



Reverse Transcription Loop-Mediated Isothermal Amplification Assay with High Sensitivity to Rapid Detection of Viable *Salmonella* in Foods

Dong Chen ¹, Fangju Tian ², Wanyu Liu ², Jingyi Yu ³ and Dafeng Song ^{2,*}

¹Hangzhou Anxu Biological Technology Co., Hangzhou, China

²Zhejiang Gongshang University, Hangzhou, China

³School of Accounting for Zhejiang Gongshang University, Hangzhou, China

*Corresponding author: Key Laboratory for Food Microbial Technology of Zhejiang Province, Zhejiang Gongshang University, Hangzhou, China. Email: dfsong@zjsu.edu.cn

Received 2021 July 26; Revised 2021 December 19; Accepted 2021 December 19.

Abstract

Background: *Salmonella* is one of the main foodborne bacterial pathogens, causing diseases and death. The study used reverse transcription loop-mediated isothermal amplification (RT-LAMP) to detect *Salmonella*.

Objectives: To design six primers and detect *Salmonella* using RT-LAMP to facilitate the rapid detection of pathogenic bacteria in food.

Methods: We designed six primers based on the gene coding sequences of *inv A*, specific to *Salmonella*. Each reaction solution contained 6.0 mM MgSO₄, 1 M betaine, 1.6 mM dNTPs, 160 U/mL Bst DNA polymerase, 0.2 μ M of both external primers, 0.8 μ M of both internal primers, and 0.2 μ M of both loop primers. The reaction temperature was 65°C.

Results: Our amplified products were separated by 2% agarose gel electrophoresis. The detection limit was 10 CFU per reaction.

Conclusions: RT-LAMP exhibited the same accuracy as the Chinese National Standard assay (GB 4789.30-2016) assay in detecting *Salmonella* in foods. RT-LAMP was highly specific and sensitive; hence, it may serve as an effective tool in detecting *Salmonella*.

Keywords: *Salmonella* D012475, RT-LAMP, Foodborne Pathogens, Rapid Detection

1. Background

Salmonella is one of the main foodborne pathogens in the *Enterobacteriaceae* family. *Salmonella* spp. cause diarrhea, vomiting, abdominal colic, pneumonia, and bacteremia (1). On the other hand, in the cutting-edge technology of bacterial therapy, scientists utilize bacteria such as *Salmonella typhimurium* and *Shigella flexneri* as antiproliferative and potential chemotherapeutic agents to treat gastrointestinal cancers (2, 3). Livestock and poultry products are susceptible to *Salmonella* contamination. Foods leading to disease outbreaks are mainly eggs and poultry meat; however, the predominant foodborne diseases are enteritis and typhoid fever (4, 5). Due to its excellent survival characteristics, *Salmonella* is found in various fresh vegetables and meat products. When adhered to the surface of heat-treated foods, the bacteria rapidly multiply even at low temperatures (6). Accordingly, the detection of *Salmonella* is of utmost importance in the food industry (7).

Routine culture is the gold standard for detecting, isolating, and identifying *Salmonella* spp. However, the cultural process is time-consuming. Samples should be

placed in an enrichment solution for 18 - 24 h, transferred to chromogenic plates for 18 - 24 h, and purified for 18 - 24 h. Following these steps, the pathogen can be preliminarily and serologically identified before its identity is confirmed. In this regard, it takes 6 - 7 days to reach results, and specialized laboratories are required (8). Compared with routine culture, polymerase chain reaction (9), DNA microarrays (10), and antibody assays (11) are more appropriate detection methods; however, they are time-consuming. Accordingly, we need a more efficient and straightforward protocol.

In 2000, loop-mediated isothermal amplification (LAMP) was proposed as a novel approach to molecular identification (12). LAMP is specific, sensitive, and cost-effective. However, it fails to distinguish between live and dead cells, and this method often obtains false-positive results from food samples (13). Accordingly, a more accurate method is required to detect and monitor pathogens. Studies have reported that reverse transcription LAMP (RT-LAMP) may overcome the LAMP limitations. RT-LAMP can be used to detect viruses (14). In this study, we collected samples from Hangzhou, Zhejiang Province, China, and

used RT-LAMP to detect *Salmonella* to facilitate the rapid detection of pathogenic bacteria in food.

2. Objectives

To design six primers and detect *Salmonella* using RT-LAMP to facilitate the rapid detection of the pathogen in food.

3. Methods

3.1. Strains and Culture

We collected 41 strains from different areas in Hangzhou, China. Ten *Salmonella* spp. were used as positive samples; other bacterial species were selected to evaluate the specificity of RT-LAMP (Table 1). *Salmonella* strains were stored in the LB broth/agar medium, and additional species such as *Listeria* were stored in the BHI broth/agar medium. The bacterial solutions were cultured at 37°C and diluted. We measured bacterial concentrations by ultraviolet spectrophotometry. All chemicals and media were acquired from Sangon Co., Ltd. (Shanghai, China).

