s1570-0232\(03\)00398-2](https://doi.org/10.1016/s1570-0232(03)00398-2).
22. Liew YJM, Lee YK, Khalid N, Rahman NA, Tan BC. Cell-Free Expression of a Plant Membrane Protein BrPT2 From Boesenbergia Rotunda. *Mol Biotechnol*. 2021;63(4):316-26. [PubMed ID: 33565047]. <https://doi.org/10.1007/s12033-021-00304-z>.
23. Karlson CKS, Mohd Noor SN, Khalid N, Tan BC. CRISPRi-Mediated Down-Regulation of the Cinnamate-4-Hydroxylase (C4H) Gene Enhances the Flavonoid Biosynthesis in Nicotiana tabacum. *Biology (Basel)*. 2022;11(8). [PubMed ID: 36009753]. [PubMed Central ID: PMC9404795]. <https://doi.org/10.3390/biology11081127>.
24. Zhao B, Fu Y, Wang W, Zu Y, Gu C, Luo M, et al. Enhanced extraction of isoflavonoids from Radix Astragali by incubation pretreatment combined with negative pressure cavitation and its antioxidant activity. *Innov Food Sci Emerg Technol*. 2011;12(4):577-85.
25. Temidayo AR. Extraction and isolation of flavonoids present in the methanolic extract of leaves of Acanthospermum hispidum Dc. *Glob J Med Plant Res*. 2013;1(1):111-23.
26. Jamalova D, Mustafina F, Kurbaniyazova G, Turdiev D. [The role of Exogenous Phytohormones as a Key in vitro Factor in the Morphogenesis of Some Species of the Ferula L. (Apiaceae Lindl.) Genus]. *Biotehnologiya*. 2022;4:46-55. RU. <https://doi.org/10.56304/s0234275822040160>.
27. Mustafina FU, kizi Jamalova DN, Zarekarizi AR, kizi Juraeva HK, ogli Khazratov AT, Hoe Jin K, et al. Optimized microclonal propagation protocol and antioxidant activity of callus cultures from the endangered Ferula tadzhikorum Pimenov (Apiaceae Lindl.). *Plant Cell Tissue Organ Cult*. 2025;161(2). <https://doi.org/10.1007/s11240-025-03048-9>.