3.2. RNA Extraction and cDNA Synthesis

We extracted total RNA from all 41 bacterial strains using an RNA Rapid Extraction Kit (Sangon Co., Ltd. Shanghai, China) and measured yield using a NanoDrop spectrophotometer (Eppendorf, Germany). To minimize the RNA degradation, total RNA was converted into cDNA using a PrimeScript II First Strand cDNA Synthesis Kit (TaKaRa, China) (15). The reaction volume (20 μ L) consisted of 6 μ L of RNA, 4 μ L of 5 \times RT buffer, 2 μ L of dNTPs (10 mM each), 50 pmol of random hexamers, 10 U of AMV reverse transcriptase XL (TaKaRa), and 10 U of RNase inhibitor (TaKaRa). The reaction solution was maintained for 60 min at 42°C and 15 min at 70°C.

3.3. Primer Design

We designed primers based on an internally transcribed spacer sequence (GenBank Accession No. NC_987035) using Primer Explorer V5 (Eiken Chemical Co. Ltd., Tokyo, Japan). Six pairs of primers were created, which targeted distinct regions of the LAMP sequence and corresponded to two outlying primers (F3 and B3), two internal primers (FIP and BIP), and two loop primers. Table 2 and Figure 1 show the primer sequences and locations, respectively.

Table 1. Bacterial Strains Used in this Study

Types and Species	Strains
Target- <i>Salmonella</i> spp.	
<i>Salmonella enterica</i>	ZFM New1
<i>S. enterica</i>	CGMCC 1.1552
<i>S. enterica</i>	CGMCC 1.10603
<i>S. typhimurium</i>	CGMCC 1.1174
<i>S. typhimurium</i>	CGMCC 1.1190
<i>S. pullorum</i>	ZFM 124694
<i>S. anatum</i>	ZFM New2
Non- <i>Salmonella</i> spp.	
<i>Micrococcus luteus</i>	CGMCC 1.193
<i>Staphylococcus aureus</i>	ATCC 6538P
<i>S. aureus</i>	CGMCC 1.2386
<i>S. aureus</i>	CGMCC 1.879
<i>S. aureus</i>	CGMCC 1.128
<i>Lactobacillus plantarum</i>	CGMCC 1.11
<i>L. plantarum</i>	CGMCC 1.124
<i>L. plantarum</i>	CGMCC 1.551
<i>L. plantarum</i>	CGMCC 1.556
<i>L. plantarum</i>	CGMCC 1.511
<i>L. lactis</i>	ATCC 15577
<i>Bacillus subtilis</i>	CGMCC 1.1627
<i>Enterococcus faecalis</i>	ZFM BM3
<i>E. faecalis</i>	CGMCC 1.125
<i>Shigella flexneri</i>	CGMCC 1.1868
<i>Pseudomonas aeruginosa</i>	CGMCC 1.647
<i>S. dysenteriae</i>	ATCC 9753
<i>Escherichia coli</i>	CGMCC 1.1580
<i>E. coli</i>	O104
<i>E. coli</i>	JM109
<i>Listeria seeligeri</i>	ZFM 51335
<i>L. monocytogenes</i>	ATCC 7648
<i>P. putida</i>	CGMCC 1.645
<i>Rhodotorula Rubra</i>	CGMCC 2.1034
<i>Saccharomyces cerevisiae</i>	CGMCC 2.1643

Abbreviations: ATCC, American Type Culture Collection; CGMCC, China General Microbiological Culture Collection Center; ZFM, Key Laboratory of Food Microbiology Research, Zhejiang Province of China.

3.4. LAMP Assay

The following six primer pairs were added to a 25- μ L tube: 1.6 mM dNTPs, 0.8 M betaine (Sigma-Aldrich, USA), 20 mM Tris-HCl (pH 8.8), 10 mM KCl, 10 mM (NH4)2SO4, 4 mM

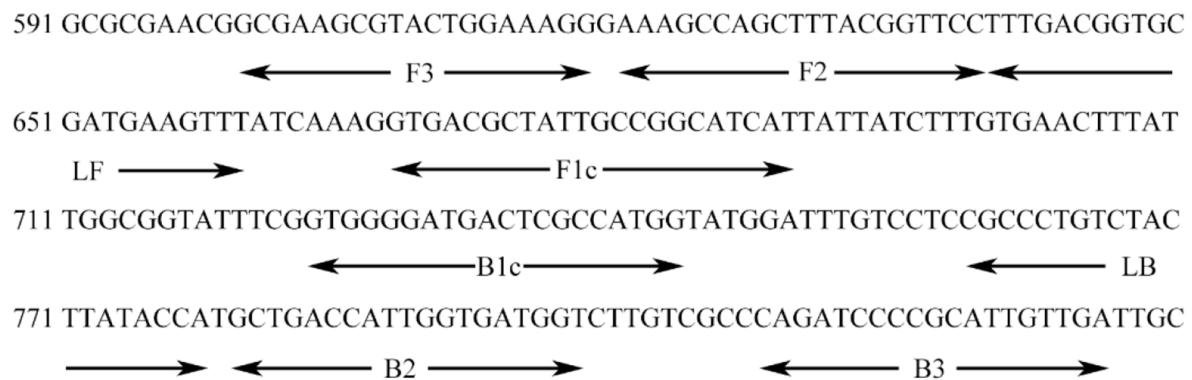


Figure 1. Sequence of *inv A* gene for detecting *Salmonella* and primer locations. The sequences of each primer-binding site are provided under the line.

Table 2. Primers Positions of *inv A* Gene Coding Sequences of *Salmonella*

Primer Name	Type	Sequences (5' - 3')	Length (bp)
F3	Forward outer	GCGAACGGTACTGGAAAGG	19
B3	Reverse outer	TCAACAATGCCGGGATCTG	19
FIP	Composed of F1c and F2	ATGATGCCGGATAGCGTCACAAAGCCAGCTTACGGTCC	40
BIP	Composed of B1c and B2	GTGGGATGACYCGCATGGACCATCACCAA ^a	38
LF		AAACTTCATCGCACCGTCAA	21
LB		CCGCYCTGTCTACTTATACCA	21

MgSO₄, 0.1% Triton X-100, 80 U Bst DNA polymerase (New England Biolabs, USA), and 2 μ L of target cDNA. Different assay conditions, including different temperatures (59 to 65°C) and concentrations of MgSO₄ (2 to 8 mM), dNTPs (0.8 to 2.0 mM), and Bst DNA polymerase (40 to 320 U), were evaluated. The reaction solution was incubated for 1 h and 5 min at 80°C (16, 17). All experiments were performed in triplicate.

3.5. Detection of RT-LAMP Products

The amplified products were separated using 2% agarose gel electrophoresis and analyzed under ultraviolet light after 40 min. A white precipitate (magnesium pyrophosphate) was produced during the reaction, which was visible with the naked eye.

3.6. Specificity of RT-LAMP

We used the RNA templates of the 41 strains to perform RT-LAMP. A total of 25 non-*Salmonella* strains were tested to assess potential cross-reactions (Table 1). The other strains were used for determining the specificity of the assay.

3.7. Sensitivity of RT-LAMP

We determined the detection limit of RT-LAMP by preparing a 10-fold dilution series (106 CFU/ μ L to 100

CFU/ μ L) of *Salmonella* (ZFM New1) in sterile phosphate-buffered saline. The strain was measured by plating it onto LB agar directly (Huankai, Guangzhou, China) followed by incubation at 37°C for 24 h. The RNA templates were obtained from 100 μ L of each dilution, and 2 μ L fractions were amplified using RT-LAMP. Each sensitivity test was performed four times.

3.8. *Salmonella* Detection in Ready-to-Eat Meats

We used the RT-LAMP assay to detect *Salmonella* spp. in ready-to-eat (RTE) meats and other foods and compared the results with those obtained from the Chinese National Standard assay (GB 4789.30-2016). The RTE meats were obtained from a local market in China. We divided each sample into 25-g portions, and each portion was isolated for the RNA extraction to act as a template for RT-LAMP. The GB assay was performed by incubating 25 g of each sample at 30°C for 24 h in 225 mL of Listeria enrichment broth I (LB1). In the next phase, we incubated 100 μ L of the samples in 10-mL of Listeria enrichment broth II (LB2) at 30°C for 24 h. Positive LB2 cultures aliquots (50 μ L) were inoculated on PALCAM agar (Huankai) and Listeria chromogenic agar (Huankai) and incubated at 37°C for 48 h. To confirm the presence of *Salmonella* spp., we used characteristic colony morphologies, Gram staining, and additional tests, includ-

ing sugar and motility tests. The strains were finally identified using API Listeria (bioMérieux, France).

3.9. Statistical Analysis

SPSS and Origin were used to analyze the experimental data and process the figures.

4. Results

4.1. Optimization of RT-LAMP Reaction Conditions

We optimized the RT-LAMP conditions by evaluating different reaction conditions. The optimal Mg and dNTP concentrations were 6.0 mM MgSO₄ (Figure 2A) and 1.6 mM dNTPs (Figure 2C), respectively. Figure 2B shows a minor difference in temperature (63 and 65°C). No positive product was amplified at 40 U/mL Bst DNA polymerase, while the optimal results were obtained at 160 U/mL Bst DNA polymerase (Figure 2D). Accordingly, optimum conditions were 6.0 mM MgSO₄, 1.6 mM dNTPs, and 160 U/mL Bst DNA polymerase at 65°C.

4.2. Detection of RT-LAMP Products

Our amplified products were separated by 2% agarose gel electrophoresis (Figure 3A), and the strains were observed with the naked eye (Figure 3B). We noted turbidity changes under normal lighting conditions. Positive cDNA samples were successfully amplified. The results revealed that *Salmonella* could be rapidly and easily detected using RT-LAMP.

4.3. Specificity of RT-LAMP Assay

Primer specificity (Figure 1) was investigated by extracting cDNA from eight strains of *Salmonella* and incorporating 33 strains of other species as templates (Table 1). Each of the eight *Salmonella* strains (Figure 4) was successfully amplified, whereas none of the other species were amplified. Accordingly, our primers were highly specific to *Salmonella*.

4.4. Sensitivity of RT-LAMP Assay

We performed sensitivity analysis by incorporating 10-fold serial dilutions of *Salmonella*, corresponding to 500 ng, 50 ng, 5 ng, 0.5 ng, 50 pg, 5 pg, 0.5 pg, and 50 fg (equivalent to 1 × 10⁶ CFU/μL to 1 × 100 CFU/μL) of RNA. The RT-LAMP assay resulted in the DNA bands at all dilutions. However, the bands became weaker with decreasing RNA concentration. The sensitivity limit was 50 fg (Figure 5), equivalent to 1 CFU/μL.

4.5. Detection of viable *Salmonella* in RTE Food Samples

We assessed the capability of the RT-LAMP assay in detecting pathogenic microorganisms in the RTE foods, including a variety of vegetables, fruits, and meats (Table 3). The RT-LAMP and GB assays detected two *Salmonella*-positive chicken samples. Similar results were obtained for the other three types of food, indicating that the RT-LAMP and GB assays had similar detection capabilities.

5. Discussion

Microbial contaminants have long been a health concern. Food poisoning has been studied for a long time, and plants are fully used as drugs in traditional treatment methods (18). In recent years, the abuse of antibiotics has led to the rapid spread of antibiotic resistance in pathogens (19) as such, it is necessary to change the traditional treatment strategy. Experts have conducted in-depth studies on the pathogenicity and defense mechanisms. Hassan Mahmoudi found out that the pathogenicity of *Escherichia coli* with *iuc A* and *iut A* genes may be associated to this issue (20). Propolis and garlic water extract can play a role in preventing pathogenic bacteria (21, 22). In the present study, a rapid detection method was designed for pathogenic bacteria in food to expand food safety methods further.

Developing detection methods for *Salmonella*, one of the most common contaminants in fruits and meats, is vital to minimize outbreaks (23). From a food safety and human health perspective, detecting foodborne bacterial pathogens requires immediate attention. This study proposed the use of RT-LAMP for detecting *Salmonella* in foods. The assay had high specificity and sensitivity in detecting *Salmonella*. Moreover, RT-LAMP was accurate and fast enough to be used in routine inspections and quarantine situations. Accordingly, it may serve as a potential diagnostic tool in the food industry to detect *Salmonella* in food products rapidly.

The proposed detection method of *Salmonella* uses *inv A* as the target gene, specific to *Salmonella*. Previous studies on *inv A*-based molecular detection assays have demonstrated their high specificity and sensitivity (24). When bacteria die, RNA cleaves quickly. Accordingly, the RT-LAMP assay for detecting *Salmonella* has lower false-positive results. Mg²⁺ concentration has the largest impact on primer annealing and DNA polymerase activity. A preliminary study revealed no target amplification at high Mg²⁺ concentrations since Mg²⁺ is bound to chelating agents such as ethylenediaminetetraacetic acid and anionic groups. Hence, the DNA polymerase activity and amplification results were affected. After evaluating a range of concentrations (2 - 8 mM), we found that 6 mM MgSO₄ was optimal,

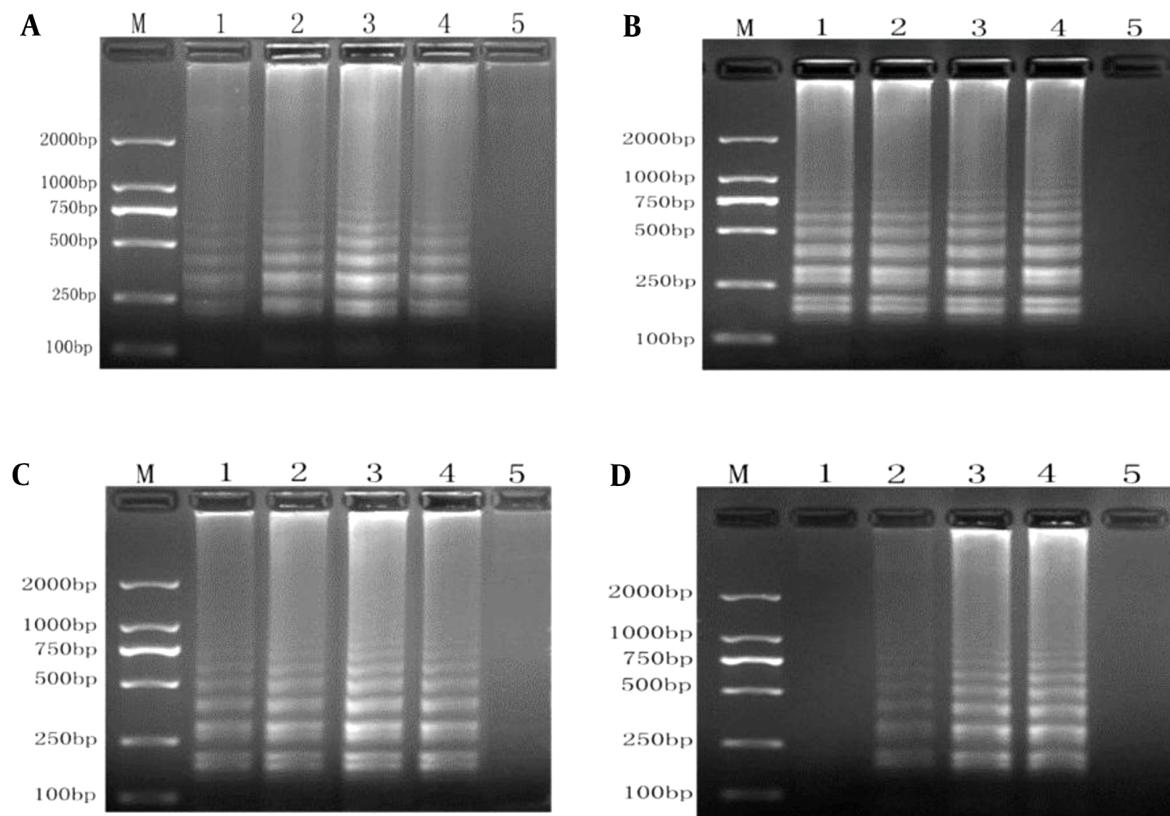


Figure 2. RT-LAMP system optimization diagram. A, optimization of Mg^{2+} concentrations (lane M, 2,000-bp ladder marker; lanes 1 - 4, the results of RT-LAMP with Mg^{2+} concentrations of 2.0 mmol/L, 4.0 mmol/L, 6.0 mmol/L, and 8.0 mmol/L; lane 5, negative control); B, optimization of annealing temperatures (lane M, 2000-bp ladder marker; lanes 1 - 4, the results of RT-LAMP at 59, 61, 63, and 65°C; lane 5, negative control); C, optimization of dNTP concentrations (lane M, 2000-bp ladder marker; lanes 1 - 4, the results of RT-LAMP with dNTP concentrations of 0.8 mmol/L, 1.2 mmol/L, 1.6 mmol/L, and 2.0 mmol/L; lane 5, negative control); D, optimization of Bst enzyme concentrations (lane M, 2,000-bp ladder marker; lanes 1-4, the results of RT-LAMP with Bst enzyme concentrations of 40, 80, 160, and 320 U/mL; lane 5, negative control).

Table 3. Detection of *Salmonella* in RTE Foods Using RT-LAMP and GB 4789.4-2016 Assay

Sample (Tested Number)	No. of Samples Tested Positive for <i>Salmonella</i>	
	RT-LAMP	GB 4789.4-2016
Beef (25)	0	0
Pork (25)	1	1
Egg (25)	2	2
Fish (25)	0	0
Total (100)	3	3

indicating that the Bst DNA polymerase was effective at 61 - 65°C. We selected 65°C as the optimal temperature based on the AT and GC contents of the primers. The number of *inv A* mRNA copies was significantly higher than the number of *inv A* genes in the cell. Copy number is the major determinant of sensitivity.

5.1. Conclusions

The findings revealed that the detection limit for *Salmonella* in RT-LAMP was 1 CFU per reaction in pure cul-

tures, which was 10-fold lower than that in PMA-LAMP (25). As an alternative to PCR, RT-LAMP is more sensitive, may be superior to PMA-LAMP in detecting *Salmonella*, and is further used to detect microorganisms in the RTE meats. Compared to GB 4789.4-2016, it was accurate; however, it took considerably less time.

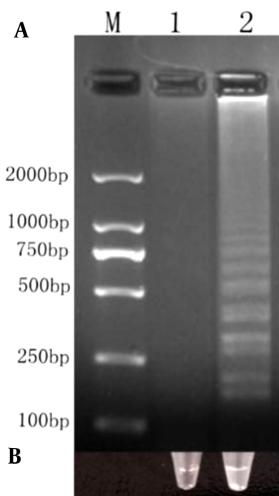


Figure 3. Detection of RT-LAMP products. A, lane M, 2000-bp ladder marker; lane 1, negative control; lane 2, the results of LAMP optimization reaction; B, physical map of corresponding lanes.

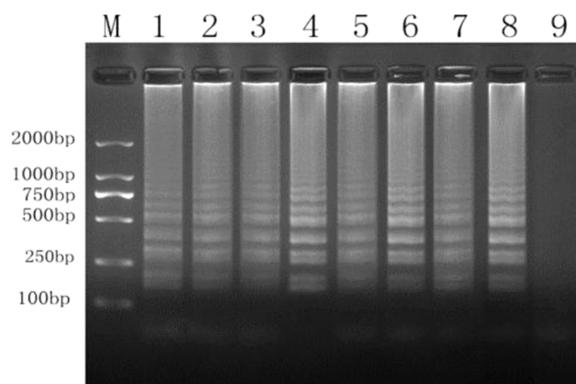


Figure 4. Specificity test of RT-LAMP assay (lane M, 2,000-bp ladder marker; lane 1, WSI; lane 2, F2365; lane 3, F8027; lane 4, J1723; lane 5, HF550; lane 6, J4045; lane 7, clip11262; lane 8, J0161; and lane 9, negative control).

Footnotes

Authors' Contribution: DongChen conducted the experiments and prepared the first draft of the manuscript. Fangju Tian, Wanyu Liu, and Dafeng Song performed the molecular biology experiments. Dong Chen and Jingyu Yu contributed to the manuscript's design, analysis, and preparation. All authors studied and approved the final manuscript.

Conflict of Interests: All authors declare that there is no conflict of interests regarding the publication of this manuscript.

Ethical Approval: This study was approved by the Food

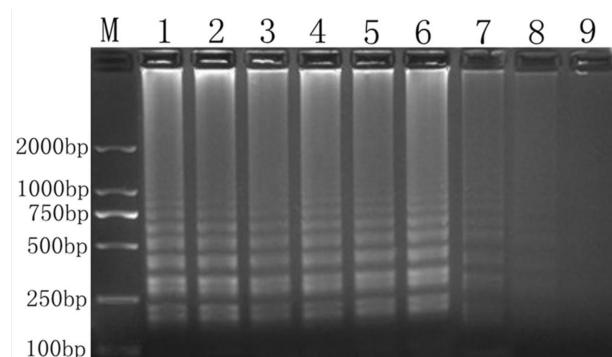


Figure 5. Sensitivity test of RT-LAMP assay. Lane M, 2000-bp ladder marker; lanes 1-8, diluted samples of *Salmonella* with 4.36×10^7 , 4.36×10^6 , 4.36×10^5 , 4.36×10^4 , 4.36×10^3 , 4.36×10^2 , 4.36×10^1 , and 4.36 ; lane 9, negative control

Microbial Technology Ethics Committee of the Zhejiang Gongshang University.

Funding/Support: Our study was supported by the Zhejiang Province key research and development program, Lingyan Plan- automatic nucleic acid detection all-in-one machine and ultra-high sensitivity monocyte gene listeria detection kit development (NO.2022C03091).

References

1. Bayrac C, Eyidogan F, Avni Oktem H. DNA aptamer-based colorimetric detection platform for *Salmonella Enteritidis*. *Biosens Bioelectron*. 2017;98:22-8. doi: [10.1016/j.bios.2017.06.029](https://doi.org/10.1016/j.bios.2017.06.029). [PubMed: 28646719].
2. Khodavirdipour A, Rouhani nejad H, Zandi M, Khodavirdipour A, Khayyam N. To study in vitro anti-proliferative and pro-apoptotic properties of *Salmonella typhi* in human pancreatic cancer cell line. *Avicenna J Clin Microbiol Infect*. 2019;6(3):77-82. doi: [10.34172/ajcmi.2019.13](https://doi.org/10.34172/ajcmi.2019.13).
3. Khodavirdipour A, Jamshidi F, Nejad HR, Zandi M, Zarean R. To study the anti-cancer effects of *Shigella flexneri* in AspC-1 pancreatic cancer cell line in approach to bax and bcl-2 genes. *J Gastrointest Cancer*. 2021;52(2):593-9. doi: [10.1007/s12029-020-00433-9](https://doi.org/10.1007/s12029-020-00433-9). [PubMed: 32524303].
4. Jarquin R, Hanning I, Ahn S, Ricke SC. Development of rapid detection and genetic characterization of salmonella in poultry breeder feeds. *Sensors (Basel)*. 2009;9(7):5308-23. doi: [10.3390/s90705308](https://doi.org/10.3390/s90705308). [PubMed: 22346699]. [PubMed Central: PMC3274138].
5. Ocana-de Jesus RL, Gutierrez-Ibanez AT, Sanchez-Pale JR, Mariezcurrena-Berasain MD, Laguna-Cerda A, Hernandez-Chinas U, et al. Motility and survival of *Salmonella enterica* subspecies enterica serovar enteritidis in tomato plants (*Solanum lycopersicum* L.). *Int Microbiol*. 2019;22(3):363-8. doi: [10.1007/s10123-019-00059-3](https://doi.org/10.1007/s10123-019-00059-3). [PubMed: 30811003].
6. Burnett SL, Chopskie JH, Podtburg TC, Gutzmann TA, Gilbreth SE, Bodnaruk PW. Use of octanoic acid as a postlethality treatment to reduce *Listeria monocytogenes* on ready-to-eat meat and poultry products. *J Food Prot*. 2007;70(2):392-8. doi: [10.4315/0362-028x-70.2.392](https://doi.org/10.4315/0362-028x-70.2.392). [PubMed: 17340874].
7. Khan I, Khan J, Miskeen S, Tango CN, Park YS, Oh DH. Prevalence and control of *Listeria monocytogenes* in the food industry - a review. *Czech J Food Sci*. 2016;34(No. 6):469-87. doi: [10.17221/21/2016-cjfs](https://doi.org/10.17221/21/2016-cjfs).

8. Liu A, Shen L, Zeng Z, Sun M, Liu Y, Liu S, et al. A Minireview of the Methods for Listeria monocytogenes Detection. *Food Anal Methods*. 2017;11(1):215–23. doi: [10.1007/s12161-017-0991-2](https://doi.org/10.1007/s12161-017-0991-2).
9. Shoukat S, Ali M, Wani H, Ali U, Para PA, Ganguly S. Food-borne listeriosis, an upcoming public health problem: A review. *J Immunol Immunopathol*. 2017;19(1):1–9. doi: [10.5958/0973-9149.2017.00001.6](https://doi.org/10.5958/0973-9149.2017.00001.6).
10. Dzupova O, Rozsypal H, Smiskova D, Benes J. Listeria monocytogenes meningitis in adults: the Czech Republic experience. *Biomed Res Int*. 2013;2013:846186. doi: [10.1155/2013/846186](https://doi.org/10.1155/2013/846186). [PubMed: [24106719](https://pubmed.ncbi.nlm.nih.gov/24106719/)]. [PubMed Central: [PMC3784069](https://pubmed.ncbi.nlm.nih.gov/PMC3784069/)].
11. Savas S, Ersoy A, Gulmez Y, Kilic S, Levent B, Altintas Z. Nanoparticle enhanced antibody and DNA biosensors for sensitive detection of *Salmonella*. *Materials (Basel)*. 2018;11(9). doi: [10.3390/matl091541](https://doi.org/10.3390/matl091541). [PubMed: [30150524](https://pubmed.ncbi.nlm.nih.gov/30150524/)]. [PubMed Central: [PMC6163637](https://pubmed.ncbi.nlm.nih.gov/PMC6163637/)].
12. Notomi T, Okayama H, Masubuchi H, Yonekawa T, Watanabe K, Amino N, et al. Loop-mediated isothermal amplification of DNA. *Nucleic Acids Res*. 2000;28(12). E63. doi: [10.1093/nar/28.12.e63](https://doi.org/10.1093/nar/28.12.e63). [PubMed: [10871386](https://pubmed.ncbi.nlm.nih.gov/10871386/)]. [PubMed Central: [PMC102748](https://pubmed.ncbi.nlm.nih.gov/PMC102748/)].
13. Mei X, Zhai X, Lei C, Ye X, Kang Z, Wu X, et al. Development and application of a visual loop-mediated isothermal amplification combined with lateral flow dipstick (LAMP-LFD) method for rapid detection of *Salmonella* strains in food samples. *Food Control*. 2019;104:9–19. doi: [10.1016/j.foodcont.2019.04.014](https://doi.org/10.1016/j.foodcont.2019.04.014).
14. Parida MM, Santhosh SR, Dash PK, Tripathi NK, Saxena P, Ambuj S, et al. Development and evaluation of reverse transcription-loop-mediated isothermal amplification assay for rapid and real-time detection of Japanese encephalitis virus. *J Clin Microbiol*. 2006;44(11):4172–8. doi: [10.1128/JCM.01487-06](https://doi.org/10.1128/JCM.01487-06). [PubMed: [17005741](https://pubmed.ncbi.nlm.nih.gov/17005741/)]. [PubMed Central: [PMC1698363](https://pubmed.ncbi.nlm.nih.gov/PMC1698363/)].
15. Zhang Y, Chen M, Liu C, Chen J, Luo X, Xue Y, et al. Sensitive and rapid on-site detection of SARS-CoV-2 using a gold nanoparticle-based high-throughput platform coupled with CRISPR/Cas12-assisted RT-LAMP. *Sens Actuators B Chem*. 2021;345:130411. doi: [10.1016/j.snb.2021.130411](https://doi.org/10.1016/j.snb.2021.130411). [PubMed: [34248284](https://pubmed.ncbi.nlm.nih.gov/34248284/)]. [PubMed Central: [PMC8257267](https://pubmed.ncbi.nlm.nih.gov/PMC8257267/)].
16. Itano T, Kawakami H, Kono T, Sakai M. Detection of fish nocardiosis by loop-mediated isothermal amplification. *J Appl Microbiol*. 2006;100(6):1381–7. doi: [10.1111/j.1365-2672.2006.02872.x](https://doi.org/10.1111/j.1365-2672.2006.02872.x). [PubMed: [16696687](https://pubmed.ncbi.nlm.nih.gov/16696687/)].
17. Yeh HY, Shoemaker CA, Klesius PH. Evaluation of a loop-mediated isothermal amplification method for rapid detection of channel catfish *Ictalurus punctatus* important bacterial pathogen *Edwardsiella ictaluri*. *J Microbiol Methods*. 2005;63(1):36–44. doi: [10.1016/j.mimet.2005.02.015](https://doi.org/10.1016/j.mimet.2005.02.015). [PubMed: [16157211](https://pubmed.ncbi.nlm.nih.gov/16157211/)].
18. Jalali H, Mozaffari Nejad AS, Ebadi AG, Laey G. Ethnobotany and folk pharmaceutical properties of major trees or shrubs in northeast of Iran. *Asian J Chem*. 2009;21(7):5632–8.
19. Kavyani B, Arabestani MR, Mozaffari Nejad AS, Moradkhani S, Kamarehei F, Taheri M, et al. The effect of Cinnamon bark on the expression of quorum sensing system and virulence genes in *pseudomonas aeruginosa* strain PAO1. *Res J Biotechnol*. 2019;14(2):42–8.
20. Mahmoudi H, Hossainpour H, Moradi M, Alikhani MY. Distribution of iron uptake systems encoding genes among the clinical isolates of *Escherichia coli* compared to foodstuffs isolates. *Infect Disord Drug Targets*. 2020;20(4):517–22. doi: [10.2174/1871526519666190119112542](https://doi.org/10.2174/1871526519666190119112542). [PubMed: [30659552](https://pubmed.ncbi.nlm.nih.gov/30659552/)].
21. Shariatifar N, Janghorban A, Rahimnia R, Mozaffari Nejad AS. Antimicrobial, antifungal and antioxidant activities of bee glue ethanol and aqueous extracts. *J Biol Res*. 2018;90(2). doi: [10.4081/jbr.2017.6452](https://doi.org/10.4081/jbr.2017.6452).
22. Mozaffari Nejad AS, Shabani S, Bayat M, Hosseini SE. Antibacterial effect of garlic aqueous extract on *Staphylococcus aureus* in hamburger. *Jundishapur J Microbiol*. 2014;7(11). e13134. doi: [10.5812/jjm.13134](https://doi.org/10.5812/jjm.13134). [PubMed: [25774277](https://pubmed.ncbi.nlm.nih.gov/25774277/)]. [PubMed Central: [PMC4332239](https://pubmed.ncbi.nlm.nih.gov/PMC4332239/)].
23. Tamariz J, Guevara V, Guerra H. Rapid detection of salmonellosis due to *Salmonella enterica* serovar Typhimurium in Peruvian commercially bred cavies, using indigenous wild bacteriophages. *Germs*. 2018;8(4):178–85. doi: [10.18683/germs.2018.1144](https://doi.org/10.18683/germs.2018.1144). [PubMed: [3075336](https://pubmed.ncbi.nlm.nih.gov/3075336/)]. [PubMed Central: [PMC6363402](https://pubmed.ncbi.nlm.nih.gov/PMC6363402/)].
24. Bai J, Trinetta V, Shi X, Noll LW, Magossi G, Zheng W, et al. A multiplex real-time PCR assay, based on *invA* and *pagC* genes, for the detection and quantification of *Salmonella enterica* from cattle lymph nodes. *J Microbiol Methods*. 2018;148:110–6. doi: [10.1016/j.mimet.2018.03.019](https://doi.org/10.1016/j.mimet.2018.03.019). [PubMed: [29621581](https://pubmed.ncbi.nlm.nih.gov/29621581/)].
25. Wan C, Yang Y, Xu H, Aguilar ZP, Liu C, Lai W, et al. Development of a propidium monoazide treatment combined with loop-mediated isothermal amplification (PMA-LAMP) assay for rapid detection of viable *Listeria monocytogenes*. *Int J Food Sci Technol*. 2012;47(11):2460–7. doi: [10.1111/j.1365-2621.2012.03123.x](https://doi.org/10.1111/j.1365-2621.2012.03123.x).